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SIMULATION STUDIES ON *DIGITARIA ERIANTHA* STEUD. SUBSP. *ERIANTHA* AT DIFFERING SOIL NITROGEN LEVELS.

 $\mathbb{B}\mathbb{Y}$

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A DISSERTATION SUBMITTED TO THE FACULTY OF U. O. V. S. IN FULFILMENT OF THE REQUIREMENT FOR THE DEGREE OF MASTER OF SCIENCE IN AGRICULTURE IN THE DEPARTMENT OF AGROMETEOROLOGY.

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DECLARATION

I declare this dissertation to be my own work. It has been submitted for the degree of Master of Science in Agriculture in the Department of Agrometeorology, University of the Orange Free State, Bloemfontein, South Africa. It has not been submitted for any degree in any other University or faculty.

M.D. HOWARD

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Note: Programme statements were expressed in the BASIC computer Language format.

TABLE OF CONTENTS

i

CHAPTER 1

INTRODUCTION	I .	1
1.1	Description of the problem	1
1.2	Objectives of the study	2
1.3	Modelling Approach	2
1.4	Description of the study area	3

CHAPTER 2

MODEL DESCRIPT	TION AND REFINEMENT	6
2.1	Choice of PUTU 13 as a basis	6
2.2	Model structure	7
2.3	Phenology	9
2.4	Leaf development	14
2.5	Dry matter assimilation	15
2.6	Carbohydrate translocation	29
2.7	Soil mass balances	35
	2.7.1 Water balance	35
	2.7.2 Nitrogen balance	41
2.8	Calibration of the PUTU 14 model for D. eriantha	50
	2.8.1 Method	50
	2.8.2 Results	50
· ·	2.8.3 Discussion	52

CHAPTER 3

VALIDATION OF THE PUTU 14 MODEL		THE PUTU 14 MODEL	57
	3.1	Introduction	57
	3.2	Results and discussion	57

CHAPTER 4 MODEL APPLICATIONS 65 4.1 Introduction 65 4.2 Interaction between climate and nitrogen application 67 4.3 Optimization of nitrogen application and climatic risk 70 CHAPTER 5 SUMMARY AND CONCLUSIONS 74

80

89

LITERATURE SURVEY

APPENDIX

ii

iii

LIST OF SYMBOLS

SOIL PARAMETERS

CLAY	Average clay content of the A and B horizon of the soil	(%)
DRAIN1	Water drainage from soil level one	(mm)
DRAIN2	Water drainage from rest of soil profile	(mm)
DRAINP	Drainage coefficient (0.9)	(mm d ⁻¹)
FC1	Field capacity of soil level one 100 mm	(%)
FC2	Field capacity of remainder of soil profile	(%)
GWP	Soil water potential	(kPa)
HYCON	Hydraulic conductivity (0.01)	(mm d ⁻¹ kPa ⁻¹)
INFIL	Infiltration of water into soil	(mm)
PWP1	Permanent wilting point of soil level one	(%)
PWP2	Permanent wilting point of remainder of soil profile	(%)
RUNPAR	Run-off coefficient (0.1)	(mm d ⁻¹)
SDP	Effective root depth of pasture	(mm)
SDP1	Soil depth of level one	(100 mm)
SDP2	Soil depth of level two, remainder of profile	(mm)
SLWAT1(1)	Soil water content for soil profile one (100 mm)	(mm)
SLWAT2(1)	Soil water content for remainder of soil profile	(mm)
SLWP1(1)	Soil water of level one (1 st 100 mm)	(%)
SLWP2(1)	Soil water of remainder of profile	(%)
WHC1	Water holding capacity at field capacity 10 Kpa	(%)
WHC4	Water holding capacity at permanent wilting point 2440 Kpa	(%)
WHCTEMP	Water holding capacity from previous season	(%)

ENVIRONMENTAL PARAMETERS

AE	Actual evaporation	(mm d ⁻¹)
AMXT	Maximum air temperature	(°C)
AMNT	Minimum air temperature	(°C)
ATEMP	Average daily air temperature	(°C)
EE	Equilibrium evaporation	(mm d ⁻¹)
EVAP	Daily evaporation	(mm)

ermal period required to trigger next growth stage	(°C)
Minimum value to which HUCRIT may be reduced by	
ter stress	(°C)
tential evaporation	(mm d ⁻¹)
ily precipitation	(mm)
diant flux density	(J m ⁻² d ⁻¹)
ily sunshine duration	(h)
l evaporation	(mm d ⁻¹)
	ermal period required to trigger next growth stage nimum value to which HUCRIT may be reduced by ter stress eential evaporation ily precipitation diant flux density ily sunshine duration l evaporation

PLANT PARAMETERS

AL	Leaf area index	
BCFUNC	Basal cover function	(%)
DBL	Leaf mass change demand at 100 % basal cover	(kg ha ⁻¹)
DMG	Daily dry matter gain	(kg ha ⁻¹)
GRSTGE	Growth stage	(1-5)
JDMAKS	Day on which maximum biomass is reached	(1-365)
LWP	Leaf water potential	(kPa)
PPRODO	Previous production	(kg ha ⁻¹)
PRVISC	Previous biomass production	(kg ha ⁻¹)
SPL	Specific leaf area	(kg ha ⁻¹)
TNEXT	Next growth stage	
TPMAKS	Maximum biomass production of previous season	(kg ha ⁻¹)
TRANS1	Daily transpiration from soil level one	(mm)
TRANS2	Daily transpiration from remainder of soil profile	(mm)
WPC	Wilting point capacity	(-1800 kPa)

PLANT COMPONENTS

BBL	Live leaf mass	(kg ha ⁻¹)
BBD	Dead leaf mass	(kg ha ⁻¹)
BCL	Live culm mass	(kg ha ⁻¹)
BCD	Dead culm mass	(kg ha ⁻¹)
BGL	Live grain mass	(kg ha ⁻¹)
BGD	Dead grain mass	(kg ha ⁻¹)

iv

BSL	Live stubble mass	(kg ha ⁻¹)
BSD	Dead stubble mass	(kg ha ⁻¹)
BRL	Live root mass	(kg ha ⁻¹)
BRD	Dead root mass	(kg ha ⁻¹)
BL	Live leaf mass	(kg ha ⁻¹)
BD	Dead leaf mass	(kg ha ⁻¹)
CL	Live culm mass	(kg ha ⁻¹)
CD	Dead culm mass	(kg ha ⁻¹)
GL	Live grain mass	(kg ha ⁻¹)
GD	Dead grain mass	(kg ha ⁻¹)
RL	Live root mass	(kg ha ⁻¹)
RD	Dead root mass	(kg ha ⁻¹)
SL	Live stubble mass	(kg ha ⁻¹)
SD	Dead stubble mass	(kg ha ⁻¹)
BTB	Total above ground leaf biomass	(kg ha'l)
BTG	Total above ground grain biomass	(kg ha ⁻¹)
BTC	Total above ground culm biomass	(kg ha ⁻¹)
BTRO	Total root biomass	(kg ha ⁻¹)
BTS	Total above ground stubble biomass	(kg ha ⁻¹)
RES	Reserve pool for carbohydrate	(kg ha⁻¹)

v

RATE PROCESS

DPROB	Desired proportion of DMG routed to leaf	(Partitioning factor)
DPROC	Desired proportion of DMG routed to culm	(Partitioning factor)
DPROG	Desired proportion of DMG routed to grain	(Partitioning factor)
DPROR	Desired proportion of DMG routed to root	(Partitioning factor)
DPROS	Desired proportion of DMG routed to stem	(Partitioning factor)
R(1)	Daily translocation rate to living leaves	(kg ha'l)
R(2)	Daily translocation rate to living stems	(kg ha' ¹)
R(3)	Daily translocation rate to living roots	(kg ha')
R(4)	Daily translocation rate to living culm	(kg ha ⁻¹)
R(5)	Daily translocation rate to living grain	(kg ha' ^ı)

X(1)	Translocation rate to dead stems	(kg ha ⁻¹)
X(2)	Translocation rate to dead leaves	(kg ha ⁻¹)
X(3)	Translocation rate to dead roots	(kg ha ⁻¹)
X(8)	Translocation rate to dead grain	(kg ha ⁻¹)
X(9)	Translocation rate to dead culm	(kg ha ⁻¹)
X(4)	Translocation rate from dead grain to trash	(kg ha ⁻¹)
X(5)	Translocation rate from dead culm to trash	(kg ha ⁻¹)
X(6)	Translocation rate from dead stems to trash	(kg ha ⁻¹)
X(7)	Translocation rate from dead leaves to trash	(kg ha ⁻¹)
ITERM	Period over which mean determined	(1)
CUT	Cutting date of vegetation	(CUT)

SOIL NITROGEN

AN	Available inorganic nitrogen in soil	(kg ha'')
ANC	Nitrogen content of soil water	(kg ha ⁻¹ mm ⁻¹)
CNR	C:N ratio	
CNRFOM	C:N ratio of fresh organic matter	
CNRHUM	C:N ratio of humus	
DECR	Decomposition rate of fresh organic matter	(d ⁻¹)
DMINR	Maximum rate of N-mineralization from humus	
DOYFERT	Day when fertilized	(DoyFert(i))
FCNR	C:N control function for decomposition	
FERTH4	Nitrogen in NH₄ form	(kg ha ⁻¹)
FERTO3	Nitrogen in NH ₃ form	(kg ha ⁻¹)
FOM	Fresh organic matter	(kg ha ⁻ⁱ)
HUM	Humus content of soil	(kg ha'')
IFOM	Fresh organic matter in soil after last application	(kg ha' ^ı)
L	Soil layer number	(1-2)
NFERT	Number of nitrogenous applications	
NFOM	Nitrogen content of fresh organic matter	(kg ha ⁻¹)
NH4	Ammonium content of soil	(kg ha' ^ı)
NHUM	Humus in soil	(kg ha ⁻¹)
NIMMFOM	Nitrogen immobilization from fresh organic matter	(kg ha ⁻¹ d ⁻¹)
NINFLO	Nitrogen inflow in soil	(kg ha ⁻¹ d ⁻¹)

vi

NIT	Nitrification rate	(kg ha ⁻¹ d ⁻¹)
NLEACH	Nitrogen leaching from soil	(kg ha ⁻¹ d ⁻¹)
NMASFLO	Nitrogen uptake as a result of mass flow	(kg ha ⁻¹ d ⁻¹)
NMINHUM	Nitrogen mineralization from humus	(kg ha ⁻¹ d ⁻¹)
NMINNET	Net nitrogen mineralization rate	(kg ha ⁻¹ d ⁻¹)
NO3	Nitrate content of soil	(kg ha ⁻¹)
NOUTFLO	Nitrate flow from a given soil layer	(kg ha ⁻¹ d ⁻¹)
PERC	Water flow from given soil layer	(mm d ⁻¹)
RDECR	Maximum fraction of organic matter decomposed	(kg ha ⁻¹ d ⁻¹)
TOTAN	Total available nitrogen in soil	(kg ha ⁻¹)

PLANT NITROGEN

COMP	Plant component (1 root, 2 culm, 3 leaf, 4 grain)	
FACN	Nitrogen control function for growth	
MASS	Plant component mass	(kg ha ⁻¹)
Ν	Nitrogen content of each plant component	(kg ha ⁻¹)
NC	Nitrogen concentration of each plant component	(kg kg ⁻¹)
NCMAX	Maximum nitrogen content of plant component	(kg kg ⁻¹)
NCMIN	Minimum nitrogen content of plant component	(kg kg ⁻¹)
NCOPT	Optimum nitrogen content of plant component	$(kg kg^{-1})$
NDEM	Nitrogen demand	(kg ha')
NDIFFLO	Nitrogen uptake by diffusion	(kg ha ⁻¹)
NDIFFLOO	Maximum nitrogen uptake by diffusion	(kg ha ⁻¹)
NUP	Nitrogen uptake	(kg ha ⁻¹ d ⁻¹)
NUPO	Maximum nitrogen uptake rate	(kg ha ⁻¹ d ⁻¹)
PAN	Plant available nitrogen	(kg ha ⁻¹)
TNDEM	Total plant nitrogen demand	(kg ha ⁻¹)
TPAN	Total plant nitrogen available for re-mobilization	(kg ha ⁻¹)

ENVIRONMENTAL FACTORS IN NITROGEN SUBROUTINE

FTMIN	Temperature control function for mineralization	
FWMIN	Water control function for mineralization	
Т	Average daily soil temperature	(°C)

vii

(mm m⁻¹) (mm)

W

V

INDEX OF	TABLES
----------	--------

Table 2.1	Models which simulate certain photo- and biochemical	
	events in various plant species.	16
Table 2.2	Senescence factors accounting for the daily transfer of mass	
	(kg ha ⁻¹ d ⁻¹) from living plant structures to dead tissue in the	
	different growth stages. (Source code statement numbers are	
	given in parenthesis).	. 33
Table 2.3	Trash factors for the transfer of dead tissue to trash, in the	
	different growth stages. (Source code reference are given in	
	parenthesis).	34
Table 2.4	Model performance during calibration (1978/79) on a	
	Westley $(n = 3)$ soil. MAE = mean absolute error, RMSE =	
	root mean square error, $RMSE_s = systematic error, RMSE_u$	
	= unsystematic error, \mathbb{D} = Wilmott index of agreement and	
	r^2 = coefficient of determination.	52
Table 2.5	Mean monthly rainfall (mm) for the Agriculture	
	Development Institute of Potchefstroom in relation to the	
	monthly mean for 1978/1979.	53
Table 2.6	New values of variables resulting from the calibration of	
	PUTU 14 on 1978/79 season data.	54
Table 3.1	Mean monthly rainfall (mm) for the Agriculture	
	Development Institute of Potchefstroom in relation to the	
	monthly long term mean for 1979/1980.	58
Table 3.2	Model performance and test for Westley $(n=3)$, Avalon	
	(n=6) and Valsrivier $(n=6)$ soils at various N-levels and	
	data from all three lumped together (n = 15). $MAE = mean$	
	absolute error, \mathbb{RMSE} = root mean square error, \mathbb{RMSE}_s =	
	systematic error, \mathbb{RMSE}_{u} = unsystematic error, \mathbb{D} =	
	Wilmott index of agreement and r^2 = coefficient of	
	determination.	61

ix

Monthly total rainfall (mm) with the given chances of non-Table 4.1 exceedance (cumulative probability) for Potchefstroom. 66 Years in which the monthly rainfall totals given in Table 4.1 Table 4.2 occurred at Potchefstroom. 67 **Rate of N-applications.** Table 4.3 67 Average, Median, Standard deviation, Low and High values Table 4.4 simulated for Potchefstroom from 1916-1991, at different Nlevels.

71

х

xi

INDEX OF FIGURES

Figure 1	Climatological characteristics of Potchefstroom with absolute	
	daily extremes (${\mathbb T}$) and average values reported (Anonymous,	
	1986; Koch, 1988).	4
Figure 2.1	Schematic representation of the model structure for the	
	PUTU family of models. The symbols are described in the	
	text.	7
Figure 2.2	The relationship between plant development rate and	
	prevailing temperature (after van Keulen, 1987).	10
Figure 2.3	The relation between photosynthesis and some limiting	
	factor as given by Devlin & Whitham (1983).	17
Figure 2.4	Typical relationship between the fraction of the maximum	
	translocation rate and the desired proportion of plant mass	
	shortage (Fouché <u>et</u> al 1986).	30
Figure 2.5	Partitioning factor for dry matter assimilate (kg ha ⁻¹ d ⁻¹) in	
	the different growth stages. For the status variables culm,	,
,	leaf, stubble, grain and root.	31
Figure 2.6	Components of the water balance model at a vegetative	
	surface.	35
Figure 2.7	Difference in calculated soil water potential between the	
	PUTU 11 and PUTU 13 and PUTU 14 simulation models.	39
Figure 2.8	Nitrogen balance in an ecosystem (Haynes, 1986)	42
Figure 2.9	Relationship between mean daily soil temperature and a	
	function describing how temperature limits mineralization	
	rate.	44
Figure 2.10	Relationship between soil water potential and the function	
	for describing how soil water limits the rate of	
	mineralization.	44
Figure 2.11	Relationship between C:N ratio and the function describing	
	how C:N ratio limits rate of decomposition of organic	
	matter.	44

Figure 2.12	Simulated nitrogen applied, simulated N-uptake, simulated	
	production and measured final yield at three N-levels for the	
	1978/79 growing season at Potchefstroom.	51
Figure 3.1	Comparison of measured and simulated yields for the	
	1978/80 and 1979/80 growth seasons.	58
Figure 3.2	Simulated nitrogen applied, simulated N-uptake, and	
	simulated production and measured final yield at three N-	
	levels for the 1979/80 growing season at Potchefstroom.	60
Figure 3.3	Comparison of measured and simulated yield attained for a	
	Westley, Avalon and Valsrivier soil.	62
Figure 3.4	Measured and simulated yield for Avalon soil at different	
	soil N-levels for 1981/82, 1982/83 and 1983/84 season.	63
Figure 3.5	Measured and simulated yields on a Valsrivier soil at	
	different soil N-levels for 1981/82, 1982/83 and 1983/84	
	season.	63
Figure 4.1	Variation in annual rainfall for period 1916 to 1990 at	
	Potchefstroom.	65
Figure 4.2	Influence of various climatic regimes and N-application rate	,
	on yield. The bars indicate the range in yield obtained as a	
	result of different time of application.	68
Figure 4.3	Influence of various climatic regimes and time of N-	
	application on yield. The bars indicate the range in yield	
	obtained as a result of the different levels of applied	
	nitrogen.	69
Figure 4.4	Climatic risk — the cumulative distribution function of	
	simulated yield for different soil N-levels.	71
Figure 4.5	The expected outcome-variance space - A. Plot of mean yield	
	against standard deviation at differing N-applications. A 2:1	
	line is also indicated. Data for all 76 seasons was included.	72
Figure 4.6	Chance of realising an increased yield were a higher N-	
	application utilized (\vartriangle) and the mean yield (*) at given	
	nitrogen application rates. The bars indicate the range	
	(maximum - minimum) in yields due to seasonal weather	
	variability.	73

xiii

INDEX OF APPENDICES

Appendix A	Source code for the PUTU 14 model (Digitaria	
	eriantha).	89
Appendix B	Source code for the soil nitrogen balance used in the	
	PUTU 14 model.	104
Appendix C	Source code for the plant nitrogen balance used in	
	the PUTU 14 model.	106
Appendix D.1	Table of high, low and average yield values (t ha ⁻¹)	
	simulated for a bad year.	108
Appendix D.2	Table of high, low and average yield values (t ha ⁻¹)	
	simulated for a poor year.	108
Appendix D.3	Table of high, low and average yield values (t ha ⁻¹)	
	simulated for a moderate year.	108
Appendix D.4	Table of high, low and average yield values (t ha-1)	
	simulated for a good year.	109
Appendix D.5	Table of high, low and average yield values (t ha ⁻¹)	,
	simulated for a wet year.	109
Appendix D.6	Simulated seasonal yield kg ha ⁻¹ from 1916-1991 for	,
	different N-levels.	109

PREFACE

All the glory to our God almighty who reigns forever, Amen.

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May God's richest blessing be bestrewed upon you.

CHAPTER 1

INTRODUCTION

1.1 Description of the problem

Growth modelling entails multi-disciplinary investigations concerning:

- collection and classification of state-of-the-art knowledge,
- mathematical collation of such knowledge, and
- verification and validation of the algorithms produced from such information.

Algorithms are used to build a growth model and reflect the extent of present knowledge. Skoog (1955) concluded, "We can claim to understand the plant when we can express it all in a mathematical model". Since then, much literature has been published on the subject. Of particular relevance here, are the reviews by Van Veen & Frissel (1981), Van Keulen (1982), Van Keulen, Penning de Vries & Drees (1982). Booysen (1983), Fouché (1984), Van Keulen & Seligman, (1987) Du Pisani (1992) and Fouché (1992). Mostly, models have been developed for agronomic species. Southern African, grassland scientists have only really commenced work in this field over the past decade (De Jager, 1976; De Jager, Opperman & Booysen, 1980; Booysen, 1983; Fouché, 1984; Du Pisani, 1992 and Fouché, 1992).

Of particular interest is the PUTU 13 model used by Du Pisani (1992) to demonstrate the practical value of a grassland model in determining animal/veld performance under different stocking rates. This model was adapted from the PUTU 11 (Booysen, 1983; Fouché, 1984 and Fouché, 1992) which was based upon the original PUTU 2 climax grassland model developed by De Jager (unpublished monograph, 1976).

Du Pisani (1992) further developed the PUTU 13 model to make possible the simulation of daily dry matter production of *Cenchrus ciliaris* L. cv. Molopo. Inability to simulate the influence of nitrogen on dry matter production however restricted the use of this model. PUTU 13 however did represent the first deterministic model developed for a 2

subtropical dryland pasture and could well prove to be the basis for developing new models for other dryland cultivated pastures, e.g. Anthephora pubescens, Digitaria eriantha, Eragrostis curvula, Medicago sativa and Sorghum almum.

Numerous key questions arose following the work of Du Pisani (1992) viz. is it possible to :

- 1. use the PUTU 13 model as a basis for developing deterministic models for other dryland cultivated pasture such as *D. eriantha* ?
- 2. include a mechanistic computational procedure into the PUTU 13 model, which will enable simulation of the impact of nitrogen upon daily dry matter production ?
- 3. apply such model to solving questions as to how much and when to apply nitrogen ?
- 1.2 Objectives of the study

With these questions in mind, it was decided to:

- adapt PUTU 13 to simulate daily dry matter production of *D. eriantha*.
- build therein a routine accounting for the influence of nitrogen upon growth.
- validate the new model under different climatic and soil conditions, and
- undertake preliminary demonstration of the new model's suitability for determining nitrogen application strategies for different climatic scenarios.

1.3 Modelling approach

Throughout the thesis reference will be made to the scientific literature in the relevant sections as this becomes necessary. No separate literature study has been undertaken. Mathematical approaches towards crop growth modelling could involve the use of linear regression, stochastical or statistical techniques, or mechanistic procedures. Furthermore, plant growth modelling could take place at different levels (resolution) e.g. whole

crop, plant organs, tissues, cells etc. (see Thornley & Johnson, 1990). The complexity of the modelling will depend on :

- the current state of knowledge,
- what data are available to compose and validate the model, and
- the accuracy required of the model output.

In this study, an intermediate level of modelling complexity was adopted. This entailed simplification of the PUTU 11 model and required the quantification of :

- growth and development in the plant organs i.e. roots, leaves, culm, stubble and seed, and
- the influence of daily status of radiation, temperature, precipitation and nitrogen on plant growth and development.

The model output was expected to be sensitive to differences in N-application rates of 20 kg N ha⁻¹. For modelling purposes, the study was restricted to use of available field data, which unfortunately were relatively few. The study thus represents a first attempt, but as the results will show, one offers much potential for future refinement and practical application.

1.4 Description of the study area

Data used to calibrate and validate the PUTU 13 model for *D. eriantha* were obtained for Pochefstroom from the High Veld Region Agriculture Development Institute. The experimental station is situated at 26° 44' latitude and 27° 05' longitude, 1345 m above sea-level. The data were collected by Dannhauser (1985), Dannhauser, Van Rensburg, Opperman & Van Rooyen (1987) and Dannhauser (1988). In general the temperature of Potchefstroom reflects cold to moderate winters with warm to hot summers (Figure 1.A & 1.B). The location receives a summer rainfall (Figure 1.C). with a long term mean precipitation of 625 mm per annum. The average daily sunshine duration varies between 8 and 10 hours (Figure 1.D) (Anonymous, 1986; Koch, 1988).



Figure 1 Climatological characteristics of Potchefstroom with absolute daily extremes (1) and average values reported (Anonymous, 1986; Koch, 1988).

1.4.2 Soil

The soil profile can be characterized as a Westley form with a clay content of 25 %, soil depth of A & B horizon is 500 mm. This was covered by *D. eriantha* with an effective rooting depth of 300 mm. Results of soil analyses showed that the soil contained:

 $pH(H_2O) = 6.4$; pH(KCI) = 5.3; P = 35 and K = 62 ppm, respectively (Dannhauser, 1985).

CHAPTER 2

MODEL DESCRIPTION AND REFINEMENT

2.1 Choice of PUTU 13 as a basis

Certain factors were taken into account when considering models for possible refinement for use on cultivated pastures :

- the accessibility of source code and required level of complexity,
- the ease with which the new model may be interfaced with an animal production model such as STEER (Du Pisani, 1992), and
- compatibility of the fundamental structure of the basic model with the growth characteristics of a cultivated pasture such as *D. eriantha*.

The PUTU 13 model was selected as a basis for refinement due to the fact that both *C*. *ciliaris* and *D. eriantha* are all classified as subtropical dryland species and PUTU 13 had demonstrated reliability in the semi-arid regions of the Orange Free state. Fouché (1992) found with PUTU 11 for climax grassland that $R^2 = 0.92$ for total dry matter production simulated versus measured over 12 seasons. Furthermore, du Pisani (1992) found over 5 seasons that $R^2 = 0.96$ between measured and simulated yield for *C*. *ciliaris* (PUTU 13) and $R^2 = 0.86$ where measured animal mass was correlated against animal mass simulated using the STEER and PUTU 13 models.

PUTU 13 has evolved as the latest in a series of crop growth models initiated in 1973 (see De Jager, 1974). Fouché (1992) chronicals the history of the development of the PUTU models.

In view of these facts, a new model PUTU 14 for *D. eriantha* was developed by appropriately simplifying and modifying PUTU 11 and 13.

2.2 Model Structure

Fouché, De Jager & Booysen (1986) gave an overview of the typical structure of the PUTU models (see figure 2.1 for a schematic representation of the structure). A brief description of the PUTU 13 model will now be given. The source code for PUTU 14 is given in Appendix A. Where specific code statements are referred to the line numbers are given in parenthesis. A detailed description of the model is provided, which includes a relevant literature survey. This was done because no updated description of PUTU 11 and PUTU 13 exist and the work here reported represent a marked simplification of these models. In these models daily CO_2 assimilation is computed from weather variables and converted to dry matter (DMG) using the conversion factor 0.66 kg $CH_2O/kg CO_2$.



Figure 2.1 Schematic representation of the model structure for the PUTU family of models. The symbols are described in the text.

In Fig 2.1 the following symbols apply. Daily dry matter gain (DMG) from the assimilation process is routed to the reserve pool of carbohydrates (RES) (see [A, 419-423,434]. From this it is distributed among the different plant organs. Translocation rates of mobile carbohydrates to the living plant organs; grain (GL), culm (CL), leaf (BL), stubble (SL) and root (RL) are quantified by parameters R5, R4, R1, R2 and R3, respectively. Due to senescence, biomass transfers from live to dead tissue. These translocation rates are accounted for by factors X8, X9, X2, X1 and X3 for grain (GD), culm (CD), leaf (BD), stubble (SD) and root (RD), respectively. With grassland the partitioning of dry matter is more important than is the case for most other species, because the storage of reserves can occur.

The amount of trash (TRD) is accumulated from each plant organ which dies at a rate determined by X4, X5, X7 and X6, respectively.

Parameters CA, CB, BA, DA, BB and DB account for total above ground standing crop, below ground standing crop, above ground biomass, above ground dead biomass, root biomass and root dead biomass, respectively.

In PUTU 11 and 13, in order to make accounting for mutual shading and competition due to basal cover possible; an input parameter BCOVER = 10 (percent), is introduced, see [A,30 and 50]. This is then used to describe the influence of basal cover on competition (BCFUNC). Future work will hopefully formulate the relationship (BCFUNC) between competition effects and BCOVER which should not be a linear one. Up to the present however, the simple assumption BCFUNC = BCOVER / 100, see [A,50], was made. For convenience all computations were executed at 100 % basal cover and production status in each component simply obtained from the product of biomass and BCFUNC. Where cultivated pastures are concerned, a homogeneous nearly constant distribution is produced at the time of sowing. It was therefore decided to remove this complicated procedure in PUTU 14. Since a summer crop is simulated in daily iterations, simulations commence on 1 July and end on 30 June of the following year. All the weather files are constructed accordingly and the day counter is J.

The various facets of the model will now be described.

2.3 Phenology

Seasonal plant development is related to physiological age and morphological appearance (Penning de Vries & Van Laar, 1982; Rimmington & Charles-Edwards, 1987; Thornley & Johnson, 1990). The prediction of phenological events in plants has been attempted as early as 1735 by Réamur as quoted by Van Keulen (1987).

Innes (1978) identified seven phenological events related to plant age. The PUTU family of models is capable of dividing the season into as many as nine different stages. The grassland models consider plant development in five categories, viz.

vegetative; reproductive; seed growth; seed fall and dormancy

Plant development is genetically determined, but regulated by external factors such as temperature and day length (Daubenmire, 1962; Innis, 1978; Angus, Mackenzie, Morton & Shafer, 1981; Penning de Vries & Van Laar, 1982; Van Keulen, 1987; Thornley & Johnson, 1990).

Plant development rate, is generally controlled by what is termed thermal period. These heat (temperature) sum formulae are commonly used to describe change from one phenological event to the next. Rimmington & Charles-Edwards (1987) described such process using :

W7 /4

$$v = Vr(1 + K\Delta I)$$
 (2.1)
where:

[2 1]

v = rate of phenological development (d⁻¹)
 Vr = development rate at a given reference temperature (d⁻¹)
 K = temperature coefficient (⁰C⁻¹)
 △T = difference between actual temperature experienced by the plant and the reference temperature (⁰C)

9

Where rate of development $v(d^{-1})$ is defined as the fraction of the present growth stage through which the vegetation progresses during a given day. An example of the relationship between temperature and rate of development described by Eq. 2.1 is shown in figure 2.2



Figure 2.2 The relationship between plant development rate and prevailing temperature (after Van Keulen, 1987).

Eq. 2.1 may be rewritten in the form of the general day/degree expression, viz.

$$\mathbf{t}_{AB} = \mathbf{a}_{D} + \mathbf{b} \sum \Delta \mathbf{t}$$
 [2.2]

where, the summation takes place in daily increments. Furthermore t_{AB} is the time taken for the vegetation to progress between stages A and B and a_n and b are crop physiological constants. It is evident that t_{AB} is linearly related to the sum of the daily differences between the daily mean temperature experienced by the plant and the reference temperature (Charles-Edwards, Dooley & Rimmington, 1986).

The term $\sum \Delta t(d^{\circ}C)$ may simply be re-defined as the heat summation above a base temperature (at which development is zero) and then HU, or heat units, symbolizes what is termed thermal period.

The PUTU simulation models step from one growth stage to the next, when a minimum heat summation HUCRIT accrues to the plant:

Thermal period is computed by daily incrementation of an algorithm derived from Eq. 2.2, viz.

[A,127]	HU	$= HU + (TGROW_{d} - BO)$	[2.3]
	HU	= thermal period (d°C)	
[A,124,325] where:	TGROW _d	= daily air temperature effective in the development	
	-	process (°C)	
[A,32]	BO	= base air temperature below which no growth occurs	
		$(BO = 10 \ ^{\circ}C)$	

In order to simulate the decreased plant development at extreme temperatures, Eq. 2.3 is subjected to two constraints, viz:

[A,126]	TGROW _d	= BO for (AMXT + AMNT)/2 \leq BO, or
[A,125]	TGROW _d	= 30 for $(AMXT + AMNT)/2 > 30$.

AMXT and AMNT are daily maximum and minimum air temperature, respectively. When the thermal period, HU, in a given growth stage exceeds the HUCRIT corresponding to that growth stage, the model triggers progression to the next growth stage.

2.3.1 Growth stage one - Vegetative growth

No growth is assumed possible before photoperiod 31 st July [A,119]. The vegetation steps from vegetative to reproductive growth when it has received a thermal period HU HUCRIT [A, 120]. The HUCRIT used in PUTU 11 for the vegetative growth stage is 250 (d°C) based upon a BO of 12 °C. Furthermore, PUTU 11 & 13 with BO = 12 °C, differs from PUTU 14 which has BO = 10 °C. The effect of drought on phenology is accounted for by shortening the duration (the thermal period required) in growth stages which experience drought. The critical thermal period is shortened according to :

[A,128] HUCRIT = HUCRIT - 10 * (100 - FW) / 100 [2.4]

FW is a growth limiting factor determined by plant water status. It will be described later. However HUCRIT may not become smaller than HUCRMN. When this occurs an assumption is made and HUCRIT is set equal to a minimum value HUCRMN (Appendix A, 129). For PUTU 13 and 14 the values of HUCRMN are 225 (d°C) and 230 (d°C), respectively.

Furthermore, when minimum daily minimum air temperature drops below 2 °C after 15 March, then reproductive growth commences :

[A,121]	IF	J	> 258 then 18	
[A,122] 18	IF	AMNT	\leq 2 then terminate growth stage one	[2.5]
where:		AMNT	= minimum temperature (°C)	

2.3.2 Growth stage two - Reproductive growth

Triggers for the change from reproductive to seed growth are one of the following;

25 days of reproductive growth for *Themeda-Cymbopogon* veld type (Fouché, 1992) 60 days of growth for *C. ciliaris* L. cv. Molopo (Du Pisani, 1992), or 30 days of growth for *D. eriantha*, and culm production must exceed 25 % of leaf biomass;

[A,148]		CL	= $(.25 * BL) AND J > MKOUNT$	[2.6]
v	where:	CL	= Live culm biomass (kg ha ⁻¹)	
[A,424]		BL	= Live leaf biomass (kg ha ⁻¹)	
•		J	= days since July 1 st	
[A,146]		MKOUNT = J + 30		

Reproductive growth may not endure for longer than 316 days of growth (i.e. beyond 15 May) [A,149] for C. ciliaris L. cv. Molopo (Du Pisani, 1992) and D.eriantha, or 258 growth days (15 March) for Themeda-Cymbopogon veld type (Fouché, 1992) and the daily minimum temperature drops below 2°C.

[A,149] $J \ge 316$

2.3.3 Growth stage three - Seed formation

Triggers for the change to seed fall growth phase are :

[A,164]
 1. 80 days (MKOUNT = 80) of seed formation for *D. eriantha*. It was 100 and 50 days for *C. ciliaris* L. cv. Molopo (Du Pisani, 1992) and *Themeda-Cymbopogon* veld type (Fouché, 1992), respectively, or

[A,168] 2. the growing season exceeds day J = 316 and

 [A,170]
 3. a minimum air temperature of 3, 2 and 2 °C is reached for *Themeda-Cymbopogon* veld type (Fouché, 1992) and C. ciliaris L. cv. Molopo (Du Pisani, 1992) and D.eriantha, respectively.

2.3.4 Growth stage four - Seed fall

Dormancy will commence when a minimum temperature of 3, 2 and 2 °C is reached for *Themeda-Cymbopogon* veld type (Fouché, 1992) and *C. ciliaris* L. cv. Molopo (Du Pisani, 1992), and *D. eriantha*, respectively [A,189].

2.3.5 General

The above approach is used to simulate phenological development in the PUTU grassland models. It is simple and makes possible describing different rates of change for each specie, provided its phenological characteristics are known. The differences between *D. eriantha*, *C. ciliaris* and *Themeda-Cymbopogon* veld type have been highlighted.

From an agricultural/ecological point of view the daily temperature extremes are most important. Particularly a minimum daily temperature of around 2 °C seems to be critical in grassland simulation models.

2.4 Leaf development

Leaf growth takes place during the vegetative growth phase. Environmental factors, such as temperature and water, control the rate of leaf development. Squire (1990) described a function where rate of leaf change increases linearly with temperature up to an optimum. This theory, as applied in the PUTU grassland models, has been extended (actually simplified) to cover cultivated pastures.

Squire (1990) expressed potential (no water or other stress) change in leaf (length) per unit time $(\delta l/\delta t)$ as the product of the coefficient of thermal rate of expansion, \mathbb{P}_{ℓ} , (units mm (0 Cd)⁻¹) and the difference between the temperature (T) and the base temperature (T_{br}), thus:

$$\delta l/\delta t = P_{\ell} (T-T_{br})$$

This equation has been adapted to calculate the daily biomass increment per unit ground area per unit time (DBL) written in the form :

[A,130]	$DBL = BCON*(TGROW_d - BO) * FW/100 $ [2.7]
where:	DBL = daily increment in leaf biomass (kg ha ⁻¹ d ⁻¹)
[A,123]	BCON = leaf biomass increment per unit ground area per unit time
	per unit temperature increment above a basal
	value (3.5 kg ha ⁻¹ d ⁻¹ °C ⁻¹)
[A,124,325]	$TGROW_d$ = daily air temperature effective in the leaf development
	process (°C)
[A,32]	BO = base air temperature below which no growth occurs
	$(BO = 10 \ ^{\circ}C)$
[A,83,85,357]	FW = growth limiting factor due to plant water status

A description of how FW is computed will be given in Chapter 2.7.1. Leaf growth is constrained to a range of 10 °C and 30 °C (see A,125 and 126). Optimum growth rate will occur when TGROW_d reaches 30 °C. If the value of TGROW_d exceeds 30 °C then TGROW_d is set equal to 30 °C, simulating optimal growth rate at temperatures higher than 30 °C. Furthermore, no leaf growth will occur when the temperature drops below 12 °C or 10 °C for *C.ciliaris* and *D.eriantha*, respectively (see Table 2.6). This leaf

growth function could be modified in the future as additional information becomes available.

Furthermore, in PUTU 11 leaf development rate was initially taken as 20 % of the optimum rate until a leaf biomass of 2900 kg leaves ha⁻¹ at 100% basal cover was reached (see, Fouché, 1992). For PUTU 13 and 14 this restriction was removed and the purely linear relationship Eq 2.7 adopted throughout the leaf development (vegetative) stage.

2.5 Dry matter assimilation

Photosynthesis is the process by which absorbed radiant energy is utilized by plant material to transform chemical energy. Essentially such process embodies diffusion of carbon dioxide (CO₂), water uptake (H₂O) and the release of oxygen (O₂). Feddes, Kowalik & Zaradny (1978) describe this as follows :

	-		light	t						
$2H_2O$	+	CO ₂		······································	->	O ₂	+	(CH ₂ O)	+ H ₂ O	[2.8]
(from soil)	(fro	m atmo	osphere)		(to at	mosp	here)	(to plant)	(to atmos	sphere)

The processes involved in the formation of carbohydrates are enzymatic, photochemical and biochemical i.e. carbon dioxide-assimilation, photo-respiration and carbohydratemetabolism (Banner & Varner, 1965; Larcher, 1980; Paleg & Aspinall, 1981; Devlin & Whitham, 1983; Black & Vines, 1987; Jensen & Seftor, 1987; and Robinson, 1987). Carbohydrates act as an energy substrate in the plant and serve as building-blocks for newly formed tissues.

High resolution models have been developed to simulate assimilation and plant growth over the past two decades (see Table 2.1)

Model name	Author	Crop
SORGF	Arkin, Vanderlip & Ritchie (1976)	Sorghum
SIMED	Shreiber, Miles, Holt & Bula (1978)	Grass
CANPAS	Fick (1980)	Rye grass
GOSSUM	Mckinion & Baker (1982)	Cotton
BACROS/PHOTON	Penning de Vries & Van Laar (1982)	Rhodes grass
SUCROS	Van Keulen <u>et al</u> (1982)	Wheat
PUTU	De Jager (1992)	Maize, Wheat Grassland
CERES-MAIZE	Jones & Kiniry (1986)	Maize

Table 2.1Models which simulate certain photo- and biochemical events in various
plant species.

It is evident from Eq 2.8 that the rate of photosynthesis is determined by environmental factors such as: light, temperature, water, carbon dioxide and oxygen (Devlin & Whitham, 1983).

As early as 1860 Blackman, as quoted by Devlin & Whitham (1983) attempted to describe the control of photosynthetic rate by these factors. A typical control function is illustrated in Fig 2.3

In the PUTU grassland models, control of photosynthesis by the environmental elements is based upon the initial research of De Jager (1968). Here individual leaf gross photosynthetic rate was described in terms of incident radiant flux density, using a rectangular hyperbola given that the other environmental variables are not limiting photosynthesis i.e. they are at optimal value.



Figure 2.3 The relation between photosynthesis and some limiting factor as given by Devlin & Whitham (1983).

However, when temperature and plant water status were sub-optimal, they restricted growth, similarly to the relationship described by Fig 2.3. The original De Jager (1971a) results were eventually applied in a form where, F_w , the growth limiting factor due to plant water status, was described by a logistic function [A, 357] and, F_t , the growth limiting factor corresponding to temperature by a Gaussian function (De Jager, 1971b). Net photosynthetic rate was then expressed in terms of the product of a maximum radiation conversion coefficient and a light limiting factor derived from the rectangular hyperbola.

De Jager (1968) made the assumption that growth limiting factors could be mathematically combined in an overall growth limiting factor, F, using a law of independent mutual limitation (i.e. a multiplicative law), thus

$$\mathbf{F} = \boldsymbol{\pi}_{\mathrm{i}} \mathbf{F}_{\mathrm{i}}$$
 [2.9]

Where i denotes over the range of relevant environmental limiting factors, viz radiation, temperature and plant water status. Thus Eq. 2.9 may be written:

$$\mathbf{F} = \mathbf{F}_{q} \cdot \mathbf{F}_{t} \cdot \mathbf{F}_{w}$$
 [2.10]

Where \mathbb{F}_q , \mathbb{F}_t and \mathbb{F}_w are the growth limiting factor due to radiation, temperature and water, respectively. Thus, the net photosynthetic rate, \mathbb{P}_1 of individual leaves was expressed as

$$\mathbf{P}_{1} = \mathbf{RCC}.\mathbf{F}$$
 [2.11]

or
$$P_1 = RCC.F_q.F_t.F_w.$$
 [2.12]

Values for the parameters used in the functions \mathbb{F}_q , \mathbb{F}_t and \mathbb{F}_w are reported by De Jager (1971a) and De Jager (1971b) for Lolium multiflorum, Lolium perenne and Paspalum dilatatum.

Individual leaf photosynthesis

Notation :

The following symbol description will apply.

Variables:	
RCC	= radiation-photosynthesis conversion coefficient ($\mu g J^{-1}$)
Р	= rate of net photosynthesis ($\mu g(CH_2O) m^{-2} J^{-1}$)
Q	= vertical component of radiant flux density ($W m^{-2}$)
α	= leaf quantum yield, or photochemical efficiency
	4 (μgJ(CH ₂ O) J ⁻¹)
E	= efficiency
а	= absorbtivity
Subscripts:	
S	= level at radiation saturation
0	= maximum, or potential value
q	= radiation
e	= an individual leaf
v	= a vegetative canopy

De Jager (1974) derived a simple leaf gross photosynthesis model based upon the following principles :

$$P_{\ell} = \alpha a_{\ell} Q$$

$$Q \rightarrow 0$$

$$P_{\ell} = P_{ls}$$

$$Q \rightarrow \infty$$

These may be combined simply to form a rectangular hyperbola as follows

 $1/P_{e} = 1/ \alpha a_{e}Q + 1/P_{ls}$ thus $P_{e} = P_{ls}Q/(P_{ls}/\alpha a_{e} + Q)$ or $P_{e} = P_{ls}Q/(Q_{B} + Q) \qquad [2.13]$ where $Q_{B} = P_{es}/ \alpha a_{e} \qquad [2.14]$

Here, physically, Q_B is simply that radiant flux density at which one-half saturation leaf photosynthesis occurs. For convenient modelling purposes (as few as possible parameters); it is required to express photosynthetic rate as the product of a radiation conversion coefficient and incoming radiant flux density;

$$\mathbf{P}_{\ell} = \mathbf{RCC.Q}$$
 [2.15]

Now RCC may be evaluated in terms of the product of leaf quantum yield (α) and ϵ_q , a radiation use efficiency, which quantifies the efficiency with which incident radiation is used to fix CO₂. Thus, ϵ_q is defined as the fraction of incident radiation actually utilized in the photosynthetic process. Thus

RCC =
$$\alpha \epsilon_{q}$$
 [2.16]

The radiation use efficiency ϵ_q may be derived from Eq 2.13 as follows. By definition, the absolute potential (maximum) leaf gross photosynthetic rate at a given radiant flux density, \mathbb{P}_{ℓ_0} , will be given by :

$$\mathbf{P}_{\ell 0} = \alpha \mathbf{a}_{\ell} \mathbf{Q}$$
 [2.17]
Radiation use efficiency for individual leaves during the photosynthetic process is by definition

$$\epsilon_{q} = P_{\ell}/P_{\ell_{0}}$$
 [2.18]

Now substituting from Eq 2.13 and Eq 2.17

$$\epsilon_{q} = P_{ls}Q/(Q_{B}+Q) / \alpha a_{\ell}Q$$

= $Q_{B}/(Q_{B}+Q)$ [2.19]

 P_e may therefore be computed using Eq 2.15 and substituting for RCC estimated from Eq 2.16 and Eq 2.19. Thus

RCC =
$$\alpha \epsilon_{q}$$

= $\alpha Q_{B}/(Q_{B}+Q)$ [2.20]

A worked example is useful for illustrating the fundamentals. From Eq 2.14

$$Q_B = P_{ls}/\alpha a_\ell$$

For *Paspalum dilitatum* and *Lolium multiflorum* the values for $P_{\ell s}$ are 2400 μg m⁻² s⁻¹ and 360 μg m⁻² s⁻¹, respectively.

Thus for Lolium multiflorum

$$Q_{B} = 360 \ \mu g \ m^{-2} s^{-1} (CH_{2}O)/(4 \ \mu g \ J^{-1} \ 0.6)$$

= 70 W m⁻²
Note: 1 \mu \ell(CO_{2}) m^{-2} s^{-1} = 1.96 \ \mu \ 10^{-9} \ \mu g \ (CO_{2}) m^{-2} s^{-1}
= 1.96 \ \mu g \ (CO_{2}) m^{-2} s^{-1}

Leaf quantum yield, or α , is the initial slope of the radiant response curve for leaf net photosynthesis. For light energy it is around 12 μ g(CO₂)J⁻¹ for most plant types (see Charles-Edwards <u>et al</u> 1986). In terms of dry matter, this is equivalent to 8μ g(CH₂O)J⁻¹. Note that 44 μ g(CO₂) will produce 33 g(CH₂O) (0.68 g(CO₂) g(CH₂O)⁻¹), and when solar

Canopy photosynthesis

A similar theory can be derived for a crop canopy by modifying Eq 2.15 and 2.16 for leaf area index

For a vegetative canopy using the same symbol definition:

$$P_v = RCC Q F$$

where, now F = F_v. F_t. F_w

 \mathbb{F}_{v} the fractional radiation interception, vegetative canopy is defined as i.e. the fraction of incident radiant flux density intercepted by foliage.

For a vegetative canopy, evaluation of RCC is complicated by having to integrate ϵ_q over canopy leaf area and the full range of incident radiant flux density. There are many examples of how this is done in the literature (Duncan, Loomis, Williams & Hanan; 1967). Thus, the photosynthetic rate of an element of leaf area, subjected a radiant flux density Q_i is given by

$$P_{\ell} = RCC_{\ell} Q_{\ell i}$$

 $_{e}$ signifies finite element of leaf area, $_{i}$ signifies the class interval of radiant flux density to which the leaf is subjected.

The photosynthesis rate of an entire canopy, leaf area per unit square area is given by

$$\int_{0}^{I} = \int_{0}^{I} RCC_{t} Q_{t}$$

$$\int_{0}^{I} RCC_{t} = \alpha \int_{0}^{t} \epsilon_{q_{t}}$$

$$\epsilon_{q\ell} = P_{\ell s} / P_{\ell}$$

$$= P_{\ell s} Q_{\ell i} / (P_{\ell s} / \alpha a_{\ell} + Q_{\ell i}) / \alpha a_{\ell} Q_{\ell i}$$

$$= (P_{\ell s} / \alpha a_{\ell}) / (P_{\ell s} / \alpha a_{\ell} + Q_{\ell i})$$

$$= Q_{B} / (Q_{B} + Q_{\ell i})$$

More conveniently, an equation for canopy photosynthesis may be developed analogously to the method used for an individual leaf. This is subject to assuming the canopy to be a single entity with overall definable properties. Such assumption has been called the conglomerate hypothesis (De Jager, 1974), or big leaf theory (see Monteith, 1975). In analogy with the development of Eq 2.13, the limits of vegetative canopy (with leaf area index = L and fractional interception F_{y}) gross photosynthesis, P_{y} , may be expressed

	$\mathbf{P}_{\boldsymbol{\ell}}$ $\mathbf{Q} \to 0$	$= \alpha \mathbf{a}_{\ell} \mathbf{F}_{\mathbf{v}} \mathbf{Q}$
	P _v Q→∞	$= LP_{ls} = P_{vs}$
Yielding	$\mathbf{P}_{\mathbf{v}}$	$= \mathbf{P}_{vs} \mathbf{Q} / (\mathbf{Q}_{Bv} + \mathbf{Q})$
where, now,	Q_{Bv}	$= \mathbf{P}_{vs} / \alpha \mathbf{a}_{\ell} \mathbf{F}_{v}$

Once again, similarly to Eq 2.19, it may be shown that

	ϵ_{q}	$= Q_{Bv} / (Q_{Bv} + Q)$	[2.21]
and	$\mathbf{P}_{\mathbf{v}}$	= RCC Q	[2.22]
where	RCC	$= \alpha \epsilon_{q}$	

In the early PUTU models Eq 2.22 was expressed

$$P_{v} = RCC Q$$
$$= \alpha \epsilon_{0} Q$$

and the effect for plant water status and temperature (see Eq 2.12) accounted for using environmental growth limiting factors. Thus, by analogy, P_y may be expressed

$$P_{v} = \alpha [Q_{Bv} / (Q_{Bv} + Q)]. F_{v}. F_{t}. F_{w}. Q$$
$$= \alpha \epsilon_{q} F_{v}. F_{t}. F_{w}. Q$$
$$= \alpha \epsilon_{q} F. Q$$
$$F = \pi_{t} F_{t}.$$

with

The coding of which (see Fouché, 1992) was

P = (EFFMAX*F/100)/100 * FE * RFDMwhere FE = photochemical equivalent RFDM = radiant flux density EFFMAX = 3.51 * (VCS - 79.6114) VCS = veld condition score F = (FI/100)* (FT/100) * (FW/100) * 100

In the PUTU models the proportion of incoming radiation intercepted by the vegetation cover, the fractional interception, F_v , is computed from Campbell (1977) using:

	$\mathbf{F}_{\mathbf{v}}$	$= 1 - \exp(-kL)$	[2.23]
where	k	= 0.7	

In the case of row crops Charles-Edwards and Lawn (1984) suggest;

$$F_v = 2 \beta F_{vo} / (1 + F_{vo})$$
 [2.24]

where β is the proportion of ground surface area covered by the downward projection of the leaf canopy and \mathbb{F}_{v_0} is the proportion of the incident energy intercepted at the row centre around solar noon.

Pastures normally present a random distribution of basal cover which makes the former of these two equations the more suitable. This analysis of the impact of radiation, temperature and water constraints on photosynthetic rate, accounts for environmental limitation on vegetation growth rate. The formulation of the appropriate mathematical expressions used in the PUTU 14 model will now be given:

Fractional radiation interception, F_i , absorbed by the canopy is expressed using Eq 2.22:

	AL	= BL / SPL		
thus:	FI	= (1 - EXP(-0.7 * AI))	.)) * 100	[2.25]
where:	FI	= Fractional intercept	ion (%)	
	AL	= Leave area index		
	BL	= Live leaf mass	(kg ha ⁻¹)	
	SPL	= Specific leaf area	(500 kg ha ⁻¹)	
	thus: where:	AL thus: FI where: FI AL BL SPL	AL= BL / SPLthus:FI= (1 - EXP(- 0.7 * ALwhere:FI= Fractional interceptAL= Leave area indexBL= Live leaf massSPL= Specific leaf area	AL= BL / SPLthus:FI= $(1 - EXP(-0.7 * AL)) * 100$ where:FI= Fractional interception (%)AL= Leave area indexBL= Live leaf mass (kg ha ⁻¹)SPL= Specific leaf area (500 kg ha ⁻¹)

Thus $\mathbb{FI} \equiv \mathbb{F}_{v}$ and $\mathbb{E}q$ 2.24 differs from PUTU 13 in that the latter contained an adjustment (BCFUNC) for basal cover.

Formulation of RCC in PUTU 14 grassland

From Eq 2.21 it is evident that ϵ_q is a function of solar radiant flux density. Over weekly periods it has however been found to be reasonably constant because, in a given climate (locality), radiation over such periods during a growing season vary little (see De Jager & Venter, 1978). Because of this Monteith (1990) suggested that ϵ_q (actually RCC) is a conservative quantity. Based upon this argument it is logical to expect minor changes in RCC with locality and stage of growing season. These changes have been neglected in the PUTU models, but could form a basis for improvement in the future. This implies that the influence of construction and maintenance respiration is also accounted for in RCC.

The radiation-photosynthesis conversion coefficient, RCC, is defined as the amount of dry matter produced per unit of radiation intercepted by the foliage. The value off RCC for a wide range of crops is reported by Monteith (1990) to vary from 0.4 μ g J⁻¹ to 1.5 μ g J⁻¹. On the other hand, Jones & Kiniry (1986) use a value of (2.5 μ g CH₂O J⁻¹) in CERES-MAIZE.

The early PUTU models (De Jager, 1976) utilized a value of $\alpha = 100\mu g \text{ CO}_2 \text{ J}^{-1}$ for photosynthetically active radiation actually stored in the chloroplast. For such approach radiation use efficiency in terms of carbohydrate and total radiation for a full crop canopy is approximately $\epsilon_q = 2.7 \%$ yielding RCC = 2.7 $\mu g \text{ CH}_2\text{O J}^{-1}$.

Such approach was adapted in PUTU 11 (see Fouché, 1992) where net photosynthesis was computed as gross photosynthesis less maintenance respiration and construction respiration, the latter two approximately 50 % of gross photosynthesis (CONS = 0.5), where

P = (EFFMAX*F/100)/100 * FE * RFDM thus CONS = construction respiration (= 0.5) where COMM = carbohydrate/CO₂ coefficient (= 0.68) DMG = COMM * P * (1 - CONS)

In PUTU 14 quantum yield has been expressed in terms of the readily physically conseptualisable slope of photosynthesis curve at low radiation $\alpha = 4 \ \mu g \ CH_2 O \ J^{-1}$. Thus an RCC of approximately unity would require an $\epsilon_q = 25 \ \%$ (De Jager, 1976).

In PUTU 14 construction respiration and maintenance respiration have been accounted for in RCC. Thus,

RCC =
$$\alpha \epsilon_{\alpha}$$

For grassland the assumption was made the $\epsilon_{q} = 77 \%$, yielding

RCC = 4 (77/100)
=
$$3.1 \ \mu g \text{ CH}, \text{O J}^-$$

This operation takes place in

[A, 31] ALFA = 4 : EFFQ = 0.775 : RCC = ALFA * EFFQ

26

and [A, 102] was modified so as to read

[A,102] P = RCC * RFD * F / 100

This algorithm can be reconciled with photosynthesis in the following manner.

Consider the alternative definition

$$RCC = \nabla W / \nabla Q$$

where $\forall W$ is the gross amount of plant dry matter accumulated over a given period of time (here one day) and Q is the amount of solar energy intercepted during the same period, thus.

RCC = $(\nabla_{pv} - \nabla_R)$. 0.68/ ∇Q

where ∇_{R} denotes total respiration, and R denotes dark respiration

Following Charles-Edwards et al (1986),

where:

 $= a_0 \nabla_{pv} + b_1 \nabla Q$ ₽_R = construction respiration constant (0.14)a。 = maintenance respiration constant (between 0.0143 and 0.0054) \mathbf{a}_1 = empirical constant 0.4 μ g (CO₂)J⁻¹ (in terms of light energy) b₁ = elemental nitrogen content of the plant Ν = maintenance respiration constant b, $\mathbf{a}_{0}\nabla_{\mathbf{p}\mathbf{v}} + \mathbf{a}_{1}\mathbf{W}$ $\nabla_{\mathbf{R}}$ $\nabla_{\mathbf{R}}$ $= a_0 \nabla_{pv} + b_0 N$ $\nabla_{\mathbf{R}}$ $= \mathbf{a}_{0} \nabla_{\mathbf{n}\mathbf{v}} + \mathbf{b}_{1} \nabla \mathbf{Q}$ $= \nabla_{pv}(1-a_0)/\nabla Q - b_1$ RCC

hence

Charles-Edwards <u>et al</u> (1986) suggest that since b_1 has a magnitude of approximately 0.14 μ g (CH₂O)J⁻¹ it could decrease total photosynthesis by only one-tenth and hence could be reasonably neglected. In early PUTU models maintenance respiration was

however accounted for using the equation of McCree (1974) (see [A, 369] Fouché, 1992), thus

$$a_o = MAIN = (C30*(.044 + .0019*TNITE + .001*TNITE^2*CLIVE)$$

Where nighttime temperature was estimated using the maximum temperature from the previous day (TMXPD) and the current day minimum (AMN(J), thus

$$TNITE = (TMXPD + AMN(J)) / 4$$

This routine is still available in PUTU models, but has been neglected in the grassland PUTU 13 and PUTU 14 versions. Instead, construction respiration, $a_0 \nabla_{pv}$, and, accepting the conservativeness in ∇Q , its sum with maintenance respiration, is computed according to CONS = 0.5.

Computation of overall environmental limiting factor and plant daily dry matter gain

The existing Gaussian function for temperature, \mathbb{F}_{i} , was retained, hence:

[A,325]	ATEMP = (AMXT + AMNT) / 2		
[A,79] thus:	$F_t = (EXP(-1 * ((ATEMP - 30)^2) / 360)) * 100$ [2	.26]	

As was the logistic control of water status on photosynthetic rate, F_w

[A,351-357]	$\mathbf{F}_{\mathbf{W}}$	= 1 / (1 + EXP[0.1*(WPC - LWP)]) [2.27]	7]
[A,83]	$\mathbf{F}_{\mathbf{W}}$	$= \mathbf{FW} * 100$	
[A,29] where:	WPC	= Critical daily leaf water potential at which wilting commences (kPa	1)
	LWP	= Existing daily leaf water potential (kPa)	

Leaf water potential was computed using an empirical daily hydraulic conductivity, HYCON,

[A,349]	LWP	$= E_{o} / HYCON$
where:	E	= Potential evaporation (mm)
[A,33]	нусо	N = Hydraulic conductivity (0.01 mm kPa ⁻¹)

28

The radiant flux density (RFD) effective in photosynthesis and evaporation is given by:

[A,332]	ALPHA	= 0.21	
[A,333]	BETHA	= 0.71	
[A,334]	DAYFRC	= SUN / DALEN(NMNTH)	
where:	DAYFRC	= Proportion of day with no cloud	
[A,69]	SUN	= Unclouded sunlight duration (h)	
[A,13,15-17]	DALEN(NM	NTH) = Maximum possible sunlight hours for month nu	mber
		NMNTH (h)	
[A,335]	ALPHA	= 0.29 for DAYFRC > .5	
[A,336]	BETHA	= 0.5 for DAYFRC > $.5$	
[A,337]	RFD	= SOLK * 10 ⁶ *(ALPHA + BETHA * DAYFRC)	[2.28]
where:	RFD	= Radiant flux density (J $m^{-2} d^{-1}$)	

The solar constant, SOLK, is computed from:

[A,329]	Х	= ((JDA + 10) / 365) * 360
[A,330]	X	= X * 0.01745
[A,331]	SOLK	= $30.85 + 12.65 * COS(X)$ (MJ m ⁻² d ⁻¹)
	JDA	= Calender day counter from 1 st January

Potential CH₂O assimilation ($P = kg CH_2O ha^{-1} d^{-1}$) is simulated as follows;

	Р	= EFF/100 *	FE * RFDM	(PUTU 11)	[2.29]
[A,102]	Р	= RCC * RH	FD * (F / 100)	(kg m ⁻² d ⁻¹)(PUTU 14)	[2.30]
	RCC	= Radiation	-photosynthesis co	onversion coefficient ($\mu g J^{-1}$)	
thus:	Р	= P * DALE	N(NMNTH) * 36	500 * 10^4 (PUTU 11)	[2.31]
[A,103]	Р	= P * 10	(kg ha ⁻¹ d ⁻¹)	(PUTU 14)	[2.32]

The value of RCC in the PUTU 14 model is around 3, inferring a $\alpha = 4 \ \mu g \ J^{-1}$ and $\epsilon_q = 77 \%$. Charles-Edwards <u>et al</u> (1986) and Monteith (1990) reported RCC values of 1.3 $\mu g \ J^{-1}$ and 1.5 $\mu g \ J^{-1}$ respectively.

Results from the PUTU 13 and PUTU 14 model suggest that this approach towards simulating dry matter assimilation is reliable (see Fouché, 1992, Du Pisani, 1992 and the

present study). Furthermore it suggests similarities in the growth patterns between *C. ciliaris* L. cv. Molopo and *D. eriantha*. This was the main reason for the belief that PUTU grassland models, especially PUTU 13 are capable of simple adaptation to the simulation of dry matter assimilation in other subtropical dryland pastures. Little change in Eq 2.31 need be made.

As explained, gross photosynthesis in PUTU 11 and PUTU 13 is reduced by construction and maintenance respiration (taken to be 50% of gross photosynthesis) and calculated in kg ha⁻¹ at 100% basal cover (see Eq 2.29, 2.30, 2.31 and 2.32). Here RCC = $\alpha \epsilon_q$ = 3.1 µg CH₂O J⁻¹ In PUTU 14 this was simplified to express net photosynthesis as

[A,103]	Р	= RCC * 10	(kg ha ⁻¹ d ⁻¹)
with	RCC	= $\alpha \epsilon_{q}$	
		= 4 * 77/100	

this simplified calibration. By trial and error for $\epsilon_q D$. eriantha was found to be 77 %. The value of ϵ_q is in arte fact of the expressions utilized for factors \mathbb{F}_v , $\mathbb{F}_i \mathbb{F}_w$ and particularly \mathbb{F}_t . Thus the constant α which will have to be determined empirically for any new specie which is to be modelled. Expressing P as in Eq 2.31 does however simplify evaluation of ϵ_q .

Re-inclusion of conservation and maintenance respiration would probably improve the equation somewhat. The simplistic approached developed here does however form a convenient basis for refinement and indicates the expected magnitude of the significant variables.

2.6 Carbohydrate translocation

For the translocation rate of carbohydrates to the different organs, viz; root, stubble, culm, leaf and seed, Booysen (1983), Fouché <u>et al</u> (1986), in PUTU 11 accepted carbohydrate to be a source sink driven relationship. In PUTU 14 this was simplified using the theory of Furniss (1982). PUTU 11 assumed that carbohydrate translocation

to different plant organs, is driven by the ratio of the existing mass of the given organ to the total plant mass and that the mass in each plant organ strives to attain an optimal proportion of the total plant biomass. The translocation of carbohydrate according to this law is illustrated in Fig 2.4. A maximum translocation rate is imposed for each growth stage and translocation only proceeds if sufficient reserves are available. This hypothesis was successfully adopted and tested in the PUTU 11 model (Fouché <u>et al</u> 1986; Fouché, 1992).



Figure 2.4 Typical relationship between the fraction of the maximum translocation rate and the desired proportion of plant mass shortage (Fouché, <u>et al</u> 1986).

When developing PUTU 14 the mode of partitioning of dry matter assimilate was changed. This represent one of the major changes made to the model.

Squire (1990) modelled dry matter partitioning to the different plant components as follows. Let, W_0 , be the dry mass of the plant at growth initiation. Then during a period of growth, t_w , dry mass change can be represented as the product of the corresponding growth rate of the whole plant (Γ) and a partitioning factor, p. Mass of a plant structure or component, W_s is given by:

 $W_s = p\Gamma t_w$

and the current mass of a given plant component, W_s , maybe related to the mass of the whole plant, W_t , after a given time t_w ; as follows:

$$\mathbf{W}_{s} = \mathbf{p}(\mathbf{W}_{t} - \mathbf{W}_{o})$$

The Squire(1990) theory has been implemented in the PUTU 14 model. The partitioning factors, p, or daily PROportion of assimilate routed to root, stem, leaf, culm and grain are coded DPROR, DPROS, DPROB, DPROC and DPROG, respectively. Each partitioning factor changes from one growth stage to the next, according to Fig 2.5.

Reproductive Growth (Stage 2)

Culm	Leaf	Stubble
0.2	0.3	0.1
	Root 0.4	· ·

Culm	Leaf	Stubble	Grain	
0.34	0.17	0.2	0.09	
Root				
0.3				

Seed Growth (Stage 3)

Seed fall and dormant (Stage 4,5)

Culm	Leaf	Stubble
0.3	0.1	0.1
Ro	oot	
0.	. 5	

Figure 2.5 Partitioning factor for dry matter assimilate (kg ha⁻¹ d⁻¹) in the different growth stages. For the status variables culm, leaf, stubble, grain and root.

In the model, DPROR, DPROS, DPROB, DPROC and DPROG are set to either zero or the values given in Fig 2.5 at the commencement of each growth stage.

The algorithms for the partitioning of daily dry matter gain, DMG (kg ha⁻¹ d⁻¹), actually evaluate the amount of carbohydrate (kg ha⁻¹ d⁻¹) sent to the different plant organs. These parameters are R(1), R(2), R(3), R(4) and R(5) as defined in Fig 2.1

[A,414]	R(1)	= DBL
[A,415]	R(2)	= DPROS * DMG
[A,416]	R(3)	= DPROR * DMG
[A,417]	R(4)	= DPROC * DMG
[A,418]	R(5)	= DPROG * DMG
remembering	:	
[A,104]	DMG	= $P * FACN$ (kg ha ⁻¹ d ⁻¹)
where	FACN	= nitrogen deduction factor in leaf (see Chapter 2.7.2)

In contrast PUTU 11 and PUTU 13 computed partitioning at 100% basal cover and hence in PUTU 14 the BCFUNC term could be removed from the equations corresponding to these.

Due to senescence, live tissue mass is transferred to dead tissue. Parameters X2, X1, X3, X9 and X8 are defined as the daily rate of senescence and they differ according to growth stage. The values for the leaf, stubble, root, culm and grain components, respectively are given in Table 2.2

Similarly to dry matter partitioning, the values of the senescence factors are set at the commencement of each growth stage. In principle, the rate of senescence is assumed to be directly proportional to the current live mass of the relevant component and the senescence factors for, for example, the vegetative growth stage X1, X2, and X3 are defined:

[A,132]	· X(1)	= 0.001 * SL	for stem
[A,133]	X(2)	= 0.001 * BL	for leaf
[A,134]	X(3)	= 0.001 * RL	for root

32

Table 2.2Senescence factors accounting for the daily transfer of mass (kg ha⁻¹ d⁻¹)from living plant structures to dead tissue in the different growth stages.(Source code statement numbers are given in parenthesis).

Plant Structure			Growth stage		
	Vegetative	Reproductive	Seed	Seed fall	Dormant
Stubble (X1)	0.001 [A,132]	0.001 [A,152]	0.002 [A,171]	0.002 [A,190]	0.020 [A,205]
Leaves (X2)	0.001 [A,133]	0.001 [A,153]	0.002 [A,172]	0.010 [A,191]	0.200 [A,206]
Root (X3)	0.001 [A,134]	0.001 [A,154]	0.003 [A,173]	0.003 [A,192]	0.003 [A,207]
Grain (X8)			0.150 [A,174]	0.500 [A,200]	0.500 [A,200]
Culm (X9)			0.001 [A,175]	0.050 [A,201]	0.250 [A,201]

The current status of living tissue mass (kg ha⁻¹ d⁻¹) for leaf, stubble, root, grain and culm are denoted by parameters, BL, SL, RL, GL and CL (see, Section 2.2). Live mass balances (kg ha⁻¹) are computed in daily iterations using the following :

[A,424]	BL	= BL + R(1) - X(2)	leaf
[A,426]	SL	= SL + R(2) - X(1)	stem
[A,432]	RL	= RL + R(3) - X(3)	root
[A,430]	GL	= GL + R(5) - X(8)	grain
[A,428]	CL	= CL + R(4) - X(9)	culm

Furthermore dead tissue mass is transferred to trash, TRD, at rates determined by X7, X6, X4 and X5 for leaf, stubble, grain and culm, respectively (see, Section 2.2). These rates also differ for each growth stage (Table 2.3)

Table 2.3Trash factors for the transfer of dead tissue to trash, in the different
growth stages. (Source code reference are given in parenthesis).

Plant Structure	Growth Stage			
	Seed	Seed fall	Dormant	
Stubble (X6)	0.001 [A,176]	0.005 [A,198]	0.500 [A,210]	
Leaves (X7)	0.001 [A,177]	0.005 [A,199]	0.005 [A,211]	
Grain (X4)		0.300 [A,193-196]	0.500 [A,208]	
Culm (X5)		0.005 [A,197]	0.002 [A,209]	

The trashing rates (kg ha⁻¹ d⁻¹) in the dormant growth stage for SD, BD, GD and CD (see, Section 2.2) are as follows;

[A,208]	X(4)	= 0.5 * GD	grain
[A,209]	X(5)	= 0.002 * CD	culm
[A,210]	X(6)	= 0.005 * SD	stem
[A,211]	X(7)	= 0.005 * BD	leaf

Use of this transfer matrix X permits daily computation of the standing dead mass balance (kg ha⁻¹). The actual gain in dead tissues due to senescence for plant organs, stubble, leaf grain, and culm are defined as:

BD	= BD + X(2) - X(7)	leaf
SD	= SD + X(1) - X(6)	stem
CD	= CD + X(9) - X(5)	culm
GD	= GD + X(8) - X(4)	grain
RD	= RD + X(3)	root
	BD SD CD GD RD	BD = BD + X(2) - X(7) $SD = SD + X(1) - X(6)$ $CD = CD + X(9) - X(5)$ $GD = GD + X(8) - X(4)$ $RD = RD + X(3)$

The amount of trash accumulated (kg ha⁻¹ d⁻¹) is constituted of the sum of the daily transfers of mass from the different plant organs ;

$$[A,435] TRD = TR + X(4) + X(5) + X(6) + X(7)$$

2.7 Soil mass balances

2.7.1 Water balance

Water is indispensable to the growth, development and maintenance of vegetation. A schematic representation of the different components involved in the water balance of a vegetative surface are given in Fig 2.6



Figure 2.6 Components of the water balance model at a vegetative surface.

Stochastical and Markov matrix simulation of surface balances whereby one variable (rainfall) was measured and the rest simulated have been attempted (Richardson, Hanson & Huber, 1987). Other models have concentrated on only certain components. Such as for example, the equations for determining the effective rainfall (Snyman & Van Rensburg, 1986), evapotranspiration (Makink & van Heemst, 1975) and soil

moisture status (Renard, Shirley, Williams & Nicks (1987). Since weather determines the ultimate control on plant water use and growth, most operational models seek to use weather data input to simulate these aspects. The most important component is evapotranspiration, about which much has been written pertaining to grassland (De Jager <u>et al</u>, 1980; Hayes, O'Rourke, Terjung, & Todhunter, 1982; Van Keulen, 1982; Wight, Hanson & Whitmer, 1984; Wight, Hanson & Cooley, 1986; Wight & Hanson, 1988).

Wisiol (1987) reviewed eight equations with reference to the water balance and plant production. Most important was the equation of Penman (1948) later modified by Monteith (1965) as quoted by Thornley & Johnson (1990). The formulation of this, known as the Penman-Monteith equation is as follows;

where:

$$E = \frac{s \oint N + \lambda \gamma G_a \Delta P_{va}}{\lambda [s + \gamma (1 + G_a/G_c)]}$$

$$E = \text{Transpiration (mm d-1)}$$

$$\oint N = \text{Energy available for evaporation (J m-2 d-1)}$$

$$\Delta P_{va} = \text{Vapour density deficit (kg m-3)}$$

$$G_c, G_a = \text{Canopy and boundary layer conductances (m s-1)}$$

$$\lambda, \gamma, s = \text{Physical parameters (MJ kg-1, kg m-3 K-1, kg m-3 K-1; respectively)}$$

$$[2.34]$$

In the PUTU grassland models the Priestley-Taylor variant of this equation is used. When wind and atmospheric vapour measure are available Eq 2.34 is used.

The potential short grass evaporation rate in mm d^{-1} , E_0 , is estimated using the Priestley- Taylor equation modified for high temperatures (see De Jager, 1992). Thus the normal expression

	E	$= 1/\lambda [\Delta/(\Delta + \gamma)] 1.28(R_n - G)$
reads	E	= $1/\lambda [\Delta/(\Delta + \gamma)] 1.28[0.63 \text{RFD}]$ for $T_{mx} < 20 \text{ °C}$
or	E	= $1/\lambda [\Delta/(\Delta + \gamma)] [1.28 + 0.08(T_{mx}-20)] [0.63 RFD]$ for $T_{mx} > 20 \text{ °C}$
where:	λ	= the coefficient of latent heat of evaporation at constant
		temperature (2.45 MJ kg ⁻¹ or 2450 J g ⁻¹)

Δ	= the slope of the saturation vapour pressure temperature
	curve (kPa °C ⁻¹)
γ	= the psychometric constant (kPa °C ⁻¹)
T _{mx}	= daily maximum air temperature ($^{\circ}C^{-1}$)
R _n	= net radiant flux density (W m^{-2})
G	= soil heat flux (W m ⁻²)
RFD	= Radiant flux density (W m ⁻²)

In the PUTU 14 model, the algorithms for the Priestley-Taylor formula are:

[A,342]		PE	= PECONS * EE	[2.35]
	where:	PECON	S = Parameter in the Priestley-Taylor formula	
		EE	= Equilibrium evaporation (mm d ⁻¹)	
[A,339]		PECON	dS = 1.28 for AMXT < 20 °C	
	or:			
[A,340]		PECON	dS = 1.28 + 0.08 * (AMXT - 20) for AMXT > = 20 °C	
[A,341]		EE	= GS * 0.63 * RFD / 2.45	
	where:	GS	= $\Delta / (\Delta + \gamma)$ and is given by	
[A,326]		GS	= 0.4019914+).01725101*ATEMP-0.0001485* ATEMP ²	
		RFD	= Radiant flux density (W m ⁻²)	•

Soil properties like depth and texture determine the amount of water stored in the soil profile as well as the availability of the water for plant growth and production (Hillel, 1977). To simulate growth and production these factors need to be taken into account. Most of all, the water balance equations have to be properly calibrated and validated.

De Jager <u>et al</u> (1980), illustrated the usefulness of the water stress function, F_w , in the PUTU 11 grassland simulation model for assessing production potential in grassland. Snyman (1982) compared results given by PUTU 11 with measured Et/E_0 -coefficients. He concluded that the PUTU grassland simulation model, accurately simulates soil water status.

37

The depth of the rooting zone (SDP = 300 mm here) is an important input parameter. Computation proceeds in two layers, the surface layer being SDP1 = 100 mm [A, 38]. In PUTU 11, Booysen (1983) and Fouché (1984, 1992) simulated the soil water retention curve using the method originally developed by De Jager (1976) for PUTU 2. For the PUTU 13 and 14 models the soil moisture potential (GWP) is simulated following the work of Campbell (1977). The author and Fouché (1992) computed this as follows;

[A,324]	GWP	= -1500 * (WJ / WHC4) ^ M1	[2.36a]
where:			
[A,44]	M1	= (LOG(10) - LOG(1500)) / (LOG(WHC1) - LOG(WHC4))	
[A,404]	WJ	= SLWAT(J + 1)*100 current day soil water content (%)	
[A,27]	WHC1	= water holding capacity at 10 kPa (%)	
[A,28]	WHC4	= water holding capacity at 2440 kPa (%)	
[A,401]	SLWAT $(J + 1)$	= soil water content of the previous day (mm)	

Both WHC1 and WHC4 are calculated from the clay content of the soil, using the Hudson equation as quoted by De Jager, van Zyl, Kelbe & Singels (1987).

This differs markedly from PUTU 11 where an exponential spline function is used. Four splines apply and the computational procedure is here included for comparative purposes.

	WJ =	= SLWAT(J) / SDP * 100	
	AII	= ((log(2440)-log(148.1))/(WHC	23-WHC4)
	AI2	= ((log(148.1)-log(22.1))/(WHC	2-WHC3)
	AI3	= ((log(22.1)-log(10))/(WHC1-V	VHC2)
	WJ	= % Soil moisture	
	SDP	= Soil depth (mm)	
if	WJ	> = WHC3 then	go to A
	WST	= WHC4 : GWPST = 24.4	
	COEF	= -1 * AI1	go to C
if	WJ	> = WHC2 then	go to B
	WST	= WHC3 : GWPST = 1.48	
	COEF	$= -1 * AI2 \qquad \text{go to } C$	2
	WST	= WHC2 : GWPST = .22	
	COEF	= -1 * AI3	

where:

A

В

thus:

С

ARGU	=	COEF * (WJ-WST)	[2.36b]
GWP	=	GWPST * EXP(ARGU)	[2.36c]
GWP	=	GWP * (-100)	

The difference between the retention curves of PUTU 11 and PUTU 14 is illustrated in Fig 2.7. It is evident that only at soil moisture percentages lower than 5 %, do soil water potentials differ. The simulation in PUTU 14 is preferred, because the manner in which it is constructed ensures reliability at the dry end of the water content range.



Figure 2.7

Difference in calculated soil water potential between the PUTU 11 and PUTU 13 and PUTU 14 simulation models.

The daily soil water balance is computed using the finite layer (reservoir) cascade technique. Firstly, only when the soil water content, in the surface layer SLWAT1(J) exceeds the field capacity of that level ,FC1, will access water drain (DRAIN1) to the second soil layer. If the soil water content in the rest of the soil profile, SLWAT2(J), exceeds the field capacity, FC2, of this layer, then only will deep drainage take place out of the root zone, DRAIN2. This water is lost to the vegetation. These processes are simulated as follows:

[A,375,376]	SLWAT1(J)	= SLWAT1(J) + RAINF + IRRIG - TRANS1 - SLVAP (mm)
Should	SLWAT1(J) >	FC1 then
[A,383]	DRAIN1	= (SLWAT1(J) - FC1)* DRN1P (mm)
[A,382]	DRN1P	= 1 rate of drainage parameter
[A,67]	RAINF	= Daily Precipitation (mm)
	IRRIG	= Irrigation (mm)
[A,371]	TRANS1	= Daily transpiration (mm)
[A,370]	SLVAP	= Soil vaporation (mm)

Finally, the soil water content in the surface layer is computed

 $[A,385] \qquad \qquad SLWAT1(J) = SLWAT1(J) - DRAIN1$

A similar process occurs in the second layer, viz.

[A,386]	SLWAT2(J)	= SLWAT2(J) + DRAIN1 - TRANS2
[A,389] should	SLWAT2(J) >	> FC2 then
[A,390]	DRAIN2	= (SLWAT2(J) - FC2) * DRAINP
where,		
[A,33]	DRAINP	= 0.9
[A,391]	DRAIN2	= 0 for DRAIN2 $< = 0$
[A,392]	SLWAT2(J)	= SLWAT2(J) - DRAIN2

For grass species with shallow roots, which effectively saturate the root zone, this simple model has shown to be reliable (Fouché, 1992; du Pisani; 1992).

Soil evaporation and transpiration rate

The rate of evaporation from the soil surface and transpiration are computed following, De Jager, Van Zyl, Bristow & Van Rooyen (1982).

First the proportion of potential evapotranspiration used in transpiration, B, is determined using an empirical relationship;

[A,346-348] B = (0.1 + (AL / 3) * 0.9)[A,371] TRANS1 = B * FW * PE

40

Daily potential plant evaporation is assumed equal to the difference between PE and transpiration, TRANS. Thereafter evaporation from the soil surface SLVAP is assumed to decay exponentially (as quantified by FG the drying parameter) with time since the previous wetting event of > 5 mm, CNT(J+1).

[A,361-364] FG = EXP(-0.5 * CNT(J + 1) [A,370] SLVAP = (1 - B) * FG * PE

The surface soil layer however is not allowed to dry below permanent wilting point of layer one, PWP1, in which case it is constrained by;

[A,373]	SLVAP =	SLWAT1(J) - PWP1
[A,371]	TRANSI =	B * FW * PE

Run-off is computed using a simple run-off routine described by Jones & Kiniry (1986)[°] [A, 365 - A, 369]. It is important that each of these components appear in the soil water balance calculations.

2.7.2 Nitrogen balance

Haynes (1986) reviewed factors involved in the nitrogen balance of an ecosystem (see Fig 2.8). Nitrogen mineralization, -immobilization, -leaching and -uptake are factors involved in the maintenance and balance of such an ecosystem. The rate of these processes are however dependant on external factors i.e. temperature, carbon to nitrogen ratio in the soil organic matter, water penetration, nitrogen demand and rate of nitrogen application.

A mechanistic simulation of N-mineralization, N-immobilization, N-leaching, N-uptake and distribution through the plant was included in the PUTU 14 model. Singels & Manley (1991) gave detailed reasons for choosing the model of Seligman & Van Keulen (1981) and Van Keulen & Seligman (1987).



Figure 2.8 Nitrogen balance in an ecosystem (Haynes, 1986)

It was found to operate reliable for wheat (Singles & Manley; 1991). A brief description of this model and the processes simulated in the soil-plant system will now be given. As far as possible, shortcomings will also be highlighted. Computer code for the soil - and plant nitrogen balance is available in Appendix B and C, respectively.

Soil Nitrogen balance

Plant and animal residues decompose and make N available (mineralization) to the vegetation. This assists recirculation of carbon (CO_2) to the atmosphere (Haynes, 1986). The decomposition of fresh organic matter thus has an impact on maintaining a nitrogen balance (Floate, 1981; Woodmanse, Vallis & Mott, 1981). Factors that influence the rate of decomposition are temperature (Alexander, 1977) and soil water (Wilson & Griffin, 1975) and C/N ratio of organic matter (Aber & Melillo, 1980). Models simulating this process have been reviewed by Frissel & Van Veen (1981) and De Willegen & Neeteson (1985)

The model takes external rate limiting factors for temperature (FTMIN), soil water (FWMIN) and carbon/nitrogen ratio in the organic matter (FCNR) into account when simulating the decomposition rate of organic matter, or mineralization (DECR). The fundamental algorithm is;

[A,18]		DECR	= RDECR(L) * FTMIN * FWMIN * FCNR (d ⁻¹)
	where:	DECR	= decomposition rate of fresh organic matter (d)
[B ,9-1]		RDECR(L)	= maximum fraction of organic matter decomposed (0.8, 0.05,
			0.0095 kg ha ⁻¹ d ⁻¹ , depending on the FOM(L)/IFOM(L) ratio
•			defined below)
[B ,12]		FTMIN	= 1/(1 + EXP(-0.15 * (T(L) - 16)))
[B ,13]		FWMIN	= 1 / (1* EXP(-GWP/1500))
[B,15]		CNR	= C:N ratio
[B,16-17	']	FCNR	= EXP(-0.693 * (CNR -25) / 25)
[B ,22]		FOM(L)	= fresh organic matter in soil layer L (kg ha ⁻¹)
[A,452]		IFOM(L)	= initial amount of fresh organic matter in soil layer L (kgha ⁻¹)
[A,445]		T(L)	= average daily soil temperature in layer L (°C)
[A,324]		GWP	= soil water potential (kPa)

The expression [B,13] for FWMIN represents a marked change from the function used by Singels & Manley (1991). It appeared that the exponential function for describing water limitations on mineralization rate, better quantified these process in the current PUTU 14 model. The original expression read;

	FWMIN	= ((V(L)-V15(L))*.5/(V01(L)-V15(L)*.5)
here	V(L)	= soil water content (mm m ⁻¹)
	V01(L)	= Soil water content at -10 kPa (mm m ⁻¹)
	V15(L)	= Soil water content at -1500 kPa (mm m ⁻¹)

w

Graphical representations of factors FTMIN, FWMIN and FCNR on the decomposition rate of organic matter are given in Fig 2.9 - 2.11, respectively.

43



Figure 2.9 Relationship between mean daily soil temperature and a function describing how temperature limits mineralization rate.



Figure 2.10 Relationship between soil water potential and the function for describing how soil water limits the rate of mineralization



Figure 2.11 Relationship between C:N ratio and the function describing how C:N ratio limits rate of decomposition of organic matter.

N-mineralization from fresh organic matter and stable organic matter (humus) are computed by factors NMINFOM(L) and NMINHUM(L), respectively; in the following manner:

[B,19] NMINFOM(L) = NFOM(L) * DECR

 [B,24]
 NMINHUM(L) = NHUM(L) * DMINR * FWMIN * FTMIN

 where,
 NFOM(L) = nitrogen in fresh organic matter for soil layer L (kg ha⁻¹)

 NHUM(L)
 = nitrogen in humic fraction for soil layer L (kg ha⁻¹)

 DMINR
 = relative decay rate of humus (8.3 * 10 ⁻⁵)

Nitrogen immobilization rate from fresh organic matter is given by NIMMFOM(L), and thus net mineralization rate NMINNET(L), and computed, thus:

 [B,20]
 NIMMFOM(L)
 = DECR * FOM(L) * 0.02

 [B,25]
 NMINNET(L)
 = .8*NMINFOM(L) + NMINHUM(L)-NIMMFOM(L)

Once NMINNET(L) exceeds zero, nitrogen is mineralized and added to the ammonium fraction NH4(L). However, nitrogen can also be immobilized into the organic fraction. This phenomena will take place when NMINNET(L) drops below zero [B, 25-28).

Indications that nitrogen in the form of NH_4^+ nitrifies to NO_3^- have been reported by Van Veen & Frissel (1981,1982) and Haynes (1986). Lees & Quatel (1946) as quoted by Kruh & Segal (1980) and Hagin & Welte (1984) used the following equation to describe such nitrification rate;

 $(NO_{3}^{-})_{N} = K (t - t_{1/2})$ $M - (NO_{3}^{-})_{N}$ where, K = constant $M = \text{asymptotic value of } (NO_{3}^{-})_{N}$ $t_{1/2} = \text{time when } (NO_{3}^{-})_{N} = M/2$

In the PUTU 14 model, this process is simulated by a simple nitrification routine for each soil layer, which states that 25 % of NH_4^+ nitrifies per day [B, 32-37] (see Singels & Manley, 1991).

The balance between the loss of nitrogen through leaching in rain water and the nitrogen applied determine the amount of nitrogen available for dry matter production, AN(L), and thus daily dry matter gain [B, 50-54). The mineral nitrate status of the soil profile ANC(L) is determined as follows:

[B ,51]	AN(L)	= ((NO3(L)-1) + (NH4(L)-1)*.9)
[B ,53]	ANC(L)	= AN(L) / W(L)
where,		
	AN(L)	= available inorganic nitrogen in the soil for soil layer L
		(kg ha ⁻¹)
	ANC(L)	= Nitrate content of soil water for layer L (kg ha ⁻¹ mm ⁻¹)

Mineralization rate of organic matter is not only a function of the environmental factors. The amount and concentration of stable organic matter HUM(L) and fresh organic matter FOM(L) present in the soil also greatly influence mineralization rate. In this study these values were guesstimated, because of lack of information. This must be remembered should any application of these results be contemplated.

Unfortunately microbial biomass could not be incorporated in the present model. Van Keulen (1982) listed the problems preventing this. Studies to investigate the influence of microbial population size for decomposing organic matter under dryland conditions with species like *D. eriantha* etc. are required.

Exponential relationships for FTMIN and FCNR and FWMIN are here assumed. Whether these hold true under different climate and soil conditions are debatable.

Plant Nitrogen balance

Nitrogen uptake (NUP) by the vegetation is extracted from the transpiration flux of water (TNFLO). It is further influenced by the maximum nitrate uptake rate which the canopy can accommodate (NUPO), the demand for nitrogen from the vegetation (TNDEM), and the amount of soil nitrogen available to the plant for the growth process (TOTAN). Once these are accounted for, nitrate will be withdrawn from the soil [C, 56-

62] and distributed amongst the plant organs in proportion to there relative mass and demand for nitrate. Thus by the law of the minimum;

[C,43-47]	NUP	= MIN(TNFLO,NUPO,TNDEM,TOTAN)
where,		
	СОМР	= subscript referring to the different plant components
		root(1),culm(2) and leaf(3) (kg ha ⁻¹)
[C,42]	TNFLO	= TNMASFLO + TNDIFFLO
[C,23]	NUPO	= 6*(1-EXP(005*(TOTMASS(2) + TOTMASS(3))))
[C,19]	NDEM(COMP)	= TOTMASS(COMP)*(NCMAX(COMP)-NC(COMP))/2
[C,21]	TNDEM	= Σ NDEM(COMP)* EOSF
[C,35]	TOTAN	= AN(L)
[C,24-30]	TNMASFLO	= nitrate due to mass flow (kg ha ⁻¹ d ⁻¹)
[C,38]	TNDIFFLO	= nitrate flow by diffusion (kg ha ⁻¹)
	TOTMASS(COM	MP) = mass in different organs (kg ha ⁻¹)
[C,17]	EOSF	= factor preventing uptake after maturity

Nitrogen distribution amongst the plant organs is a function of individual plant organ demand for nitrogen in relation to the total demand and the uptake rate, hence:

[C,63]	DN(COMP)	= NDEM(COMP)/TNDEM*NUP
[C,64]	N(COMP)	= N(COMP) + DN(COMP)
[C,65]	NC(COMP)	= N(COMP)/TOTMASS(COMP)
where,		
	DN(COMP)	= fraction of nitrogen distributed to plant organ COMP
		(kg ha ⁻¹)
	N(COMP)	= nitrate content for root, culm and leaf, respectively (kg ha ⁻¹)
	NC(COMP)	= nitrate concentration for root, culm and leaf, respective-
		ly (kg kg ⁻¹)

Once simulation of the reproductive growth has been initiated, seed growth commences. The availability of nitrate for the production and formation of seeds depends on;

- the nitrogen demand created by the seeds DN(4), and
- the nitrogen status in the individual plant organs PAN(COMP) (for root, culm and leaf).

Demand set by the seeds DN(4) depends upon the mass of the seeds (TOTMASS(4)), the difference between the optimum and current nitrate concentration (ONC- NC(4)) in the seeds and a reduction factor reflecting nitrogen deficiency in the leaves (RFNS) [C, 55-60].

If the current nitrogen concentration in a given plant organ exceeds the limit set for that organ, then nitrate will be available for growth of seeds. Plant available nitrate PAN(COMP) is a function of:

Withdrawal of nitrogen from plant organs to meet the demand by seed is then computed,

[C,70]		DN(COMP)	= DN(4) * PAN(COMP)/TPAN (kg kg ⁻¹)
	thus,		
[C,71]		N(COMP)	= N(COMP) - DN(COMP)
	where,		
		TPAN	= Total plant available nitrogen for distribution to the seeds
			(kg ha ⁻¹)

Nitrogen distribution to the seeds in growth stage four has been modified from the function used by Singels & Manley (1991) in a wheat model to the approach used by Seligman & Van Keulen (1980), viz

DN(4) = MIN(NTRANSMAX, NDEM(4) (Singels & Manley; 1991) to DN(4) = TOTMASS(4) * (ONC - NC(4)) * RFNS (Seligman & Van Keulen, 1980) To compute the growth limiting factor due to the nitrate status in the leaves (FACN) for processes such as dry matter assimilation; the following equation applies;

[C,76]		FACN	= (NC(3)-NCMIN(3))/(NCOPT(3)-NCMIN(3))
	where:		
		NC(3)	= nitrate concentration in the leaves (kg kg ⁻¹)
		NCMIN(3)	= minimum nitrate concentration in the leaves (kg kg ⁻¹)
		NCOPT(3)	= optimum nitrate concentration in the leaves (kg kg ⁻¹)
			Currently assumed to 80 % of the maximum nitrogen
			concentration in the leaves.

Daily dry matter gain (DMG) is then expressed as the product of CO_2 assimilation and the growth limiting factor due to nitrogen status of the leaves (FACN).

The Van Keulen & Seligman (1987) model here employed, is a first attempt which could be refined.

Certain principal questions still remain to be answered, such as for example

- 1. What is the true maximum and minimum nitrogen concentration in the different plant organs for *D. eriantha* ?
- 2. How do these concentrations change with growth stage from vegetative through to dormancy ?
- 3. Living and dead tissue mass were lumped for each plant organ. To what extent can nitrogen be withdrawn from dead material ?
- 4. To what extent does the nitrogen content in the leaves of *D. eriantha* influence dry matter production? and
- 5. Does the seed withdraw all nitrogen available for seed growth. Since the importance of seed formation in perennial dryland cultivated pasture is merely for reproduction. While seed production constitutes the economic product of annual species such as maize and wheat.

2.8 Calibration of the PUTU 14 model for D. eriantha

2.8.1 Method

Accepted modelling procedures prescribe the use of independent data sets for calibration and validation. Field data procurable (from Dannhauser; 1985) on *D. eriantha* for the 1978/79 season were selected for calibration. Data obtained during the 1979/80 season and data obtained during the 1981/82, 1982/83 and 1983/84 season on both an Avalon and Valsriver soil were used for validation of the refined PUTU 14 model (see Chapter 3).

The simplifications and modifications brought about to the model are detailed in the preceding sections of Chapter 2. The PUTU 14 including these refinements was then calibrated by trial and error. The 1978/79 season proved to be particularly appropriate for this purpose as it was a season in which water limited production, thereby providing a thorough test of the water stress routine.

2.8.2 Results

The parameters calibrated and values obtained are given in Table 2.6, which indicates changes from the values used in PUTU 13. The realistic shape of the simulations obtained in Quadrants A and B in Fig 2.12 verify the functioning of PUTU 14, as does the good agreement between final yields illustrated in Quadrant C.

On this evidence, the model verification and calibration of PUTU 14 may be deemed to have been successful. The data available were few and hence use of the standard statistical test of significance (correlation coefficient, mean square error etc. or even the boot strapping techniques) have limited value. For interest, comparison of simulated versus measured yields for the n = 3 sets of data are given in Table 2.4.





Statistical analyses

Results of the statistical tests undertaken during calibration are given in Table 2.4.

Table 2.4Model performance during calibration (1978/79) on a Westley soil (n = 3).MAE = mean absolute error, RMSE = root mean square error, RMSEs= systematic error, RMSEu = unsystematic error, D = Wilmott index ofagreement and r^2 = coefficient of determination.

Soil Type	MAE t ha ⁻¹	RMSE t ha ⁻¹	RMSE _s t ha ⁻¹	RMSE _u t ha ⁻¹	D	r ²	Slope through origin
Westley	0.066	0.077	0.04	0.06	0.99	0.98	0.98

Indication of the good agreement between measured and simulated yield are given by the low MAE and RMSE. The MAE and RMSE are 0.06 and 0.07 t ha⁻¹, respectively. Which is relatively small by modelling standards.

Both the Wilmott index of agreement (D) (Wilmott, 1982) and the coefficient of determination (r^2) approach unity. This is not surprising in view of the view data sets (n=3).

2.8.3 Discussion

Rainfall

From Table 2.5 it is clear that only September, October and May had more rain than the long term mean for 1978/79. The annual total rainfall was 435.8 mm, or 70 % of normal (625 mm). Furthermore, apart from the early season, 1978/79 was a poor growing season in which water stress probably occurred, making this season ideal for calibration of water stress on growth.

Table 2.5Mean monthly rainfall (mm) for the Agriculture Development Institute ofPotchefstroom in relation to the monthly mean for 1978/1979.

Growth season	Month												
	Jul	Aug	Sep	Oct	Nov	Dec	Jan	Feb	Mar	Apr	May	Jun	Total
1978/79	0	7.5	23.3	83.7	16.7	79.6	84.6	54.5	34.4	17.2	33.4	1	435.8
% of Mean	0	80	119	148	20	83	72	61	46	29	233	17	70

Dry matter gain

The rates of dry matter assimilation and therefore daily dry matter production differ greatly from specie to specie. The new expression for

[A,105] P = RCC * RFD * (F / 100)

included in PUTU 14 accommodates construction and maintenance differently to the previous models. The value of RCC = 3.1 μ g J⁻¹ does however seem to provide reliable growth simulation of *D. eriantha*.

Phenology

The most marked changes came in the phenological controls. These are listed in Table 2.6. Such specie differences were to be expected.

The change in BO from 12 °C for *C. ciliaris* to 10 °C for *D. eriantha* mean that the latter reaches the reproductive stage considerably quicker than the former. Furthermore, since the adaptation for the latter for water stress permits HUCRMN to decrease to only 230 °C rather than 225 °C shows that *D. eriantha* is less adaptable to

54

Table 2.6New values of variables resulting from the calibration of PUTU 14 on 1978/79 season data.

GROWTH PROCESS	GROWTH STAGE	VARIABLE	UNITS	PUTU 13	PUTU 14	LINE
Phenology	1	HUCRMN	°C	225	230	129
	1	BO	°C	12	10	89
	1					126
	1					127
	1					130
	2	MKOUNT	d	60	30	146
	3	MKOUNT	d	100	80	164
	1	BCON	kg ha⁻¹(d ℃)⁻¹	2.0	3.5	130
Dry matter assimilation	1-4	RCC	µд Ј-	3.5	3.1	102
Translocation rate	2	DPROC		0.25	0.2	417
	3	DPROG		0.05	0.09	418
	3	DPROC		0.41	0.34	417
Growth limiting factor	1-4	COEF	kPa-1	0.5	0.1	352

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water stress than is *C. ciliaris*. It is also a shorter season specie as illustrated by shortening of the MKOUNT routines.

By a process of trial and error, the rate of change of phase from reproductive to seed formation and seed formation to seed fall were determined. These values were fixed at 30 days of reproductive growth and 80 days of seed formation, after which seed fall was initiated. Furthermore, when a minimum temperature of 2 °C was reached, dormancy commenced and carbohydrate translocation is terminated.

Use of a respectively slow and rapid leaf development phases with, BCON = 0.8 and BCON = $2 \text{ kg ha}^{-1} (\text{d }^{\circ}\text{C})^{-1}$ appeared (see Booysen; 1983) to overestimate leaf mass. Adoption of BCON = $3.5 \text{ kg ha}^{-1} (\text{d }^{\circ}\text{C})^{-1}$ during the vegetative growth stage of *D*. *eriantha* yielded good results. This single linear growth rate should be tested on other species.

Reproductive growth, seed and seed fall phases

During these three phases, carbohydrate is translocated to the different plant organs in accordance with preferred proportions. These factors differ from growth stage to growth stage. A comparison between the rate of translocation to the different plant organs for D. eriantha and C. ciliaris are given in Table 2.6. From this it is obvious how the translocation rate between leaf and culm were adjusted. It was assumed that D. eriantha requires a higher leaf : culm ratio than C. ciliaris.

Factor limiting rate of mineralization of humic compounds due to plant water status

The equation suggested by Singels & Manley (1991) is given in Section 2.7.2. This equation was replaced by [B, 13] in Section 2.7.2. It's accuracy is verified by the realistic cut-off in simulated and measured yield obtained during the 1978/79 growing season, see Quadrant C in Fig 2.12
Nitrogen distribution to seed

The demand for nitrogen in seeds is controlled by several factors. Singels & Manley (1991) suggested that a maximum nitrogen translocation rate to the seeds (NTRANSMA X) and the demand for nitrogen in the seeds (NDEM(4)) regulates the distribution process. These two factors are functions of :

- 1. The nitrogen turnover in the vegetative parts,
- 2. Plant available nitrogen,
- 3. Fraction of nitrogen in vegetative parts available for distribution,
- 4. Temperature control of translocation to seed
- 5. Maximum nitrogen demand by the seed.

These controls seem to be realistic for crop models for maize and wheat, but uncertain for perennial dryland cultivated pastures. The availability of nitrogen for distribution to seed is however of primary concern. Algorithm, RFNS, [C, 54 and 56] apparently accommodates the process adequately. Only when nitrogen is available, then does distribution to the seed take place.

CHAPTER 3

VALIDATION OF THE PUTU 14 MODEL

3.1 Introduction

Extensive work has been done by Dannhauser (1985) on *D. eriantha*. Essentially this work endeavoured to assess the feasibility of cultivating *D. eriantha* in the Western Transvaal of Southern Africa. The results however, have never been applied in deterministic models.

Data obtained on a Westley soil included two growth seasons 1978/79, 1979/80 where 3 nitrogen fertilization rates; 30 kg N ha⁻¹ + 10 kg P ha⁻¹, 90 kg N ha⁻¹ + 10 kg P ha⁻¹ and 150 kg N ha⁻¹ + 10 kg P ha⁻¹ were applied. Although nitrogen and phosphate were applied, the soil profile (Chapter 1) clearly indicates that existing phosphate was adequate to satisfy the demands of *D. eriantha*. Data from the 1979/80 season was used to validate the model, together with data from an Avalon and Valsrivier soil for seasons 1981/82, 1982/83 and 1983/84 with N-application of 60 kg N ha⁻¹ + 10 kg P ha⁻¹ and 120 kg N ha⁻¹ + 20 kg P ha⁻¹, respectively in each year (see Dannhauser; 1985 and Dannhauser, van Rensburg, Opperman & Van Rooyen, 1987).

3.2 Results and discussion

Rainfall

Annual rainfall for 1979/80 may be compared to the long term mean rainfall in Table 3.1. An annual rainfall of 714.4 mm, or 114 % of normal (625 mm) was reported, with 6 out of the 12 months recording more rain than the long term mean.

Table 3.1Mean monthly rainfall (mm) for the Agriculture Development Institute ofPotchefstroom in relation to the monthly long term mean for 1979/1980.

58

Growth season	Month												
	Juł	Aug	Sep	Oct	Nov	Dec	Jan	Feb	Mar	Apr	May	Jun	Total
1979/80	7.2	84.7	58.3	136.8	81.8	62.0	125.8	112.4	17.9	23.5	4.0	0.0	714.4
% of Mean	191	853	297	241	99	64	107	126	24	40	28	0	114

The 1979/80 season was well endowed with rainfall (114 % of normal). Thus, this fortunately offered an ideal base for testing the nitrogen limitation mechanism of the model.

Yield comparison 1979/80

Yields obtained for *D. eriantha* during 1978/79 (calibration season) and 1979/80 on a Westley soil were 2445 and 5167 kg ha⁻¹, respectively (Dannhauser, 1985). Due to the favourable rainfall, the latter growing season yielded approximately 2700 kg ha⁻¹ more than the former. Results obtained with the PUTU 14 model accurately simulated this tendency (see Fig 3.1)



Figure 3.1 Comparison of measured and simulated yields for the 1978/80 and 1979/80 growth seasons.

Regrettably no serial harvesting was undertaken so it was not possible to test the growth pattern simulated by the model through the season.

These comparisons however does suggest reliability of the PUTU 14 model for simulating yield for D. eriantha. Significantly, it appears to take into account the affect of natural physical environment (particular water see Section 2.7.1) and nitrogen. The nitrogen balance model incorporated into the PUTU 14 model seems to account for nitrogen balance influences, both in the soil as well as in the plant.

N-distribution

Results of measured and simulated yields at 30, 90 and 150 kg N ha⁻¹ obtained during the 1978/1979 growing season for a Westley soil are given Fig 2.12. The decided limitation due to water stress at high nitrogen levels is apparent. Results from the 1979/1980 growing season are shown in Fig 3.2. By contrast measured yields obtained with 30, 90 and 150 kg applied nitrogen ha⁻¹ showed a steady increase from 1456, through 3302 to 5167 kg ha⁻¹ (see Quadrant C; Fig 3.2). The model predicted this change in biomass production well, giving 1484, 3437 and 4930 kg ha⁻¹ (see Quadrant B; Fig 3.2). Nitrogen limitation still prevailed even when 150 kg nitrogen ha⁻¹ was applied, as simulated and measured yields were still increasing almost linearly (Quadrant A; Fig 3.2) with applied nitrogen. A tendency to simulate moderate nitrogen stress is apparent from the slight curve in the simulation curve in Quadrant C, Fig 3.2.

Validation on all soils and seasons

Model performance was studied under conditions prevailing on the Western Transvaal of Southern Africa, using the results provided by Dannhauser (1985) and Dannhauser <u>et al</u> (1987).

Effective rooting depth was taken as 1200 mm and 1000 mm for the Avalon and Valsrivier soil, respectively, while clay content of A and B horizon was taken as 15 %



Figure 3.2 Simulated nitrogen applied, simulated N-uptake, simulated production and measured final yields at three N-levels for the 1979/80 growing season at Potchefstroom.

60

and 35 %. Initial soil water conditions in 1981/82 were assumed the same for both soils at 19.7 and 23.5 % for the two soil layers respectively. In subsequent seasons the soil water content as simulated at the end of the previous season was carried over to the next. The statistical analyses comparing model output to measured yield on three soil types is given in Table 3.2 and Fig 3.3; 3.4 and 3.5.

Field studies, under dryland conditions in respect of nitrate uptake, nitrification, humufication, nitrogen leaching etc, were not undertaken. Thus, it was not possible to test any of the soil nitrogen outputs obtained from the model.

Table 3.2Model performance and test for Westley (n=3), Avalon (n=6) and
Valsrivier (n=6) soils at various N-levels and data from all three lumped
together (n=15). MAE = mean absolute error, RMSE = root mean
square error, RMSE_s = systematic error, RMSE_u = unsystematic error,
D = Wilmott index of agreement and r^2 = coefficient of determination.

Soil Type	n	MAE t ha ⁻¹	RMSE t ha ⁻¹	RMSEs t ha ⁻¹	RMSEu t ha ⁻¹	D	r ²	Slope through origin
Westley	3	0.23	0.15	0.11	0.11	0.99	0.99	0.98
Avalon	6	0.89	0.51	0.02	0.51	0.96	0.85	0.98
Valsrivier	6	0.43	0.72	0.58	0.41	0.94	0.89	0.78
Lumped	15	0.35	0.56	0.26	0.50	0.96	0.88	0.90

The resullts attained with the lumped data reflect an accurate validation of the PUTU 14 model. The MAE is low 0.35 t ha⁻¹; as are RMSE, RMSE_s and RMSE_u. The coefficient of determination ($r^2 = 0.88$) is highly acceptable.

The lumped data (n = 15) from all three soils are presented in the scatter diagram in Fig 3.3.



Figure 3.3 Comparison of measured and simulated yield attained for a Westley, Avalon and Valsrivier soil.

Westley soil

Evidence of good agreement between measured and simulated yield is given by MAE and RMSE of less than 0.23 t ha⁻¹ for the Westley soil. The RMSE_s and RMSE_u are small by modelling standards and nearly equal, indicating good model performance with little systematic error.

Both the Wilmott index of agreement (D) and the coefficient of determination (r^2) approach unity for the Westley soil because the data sets are few (n = 3).

Avalon soil

Model estimation between measured and simulated yield are less than 0.65 t ha⁻¹, indicating a poor fit. Both the Wilmott index of agreement (D) and the coefficient of determination (r^2) are 0.80, or higher. Results from Fig 3.4 depict poor prediction of yields at 60 kg N ha⁻¹ in only the 1981/82 season.



Figure 3.4 Measured and simulated yield for Avalon soil at different soil N-levels for 1981/82, 1982/83 and 1983/84 season.

Valsrivier soil

Model performance tests indicated a reasonable to poor model fit in these conditions. The MAE and RMSE of 0.43 t ha⁻¹ and 0.72 t ha⁻¹ approach acceptability. Both the r^2 and D value were higher than 0.89. Once again the agreement (see Fig 3.5) was adversely affected by only the 60 kg N ha⁻¹ treatment in the 1981/82 season.



Figure 3.5 Measured and simulated yields on a Valsrivier soil at different soil Nlevels for 1981/82, 1982/83 and 1983/84 season.

To conclude, it appears as though the soil N-balance yields were poorly modelled during the 1981/82 seasons at N-application rates of 60 kg N ha⁻¹. This was probably due to using too low soil N-levels at the start of the season (7.5 kg N ha⁻¹). In Fact, when a higher value of 120 kg N ha⁻¹ was subsequently substituted, this discrepancy was eliminated. In this regard, Rethman (1987) has indicated the differences in dry matter yield in *D. eriantha* attainable in seasons subsequent to seasons fertilized at different levels of nitrogen. These quantities are essential when modelling the nitrogen balance is undertaken.

Although phosphorus was applied, it appeared in any event to have been optimal. It is important to quantify low levels of phosphorus as this could seriously reduce growth rate.

CHAPTER 4

MODEL APPLICATIONS

4.1 Introduction

Nitrogen application rates need to be chosen according to expected seasonal plant growth, which is primarily determined by rainfall and to a lesser extent existing soil nitrate levels. Due to the unpredictability and variability in rainfall (see Fig 4.1 for Potchefstroom), there is always a certain amount of climatic risk involved in applying nitrate. In poor rainfall years over application occurs and in good rainy seasons under application.



Figure 4.1 Variation in annual rainfall for period 1916 to 1990 at Potchefstroom.

To diminish this risk, solutions are required for questions such as:

- 1. What is the influence of climatic regime upon yield response to both level and time of nitrogen application ?
- 2. How may climatic risk at different nitrogen application rates be quantified ?

Both these questions deal with the optimization of nitrogen application rate and climatic risk.

Five growing season rainfall scenarios were selected, viz. bad, poor, moderate, good and wet, in which to assess the influence of timing and level of N-application. These scenarios were defined as periods receiving monthly and seasonal rainfall totals not exceeding what has occurred 10; 30; 50; 70 or 90 percent of the time in the past.

Composite rainfall scenarios

In order to investigate the above two questions, it was necessary to create the five rainfall scenarios bad through wet as defined above. The method of Fouché (1992) was followed. Herein cumulative probability functions of rainfall were established for each month of the year. From these it was possible to extract the monthly totals of rainfall not exceeded 10, 30, 50, 70 or 90 % of the time. These totals are given in Table 4.1. The years in which these totals occurred appear in Table 4.2. Taking the daily rainfall values for these particular months, composite years were constructed with expected monthly and seasonal totals not exceeded 10, 30, 50, 70 or 90 % of the time. Data from these five composites were then used for running the model to estimate yields for bad through wet scenarios.

Table 4.1Monthly total rainfall (mm) with the given chances of non-exceedance(cumulative probability) for Potchefstroom.

Scenarios	Cumulative		Month											
	Probability %	Jul	Aug	Sep	Oct	Nov	Dec	Jan	Feb	Mar	Apr	May	Jun	Total
Bad	10	0	0	0	15.4	24.2	47.8	50.8	45.4	26.1	4.6	0.0	0.0	214.3
Poor [.]	30	0	0	3.4	29.9	62.6	72.1	89.4	71.0	62.0	21.8	5.3	0.0	417.5
Moderate	50	1.7	1.8	9.5	48.8	90.8	102.6	115.6	88.3	80.6	40.9	11.4	3.0	595.0
Good	70	7.4	11.3	31.6	70.8	110.5	125.8	143.6	128.6	116.8	84.1	33.4	11.9	875.8
Wet	90	32.5	30.7	58.3	106.4	149.7	163.8	188.4	179.5	167.7	105.8	54.8	19.4	1257.2

Table 4.2Years in which the monthly rainfall totals given in Table 4.1 occurred at
Potchefstroom.

Scenarios	Cumulative		Month										
	Probability %	Jul	Aug	Sep	Oct	Nov	Dec	Jan	Feb	Mar	Apr	May	Jun
Bad	10	1935	1933	1946	1968	1950	1957	1960	1971	1940	1920	1991	1936
Poor	30	1974	1970	1963	1931	1988	1988	1958	1918	1989	1939	1921	1986
Moderate	50	1942	1983	1984	1928	1976	1941	1945	1929	1985	1922	1970	1975
Good	70 .	1923	1984	1921	1964	1928	1969	1957	1974	1928	1924	1978	1949
Wet	90	1952	1943	1979	1982	1980	1973	1976	1959	1944	1942	1962	1925

Three dates for applying nitrogen were investigated for each application level. These dates were: 1st September (1), 1st November (2) and 1st January (3).

Table 4.3Rate of N-application.

Scenarios	Nitrogen applied (kg ha ⁻¹)
Bad	5,10,15.20,25
Poor	15,30,45,60,75
Moderate	25,50,75,100,125
Good	35,70,105,140,170
Wet	60,120,180.240,300

4.2 Interaction between climate and nitrogen application

The PUTU 14 model was used to simulate expected yields for the five different climatic regimes, bad through wet at each of the application rates detailed in Table 4.3. Results for each of these simulations are given in Appendix D(1-5). Fig 4.2 (A-E) illustrate the influence of these various climatic regimes and N-application rates upon yield.





A. Bad





C. Moderate





Figure 4.2 Influence of various climatic regimes and N-application rate on yield. The bars indicate the range in yield obtained as a result of different time of application.





Figure 4.3 Influence of various climatic regimes and time of N-application on yield. The bars indicate the range in yield obtained as a result of the different levels of applied nitrogen.

From these figures optimum nitrate levels for each of the five scenarios could be established. These were for optimum yields of 0.9; 2.0; 3.0; 4.7 and 7.0 t ha⁻¹, respectively.

As far as timing of N-application is concerned (Fig 4.3), it is evident that the late (3) Napplications, restricted growth, irrespective of the type of year experienced. There was however a wide range in simulated yields as the level of N-application changed (see bars in Fig 4.3). This implies that the time of N-application is dependent upon the pattern of vegetative growth. An early N-application could result in a loss of N either through leaching or an inability of the plant to immobilize N for growth throughout the rest of the growing season. It appears that a mid-season application (2) could prove to be the best time for applying nitrogen. By comparison, Dannhauser (1988) found a Napplication during December to January to be the best.

4.3 Optimization of nitrogen application and climatic risk

Climatic risk is here defined (de Jager, pers. comm.) as the chance of non-realisation of a given yield due to the weather conditions prevailing during the growing season. It is evident that this is equivalent to the cumulative probability of occurrence of a given yield and is obtainable from the cumulative distribution function, CDF (see de Jager and Singels 1990).

Simulations were carried out using actual weather data for the period 1916-1991 at various soil-N levels (0, 30, 60, 90, 120 and 150 kg N ha⁻¹ annum⁻¹). The results for Potchefstroom are given in Appendix D.6. In order to calculate the level of climatic risk, (cumulative probability of yields at different soil N levels) the techniques of Doll & Orazem (1984) are required. For first order stochastic dominance (SD) a cumulative distribution function (CDF) lying to the right of another will reflect less climatic risk than the other.

Because grassland pastures utilize (in vegetative growth) virtually every drop of water received, the CDF in Fig 4.4 are the same at low yield levels. This is an interesting phenomenon peculiar to grassland perhaps not previously appreciated.



Figure 4.4 Climatic risk — , the cumulative distribution function of simulated yield for different soil N-levels.

Simulated yields with increasing levels of N from Fig 4.4 and Table 4.4 clearly indicate that :

- 1. As might be expected, climatic risk is decreased, with increased Nfertilization, and of course, so would gross margin as well, and
- 2. Higher standard deviation and range between high and low simulated yields, but lower coefficients of variation, occur.
- Table 4.4Average, Median, Standard deviation, Low and High values simulated forPotchefstroom from 1916-1991, at different N-levels.

Description	N(0)	N(30)	N(60)	N(90)	N(120)	N(150)	
Average	469.7	1433.7	2029.5	2266.9	2363.7	2389.9	
Median	435.3	1621.8	2100.9	2100.9	2100.9	2100.9	
Standard deviation	145.9	458.2	952.3	1273.0	1458.0	1523.2	
Low	243.9	268.0	268.0	268.0	268.0	268.0	
High	1041.4	2228.9	3710.7	4863.3	5830.8	6705.6	

From Table 4.4 it is also evident that except for levels of 30 and 60 kg N ha⁻¹ applied, the median yields are always below average simulated yields, indicating that less years occur with above than below average yield. Furthermore for a probability of 0.5, that is 1 out of 2 years a yield of about 2.2 t ha⁻¹ could be expected with an application of 60 kg N ha⁻¹ (see Fig 4.4 and Fig 4.5).



Figure 4.5 The expected outcome-variance space - A. Plot of mean yield against standard deviation at differing N-applications. A 2:1 line is also indicated. Data for all 76 seasons was included.

It is a simple matter to construct an expected yield-variance diagram (E-V space), where variance is quantified using the standard deviation of yields obtained at the various N-levels. Such E-V space is illustrated in Fig 4.5. Wafula, McCown & Keating (1992) suggest that as a rule of thumb, risk averse entrepreneurs will prefer strategies found on the 2:1 line. In Potchefstroom this would be 60 and 30 kg N ha⁻¹ with preference for the former.

Lastly, a parameter for estimating possible improved yield (P) at a given N-application is defined as:

The chance of realising an improved yield above the yield corresponding to that attained with a lower N-application rate.

Thus, where:

n - Total number with simulated yields (n = 75 seasons)

 N_h - Number of seasons in which an increased yield would result were a higher N-application used.

$$P = \frac{N_h}{n} .$$

From Fig 4.6, the highest P occurred when N-application was increased from 0 - 30 kg ha⁻¹ (P = 0.32) which was virtually the same for an increment from 30 - 60 kg ha⁻¹ (P = 0.27). Thereafter the P = 0.11 for each subsequent increment in the level of N.



Figure 4.6 Chance of realising an increased yield were a higher N-application utilized (△) and the mean yield (*) at given nitrogen application rates. The bars indicate the range (maximum - minimum) in yields due to seasonal weather variability.

CHAPTER 5

SUMMARY AND CONCLUSIONS

5.1 Introduction

The need for a simulation model for cultivated pasture, particularly *Digitaria eriantha*, for the RSA has become urgent. Furthermore, since previous grassland simulation models have not included soil nitrogen subroutines. This too has become a priority. With a view to producing such model and refining former simulation exercises; numerous key questions have been identified. For example, would it be possible to use the PUTU 13 type of model for *Cenchrus ciliarus* as a basis for developing a new model for *Digitaria eriantha*, would it be possible to include a nitrogen subroutine into such model and apply it to determine efficient fertilizer strategies (how much and when nitrogen should be applied).

Objectives

Bearing this in mind, the objectives of the present study were to :

- adapt the PUTU 13 model for simulation of the growth and development of Digitaria eriantha.
- Include a nitrogen subroutine into this model and validate it, and
- Demonstrate the suitability of the new model for determining nitrogen strategies given a long series of weather data.
- 5.2 Method

General model development

The work was carried out in the Potchefstroom area, using yields of *D. eriantha* collected by Dannhauser (1985). A literature survey was included, where applicable in

75

the body of the thesis, as this facilitated model description. A detailed model description was undertaken which outlined how the PUTU 2, 11 and 13 models were refined to produce the new model PUTU 14 for *D. eriantha*.

The changes to these earlier models were highlighted. The entire functioning of the model was carefully explained. The source of the equations utilized was given, thereby meeting an urgent need of scientists interested in developing the model further.

New mathematical functions developed, included;

- Use of a constant leaf development rate in place of the rapid and slow phases used in the previous grassland models.
- Expressing photosynthetic rate in terms of the product of radiationphotosynthesis conversion coefficient and incident radiant flux density.
- Where the previous grassland models considered 100% basal cover which was later graded down according to a basal cover function; the new PUTU 14 has no such option. Where cultivated pastures are concerned a uniform cover is usually established and such operation is not nescesary.
- Where previously translocation was based on a source-sink relationship; in the new model a constant proportion of daily dry matter assimilation is partitioned to each plant organ.

The growth processes simulated in each of five growth stages consisted of carbon dioxide assimilation, translocation, senescence and death. The five growth stages defined were vegetative, reproductive, seed, seed fall and dormancy.

Water and carbohydrate mass balances are computed in each growth stage. The water balance remained virtually unchanged from previous PUTU models, use being made of the finite reservoir cascade technique. This process is clearly described in the text. An innovation is a new exponential function for the water retention curve. It is calculated using a modified Campbell (1977) equation which is now common in all PUTU models.

Modelling soil nitrogen

The nitrogen subroutine was taken from Singels & Manley (1991). Briefly this entails:

The computation of the fresh organic matter decomposition rate which is a function of an empirical maximum rate, temperature, soil water status and the C:N ratio. A modified water status function is included in the new model. The mineralisation rate of organic matter is taken as a function of a given empirical decay rate, temperature, water and nitrogen content in the humic fraction of the soil. Nitrogen immobilization is calculated similarly to the above, but at a rate approximately 2% thereof. The nitrogen balance is computed according to a logarithmic theory of van Keulen & Seligman (1987). The calculations accommodate losses due to leaching.

Modelling Plant nitrogen

In the plant, N-uptake is regulated according to the law of the minimum. Nitrogen flow is proportional to transpiration flow, a potential uptake rate which is a function of the mass in each relevant organ and a rate of demand. Nitrogen mass balance is calculated for each organ on a daily basis.

5.3 Calibration

Calibration was conducted on three sets (different N-levels) of data collected in the 1978/79 season. The major input parameter changes from values applicable to PUTU 11 and 13 were the following:

The base for calculating thermal period, which was changed from 12°C to 10°C.
The minimum decrement in thermal period accounting for adaptation to water

stress was changed from 230 d°C to 225 d°C.

- The duration of the reproductive period was altered from 60 to 30 days and the seed

formation period from 100 to 80 days.

- The minimum temperature for growth of 2° C seems as applicable to *D. eriantha* as it is to the former species.
- Furthermore, the rate constants (proportions) for partitioning of dry matter were determined for *D. eriantha* for all growth stages.
- The constant in the argument of the exponential water stress factor was altered from its previous value of 0.5 to the 0.1 used in PUTU 14.

5.4 Validation

The model was validated on data collected in 1979/80 season and the 1981/82, 82/83 and 83/84 seasons. A total of 15 sets of data were available for three different soil types. The significant tests of accuracy undertaken were the Wilmott coefficient of agreement (D), the coefficient of determination r^2 , mean absolute error (MAE), the root mean square error (RMSE), systematic (RMSE_s) and unsystematic (RMSE_u).

Values obtained during validation were

D = 0.96; $r^2 = 0.88$; MAE = 0.35; RMSE = 0.56; RMSE_s = 0.26 and RMSE_u = 0.50.

These testify to an accurate model.

5.5 Application of the model

The model was run on 75 years of weather data for the period 1916 to 1991 at Potchefstroom on a Westley soil. Seasonal yields were calculated for each growing season and used to determine the most suitable nitrogen strategy for Potchefstroom. The optimum N-level obtained from these data for each of the seasonal scenarios defined as bad, poor, moderate, good and wet were determined. The most favourable time of application was found to be 1 November. It was further demonstrated how N- applications could diminish climatic risk. Furthermore, use was made of the expected yield-variance space from which it was found that a risk averse entrepreneur would prefer a nitrogen application of 60 kg/ha.

A new parameter for estimating the chance of an improved yield resulting from an incremented nitrogen application was defined. This potential for improving yield of a given N-application (P) was evaluated. The highest P occurred when N-application was increased from 0 - 30 kg ha⁻¹ (P = 0.32) and was virtually the same for an increment from 30 - 60 kg ha⁻¹ (P = 0.27). Thereafter P = 0.11 approximated for each subsequent level of N.

The variability in yield due to weather fluctuations at each N-level was determined and the highest expected variation was found to occur at N-applications of 180 kg ha⁻¹. The potential yield at Potchefstroom for each nitrogen application was also determined. This was taken as the median yield. It varied between 1.14 and 2.1 t ha⁻¹ according to the level of nitrogen fertilization. Whereas, median yield ceased to improve with Napplications greater than 60 kg N ha⁻¹, average yields did reflect a slight increase at higher N-levels.

While the analysis was expressed in terms of yield, the results could very easily be expressed in terms of gross margin which would have great economic significance.

5.6 Conclusion

It may be concluded that the functioning of a new model for *D. eriantha* (PUTU 14), which contains a nitrogen sub-routine, how it was developed and how it differs from the previous grassland models upon which it was based, has been fully described. The model was calibrated and then validated on independent data.

The model was applied to calculating the potential (median) yield for Potchefstroom at different levels of nitrogen and the variation therein brought about by variability in climate.

The model was applied to demonstrate how climatic risk may be diminished by increased nitrogen fertilization.

The potential for improving yield by incrementing nitrogen applied level was also determined.

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APPENDIX A

Source code for the PUTU 14 model (Digitaria eriantha)

- 1 DECLARE SUB PLANTN (J, BRL, BCL, BBL, BGL, BRD, BCD, BBD, BGD, TRANS1, TRANS2, ANC(), AN(), V(), V15(), NO3(), NH4(), TAVE, FW, N(), BO, FACN, GRSTGE, TOTMASS(), NDEM(), NCMAX(), NC(), TOTAN, TPAN, TNFLO, TNMASFLO, TOTNUP)
- 2 DECLARE SUB SOILN (J, BRL, BCL, BBL, BGL, BRD, BCD, BBD, BGD, TRANS1, TRANS2, V(), V01(), V15(), ANC(), NO3(), NH4(), W01(), W(), IFOM(), FOM(), T(), INFIL, PERC(), DoyToFert(), FERTO3(), FERTH4(), TAVE, GWP, N(), BO, FACN, GRSTGE, TOTMASS(), NFERT, FTMIN, FWMIN, CNR, FCNR, DECR, RDECR(), NMINFOM(), NFOM(), NIMMFOM(), NMINHUM(), NHUM(), DMINR, NMINNET(), HUM(), NIT(), NOUTFLO(), NLEACH, AN(), NDEM(), NCMAX(), NC(), TOTAN, TPAN, TNFLO, TNMASFLO)

3 NA\$ = "C:\QB45\WEER\PCH79.TXT"

PROGRAM PUTU 14 DIGITARIA ERIANTHA MODEL **JUNE 1993** VERSTON 1 DIM TP(300), PRVISC(300), PPRODO(300), MAXPROD(300), CNT(370) DIM D(300), DATE(300), TPCUT(100), RTRIM(370), DALEN(13), DAE(13) 4 5 DIM SLWAT1(367), SLWAT2(367), SLWAT(367) 6 DIM ANC(2), NO3(2), NH4(2), IFOM(2), FOM(2), T(2) DIM DoyFert(2), FERTO3(2), FERTH4(2) DIM RDECR(2), NMINFOM(2), NFOM(2), NIMMFOM(2), NMINHUM(2), NHUM(2) 7 8 9 10 DIM NMINNET(2), HUM(2), NIT(2), NOUTFLO(2) 11 DIM AN(2), NDEM(4), NCMAX(4), NC(4), N(4)12 CLS 13 DATA 10.5,11.1,11.9,12.8,13.6,14.0,13.8,13.1,12.3,11.4,10.7,10.3,10.3 14 DATA 31, 31, 30, 31, 30, 31, 31, 28, 29, 31, 30, 31, 30 15 FOR i = 1 TO 13 16 READ DALEN(i) 17 NEXT i 18 FOR i = 1 TO 13 19 READ DAE(i) 20 NEXT i / ********** STAGE\$(1) = "VEGETATIVE" 21 **STAGE\$(2) = "REPRODUCTIVE"** 22 STAGE\$(3) = "SEED" 23 STAGE\$(4) = "SEEDFALL" 24 STAGE\$(5) = "DORMANT"25 / ***** ***** INPUT VALUES 26 CLAY = 25: SDP = 300WHC1 = (.138 + .00416 * CLAY) * 10027 WHC4 = (.0344 + .00381 * CLAY) * 10028 29 SLWP1(1) = 19.7: SLWP2(1) = 23.57 : WPC = -180030 TPMAKS = 400: PRVISC = 150: PPRODO = 500 : BCOVER = 10

89

31 32 33 34	CONS = 0.5 MKOUNT = 39 HYCON = .07 GRSTGE = 19	:ALFA = 4 5: HUCRIT 1: RUNPAR : ITERM =	4 :EFFQ = (= 250: HU(= .1: DRA 1: JDMAKS	0.775 :RCC CRMN = 230 (NP = .9 = 0: TNEX	C = ALFA *): BO = 10 (T = 1: TF	EFFQ : SPL = = 1000	500
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,	KEY:						
,	DMG=TH	IE DAYS DF	Y MATTER O	GAIN (KG C	ARBOH./HA	/DAY)	
,	GD =GF	RAIN DEAD		GL =GRAI	N LIVE		
,	CD =CU	ILM DEAD		CL =CLUM			
	BD = LE	CAVES DEAL) D	BL =LEAV	ES LIVE		
,	נפ= תפ עם– תפ	.UDDLE UEA 1075 DFAD		ВТ =2.10В Э⊓ =2.10В	905 DIVE 93 TIVE		
,	TR =TR	ASH DEAD		RES=CARR	OHYDRATE	RESERVES	
,	DA =AE	SOVE GROUN	ID DEAD	BA =ABOV	E GROUND	BIOMASS	
,	DB =RC	OT DEAD	-	BB =ROOT	BIOMASS		
,	D=DA+D	B=TOTAL C	EAD	B=BA+BB=	TOTAL STA	NDING BI	OMASS
,	CA=ABC	VE GROUND	STANDING	CROP			
,	CB=BEI	JOW GROUND	STANDING	CROP			

================== VARTABLE NAME CODE UNITS ================== ====== ==== BASAL COVER BCOVER (%) DISIRED PROPORTION OF RELEVANT ORGAN DPRO AVERAGE TEMP т (') (') (') TEMP MAX AMX TEMP MIN AMN (W/M**2) RADIANT FLUX DENSITY RFD WATER POTENTIAL OF LEAF LWP (K PA) (M**2) LEAF AREA AT. SPECIFIC LEAF AREA SPL (KG/HA) ITERM PERIOD OVER WHICH MEAN DETERMINED ITERM ***** 35 WHCTEMP = 0/ ** SET INITIAL STATUSSES AND PARAMETERS ** ' READ SOIL WATER AND CUT INFO ## IF CUTS ARE READ, IL = 1,15 IN DO AND PUTUINFO ## IF WHCTEMP <> 0 THEN WHC = WHCTEMP 36 37 CUT = CUT1' COMPUTE SOIL DEPTH FOR SECOND LEVEL ## 38 SDP1 = 100SDP2 = SDP - SDP139 ' COMPUTE FC AND PWP FOR EACH LEVEL ## FC1 = WHC1 / 100 * SDP1 FC2 = WHC1 / 100 * SDP2 40 41 PWP1 = WHC4 / 100 * SDP142 43 PWP2 = WHC4 / 100 * SDP244 M1 = (LOG(10) - LOG(1500)) / (LOG(WHC1) - LOG(WHC4))WATER CONTENT EXPRESSED IN MM/SOIL DEPTH FOR BOTH LEVELS ## 45 WJ = SLWP2(1)SLWAT(1) = (SLWP1(1) * SDP1 + SLWP2(1) * SDP2) / SDP / 100 * SDP SLWAT1(1) = SLWP1(1) / 100 * SDP1 SLWAT2(1) = SLWP2(1) / 100 * SDP2 46 47 48 DETERMINE INITIAL MASSES OF THE DIFFERENT PLANT COMPONENTS * C IS TOTAL PLANT BIOMASS AT 100 % BASAL COVER ## ALL CALCULATIONS WILL PROCEED WITH 100 % BASAL COVER ## ON COMPLETION BIOMASSES WILL BE MULTIPLIED BY ## ## BY BCOVER*BCOVER FORMULA (BCFUNC) 1 THE ASSUMPTION IS MADE THAT THE PREVIOUS PRODUCTION IS ## EQUAL TO 25 % OF THE TOTAL BIOMASS ## 49 C = PRVISC * 100 / 25 * 100 / BCOVERBCFUNC = BCOVER / 10050 51 BL = 0: BD = 0 * C: CL = 0: CD = 0 * C52 SL = 0: SD = .1 * C:RL = .8 * C: RD = .1 * C53 GL = 0: GD = 0: RES = .1 * RL54 55 TPMAKS = 056 CLIVE = BL + SL + RLSUB FOR NITROGEN APPLICATION 57 GOSUB 500
START OF SIMULATION PER DAY FOR THE GROWING SEASON * 58 T = 159 **OPEN NA\$ FOR INPUT AS #2** 60 DO UNTIL EOF(2) INPUT #2, JUNK\$ 61 JAAR = VAL(MID\$(JUNK\$, 1)) MNDE\$ = MID\$(JUNK\$, 5, 5) DAY = VAL(MID\$(JUNK\$, 10, 3)) 62 63 64 AMXT = VAL(MID\$(JUNK\$, 14, 8))65 AMNT = VAL(MID\$(JUNK\$, 21, 6))66 $\begin{array}{l} \text{RAINF} = \text{VAL}(\text{MID}(\text{JUNK}, 28, 6)) \\ \text{EVAP} = \text{VAL}(\text{MID}(\text{JUNK}, 34, 6)) \\ \text{SUN} = \text{VAL}(\text{MID}(\text{JUNK}, 40, 6)) \\ \end{array}$ 67 68 69 NMNTH = J / 30 + 170 HERE AL=LA PER HECTARE LEAF SPL = 500 kg/ha ## AL = BL / SPL71 72 IF C <> 0 THEN 73 FRACL = BL / C74 END IF ***** * WATER BALANCE * ***** 75 GOSUB 200 ' SUB WATER ROUTINE ****** * ENVIRONMENTAL FACTORS AND * * PRODUCTION ****** 'TO COMPUTE THE INFLUENCE OF SOLAR RADIATION ON THE MAX RATE OF ## 'PHOTOSYNTHESIS(FI) 'FI = PERCENTAGE SUNLIGHT ABSORBED BY THE CANOPY ## 76 $\mathbf{TRFD} = \mathbf{TRFD} + \mathbf{RFDM}$ 77 FI = (1 - EXP(-.7 * AL)) * 10078 TFI = TFI + FI' TO COMPUTE THE INFLUENCE OF TEMP ON THE MAX RATE OF ## ' PHOTOSYNTHESIS ## (FT) 79 $FT = 100 * (EXP(-1 * ((ATEMP - 30) ^ 2) / 360))$ 80 IF ATEMP < BO THEN FT = 0 81 IF AMNT < 3 THEN FT = 0 82 $\mathbf{TFT} = \mathbf{TFT} + \mathbf{FT}$ TO COMPUTE THE INFLUENCE OF WATER AVAILABILITY ON THE MAX ## ' RATE OF PHOTOSYNTHESIS (FW) ## 83 $FW = FW \times 100$ 84 TFW = TFW + FW' SHOULD SOIL BE SATURATED REDUCE PHOTOSYNTHETIC RATE BY 20 % ## 85 IF SLWAT2(J) \rightarrow FC2 THEN FW = 100 - 10 * (SLWAT2(J) - FC2) / SDP2 86 IF FW $\langle = 0$ THEN FW = 0 ' COMPUTE BEGINING AND END, AS WELL AS NUMBER OF MOISTURE STRESS DAYS 87 IF GRSTGE < 5 THEN GOTO 6 ELSE GOTO 7 89 6 IF FW < 50 AND ATEMP > BO THEN ITTRE = ITTRE + 1 90 IF FW \langle 50 AND ATEMP \rangle BO THEN STRESS = .47 ELSE STRESS = 1

91	' WPC = CRITICAL LEAF WATER POTENTIAL IF FW < 50 AND ATEMP > BO THEN WPC = WPC - 15:	
92 93	IF WPC <= -3000 THEN WPC = -3000 WPC = -1800	
94	' IF FW < 50 AND ATEMP > BO THEN PRINT "STRESS DAY ON"; J, ITT 7 'CONTINUE	RE, FW
	' THE INFLUENCE OF ENVIRONMENT ON THE EFFICIENCY OF PHOTO- ' SYNTHESIS = F	## ##
95 96	F = (FI / 100) * (FT / 100) * (FW / 100) * 100 TF = TF + F	
	'ACTUAL EFFICIENCY OF PHOTOSYNTHESIS = EFF	##
97 98	EFF = F / 100 TEFF = TEFF + EFF	
	'THE POTENTIAL OF CO2-ASSIMALATION = P (KG CO2/HA/DAY)	##
99 100 101 102 103	IF RFDM >= 10 THEN GOTO 10 P = 0 GOTO 11 10 $P = RCC * RFD * (F / 100))$ 11 $P = P * 10$	
	' NETTO DRY MATTER GAIN	
104 105	DMG = P * FACN $TDMG = TDMG + DMG$	
	<pre> / ************************************</pre>	
106 107 108 109 110 111 112 113 114 115 116 117	SELECT CASE GRSTGE CASE IS = 1 GOTO 13 CASE IS = 2 GOTO 25 CASE IS = 3 GOTO 30 CASE IS = 4 GOTO 35 CASE IS = 5 GOTO 37 END SELECT	-
	<pre>' ************************************</pre>	*## ### ### ### ### ###
118	13	

' NO GROWTH BEFORE PHOTOPERIOID = DAY 31 (31st July)

119 IF J < 30 THEN 38

120 IF HU > HUCRIT THEN GOTO 24 ELSE GOTO 17

SHOULD GROWTH STAGE 1 LAST TO LONG (BEYOND 258 DAYS) ## SWITCH TO TNEXT GROWTH STAGE ## AND IF TEMP IS TO LOW TERMINATE GROWTH ## 121 17 IF J > 258 THEN GOTO 18 ELSE GOTO 19 18 IF AMNT <= 2 THEN GOTO 23 122 , BCON=KG/HA/DAY/DEGREE CELSIUS AT 100% COVER ## 19 BCON = 3.5123 124 TGROW = ATEMPIF TGROW > = 30 THEN TGROW = 30 125 IF TGROW <= BO THEN TGROW = BO 126 HU = HU + (TGROW - BO) HUCRIT = HUCRIT - 10 * (100 - FW) / 100 127 128 IF HUCRIT <= HUCRMN THEN HUCRIT = HUCRMN 129 ' CALCULATION OF LEAVE MASS CHANGE (KG/HA/DAY) ## DBL = BCON * (TGROW - BO) * FW / 100130 20 131 BBL = BL'CALCULATE MASS FLOW VARIABLES FOR MASS FLOW FROM LIVING PARTS ## ' TO DEAD PARTS 132 X(1) = .001 * SL133 X(2) = .001 * BLX(3) = .001 * RL134 135 X(8) = 0136 X(4) = 0137 GOTO 38 'PRINT "DAY="; JDA; "MIN.TEMP.="; AMNT; "<<<<>>>> THEREFORE 138 23 TERMINATE GROWTH" 139 24 ' PRINT STAGE\$(2); " - GROWTHSTAGE 2"; JDA ' TRANSLOCATION RATES ARE PROPORTIONAL TO DIFFERENCES ## BETWEEN EXISTING AND DESIRED ORGAN PROPORTIONS ## ' DESIRED PROPORTIONS = DPRO ...FOR REPRODUCTIVE GROWTH STAGE ## 140 DPROG = 0DPROC = .2141 142 DPROB = .3143 DPROS = .1DPROR = .4144 145 GRSTGE = 2146 MKOUNT = J + 30, , GROWTH STAGE TWO ## SECOND GROWTH STAGE IS REPRODUCTIVE GROWTH. TRIGGER FOR ## CHANGE TO 3RD GROWTH STAGE IS 30 DAYS OF GROWTH WITH ## A MINIMUM CULM PRODUCTION EXCEEDING 25 % OF THE MASS OF ## LEAVES AT A THEORETICAL BASAL COVER OF 100 % ## 147 25 148 IF CL > (.25 * BL) AND J > MKOUNT THEN GOTO 29 149 IF $J \ge 316$ THEN GOTO 26 150 GOTO 27 151 26 IF AMNT <= 2 THEN GOTO 28 ' CALCULATE THE MASS FLOW OF LIVING PLANT MATERIAAL TO DEAD ## 152 X(1) = .001 * SL27 X(2) = .001 * BLX(3) = .001 * RL153 154

155 **GOTO 38** PRINT "DAY = "; JDA; " MIN. TEMP. = "; AMNT; "<<<<>>>> 28 156 THEREFORE TERMINATE GROWTH" 29 PRINT STAGE\$(3); " - GROWTHSTAGE 3"; JDA 157 ' DESIRED PROPORTIONS = DPRO ...FOR SEED GROWTH STAGE ## 158 DPROG = .09DPROC = .34159 160 DPROB = .17DPROS = .2161 DPROR = .3162 GRSTGE = 3163 MKOUNT = J + 80164 IKOUNT = MKOUNT - 30165 ## **GROWTH STAGE THREE** ## , ## , THIRD GROWTH STAGE IS SEED. TRIGGER FOR 4TH GROWTH STAGE ## , IS 80 DAYS OF SEED FORMATION, OR A GROWING SEASON STRETCHING BEYOND DAY J = 316 AND TEMP < 2 C TERMINATE ## ## GROWTH ## 166 30 IF J > MKOUNT THEN GOTO 34 167 168 IF $J \rightarrow = 316$ THEN GOTO 31 **GOTO 32** 169 170 IF AMNT < 2 THEN GOTO 33 31 X(1) = .002 * SL171 32 X(2) = .002 * BL172 X(3) = .003 * RL173 174 IF $J \rightarrow =$ IKOUNT THEN X(8) = .15 * GL175 X(9) = .001 * CLX(6) = .001 * SD176 X(7) = .001 * BD177 178 **GOTO 38** PRINT "DAY = "; JDA; " 179 MIN. TEMP. = "; AMNT; "<<<<>>>> 33 THEREFORE TERMINATE GROWTH" " - GROWTHSTAGE 4"; JDA 180 34 PRINT STAGE\$(4); ' DESIRED PROPORTIONS = DPRO ...FOR SEEDFALL GROWTH STAGE ## 181 DPROG = 0182 DPROC = .3DPROB = .1183 DPROS = .1184 185 DPROR = .5186 GRSTGE = 4187 GLO = GD + GL + 1, ## GROWTH STAGE FOUR ## 1 ## , FOURTH GROWTH STAGE IS SEEDFALL. THIS PROCEEDS UNTIL A ## A MINIMUM AIR TEMPERATURE < 2 ^OC , ## 188 35 189 IF AMNT < 2 THEN GOTO 36 X(1) = .002 * SL190 191 X(2) = .01 * BL192 X(3) = .003 * RL193 X(4) = 0194 IF (GD + GL) > 0 THEN X(4) = (GLO - (GD + GL)) * .3195 IF $X(4) \rightarrow GD$ THEN X(4) = GD196 IF $X(4) \rightarrow = GD$ THEN GL = 0X(5) = .005 * CD197

198 199 200 201 202	X(6) = .005 * SD X(7) = .005 * BD X(8) = .5 * GL X(9) = .05 * CL GOTO 38	
	' DESIRED PROPORTIONS = DPROFOR DORMANT GROWTH STAGE##' IS THE SAME AS FOR THE SEEDFALL GROWTH STAGE##	
203	36 GRSTGE = 5 ' PRINT STAGE\$(5); " - GROWTHSTAGE 5"; JDA ' Attain the stage for the	
204 205 206 207 208 209 210 211 212 213	37 X(1) = .02 * SL X(2) = .2 * BL X(3) = .003 * RL X(4) = .5 * GD X(5) = .002 * CD X(6) = .003 * SD X(7) = .005 * BD X(8) = .5 * GL X(9) = .25 * CL	
	' DETERMINE WHETHER THE PASTURE BEEN CUT ##	
214 215 216 217 218 219 220 221 222 223	<pre>38 IF JDA <> CUT THEN GOTO 39 TP = BTG + BTC + BTB 'PRINT "A BIOMASS PRODUCTION OF "; TP; " (KG/HA) WAS REACHED CUTTING DATE "; JDA 'PRINT "A BIOMASS PRODUCTION OF "; TP; " (KG/HA) WAS REACHED CUTTING DATE "; JDA GRSTGE = 1 WPC = -1800 TNEXT = 2 HUCRIT = HUCRIT * .8 HUCRMN = HUCRMN * .8 HU = 0</pre>	ON
224 225 226 227 228 229 230 231 232 233 234	IF CUT = CUT3 THEN CUT = CUT4 IF CUT = CUT2 THEN CUT = CUT3 IF CUT = CUT1 THEN CUT = CUT2 IF J > 258 THEN GRSTGE = 5 IF GRSTGE = 5 THEN DPROG = 0 DPROC = 0.3 DPROB = 0.1 DPROS = 0.1 DPROR = 0.5 END IF	
235 236 237 238 239 240 241 242 243 244 245 246	BGL = 0 BGD = 0 BTB = 100 - BTG - BTC BBL = .4 * BTB BCL = .1 * BTB + BTC BCD = .1 * BTB BTC = BCL + BCD BTB = BBL + BBD CL = BCL CD = BCD BL = BBL	

247 248 249	BD = BBD $GL = 0$ $GD = 0$
	<pre></pre>
250	39 GOSUB 300 'TRANSLOCATION
251	IF BL > 0 THEN GOTO 40
252	BL = 0
253	BD = BD
254	40 IF SL > 0 THEN GOTO 41
255	SL = 0
256	SD = SD
257	41 IF RL > 0 THEN GOTO 42
258	RL = 0
259	RD = RD
260	42 IF CL > 0 THEN GOTO 43
261	CL = 0
262	CD = CD
263	43 IF $GL > 0$ THEN GOTO 45
264	GL = 0
265	GD = GD
266	45 TFI = TFI / ITERM
267	TFT = TFT / ITERM
268	TFW = TFW / ITERM
269	TF = TF / ITERM
270	TEFF = TEFF / ITERM
271	TRFD = TRFD / ITERM
272	FI = TFI
273	FT = TFT
274	FW = TFW
275	F = TF
276	ALAI = AL
277	A = J / ITERM
	' CALCULATION OF TOTAL PRODUCTION (TP) FOR EACH DAY
279	BDBL = DBL
280	BRES = RES
281	BGL = GL
282	BGD = GD
283	BCL = CL
284	BCD = CD
285	BBL = BL
286	BBD = BD
288	BSL = SL
389	BSD = SD
290	BRL = RL
291	BRD = RD
292	BTR = TRD
293	BDMG = DMG
294	BP = P
295	BTG = BGL + BGD
296	BTC = BCL + BCD
297	BTB = BBL + BBD
298	BTS = BSL + BSD
299	BTRO = BRL + BRD
300	TP = BTG + BTB + BTC
301	PPROD = PPRODO * $(1 - J / 210)$
302	IF PPROD <= 0 THEN PPROD = 0
303	TPROD = TP + PPROD

304 305 306 307	' DETERMEN MAXIMUM PRODUCTION AND DAY WHEN REACHED IF TP > TPMAKS THEN JDMAKS = JDA TPMAKS = TP END IF	
308	' SET VARIABLES TO ZERO TFI = 0: TFT = 0: TFW = 0: TF = 0: TEFF = 0: TP = 0: TRFD =	0
309	' NEXT DAY J = J + 1	
310 311	' OUPUT TO SCREEN PRINT USING "#########; J; TPMAKS 46 LOOP	
312 313 314 315 316 317	<pre>' VARIABLES AS INPUT FOR NEXT YEAR WHCTEMP = WHC SLWP1 = SLWAT1(J - 1) / SDP1 * 100 SLWP2 = SLWAT2(J - 1) / SDP2 * 100 PRVISC = C * 25 / 100 * BCOVER / 100 IF PRVISC > 2000 THEN PRVISC = 2000 PPRODO = TPROD</pre>	
318	END	
319	200 '***********************************	
320 321 322 323	DRAIN2 = 0 DRAIN1 = 0 TRANS2 = 0 ae = 0	
	<pre>' PROGRAM TO COMPUTE AND TABULATE CROP EVAPORATION AND ## ' WATER BALANCE OVER A PERIOD OF 365 DAYS ON A DAILY ## ' BASIS , DATA OBTAINED FROM WEATHER STATIONS ## ' DATA FORTRAN DESIGNATION ###</pre>	
	AINFALLRAIN(MM)##' RAINFALLRAIN(MM)##' MAX TEMPAMX(C)##' MIN TEMPAMN(C)##' SUNSUN(HOURS)##' EVAPORATION(CLASS-A PAN)EVAP(MM)##' GROUND WATER POTENTIALGWP(KPA)##' SPEC LEAF AREASPL##' SOIL WATERSLWAT(MM/M)##' SOIL WATERSLWAT(MM)##' ACTUAL EVAPORATIONAE(MM)##' SOIL VAPORATIONAE(MM)##' SOIL VAPORATIONSLVAP(MM)##' SOIL WATER LEVEL 1SLVAP(MM)##' SOIL WATER LEVEL 1SLWAT1(MM)##' WATER AVALIBLE AS A % (ESTIMATED VOLUMETRICALY)##' ON DAY ONEWHC(%)I.E. FIELD CAPACITY (FC)	
	AT 244U K PA WHC4 (%) I.E. PERMANENT WILTING PIONT ## P.S. PERMANENT WILT FOR A MESOPHYTE IS 1500 K PA AND FOR ##	

A ZEROPHYTE 2400 K PA BUT THE DIFFERENCE IN % MOISTURE IS ## NEGLIGIBLE ## SOIL DEPTH SDP ## (MM) ATMOS TRANSMISSIVITY 0.75 ## ## ' WJ IS THE WATER CONTENT EXPRESSED AS A % ## ' CALCULATE GWP ## $GWP = -1500 * (WJ / WHC4) ^ M1$ 324 ' MEAN DAILY TEMPERATURE IS EQUAL TO THE MEAN OF ## THE ' DAILY MAXIMUM AND THE DAILY MINIMUM TEMPERATURE ## 325 ATEMP = (AMXT + AMNT) / 2GS = .4019914# + .01725101# * ATEMP - .0001485# * ATEMP ^ 2 326 ' JDA=JULIAN DAY ## 327 JDA = J + 181IF J > 184 THEN JDA = JDA - 365 328 ' SOLK=SOLARCONSTANT ## ' RFD=RADIANT FLUX DENSITY X = ((JDA + 10) / 365) * 360329 X = X * .01745330 $SOLK = 30.85 + 12.65 \times COS(X)$ 331 ALPHA = .21332 333 BETHA = .71DAYFRC = SUN / DALEN(NMNTH) 334 IF DAYFRC > .5 THEN ALPHA = .29 IF DAYFRC > .5 THEN BETHA = .5 RFD = SOLK * (ALPHA + BETHA * DAYFRC) 335 336 337 $RFDM = RFD * 10^{6} / (DALEN(NMNTH) * 3600)$ 338 ' CALCULATION OF EQUILIBRIUM EVAPORATION ## 339 PECONS = 1.28340 IF AMXT ≥ 20 THEN PECONS = 1.28 + .08 * (AMXT - 20) 341 EE = GS * .63 * RFD / 2.45' CALCULATION OF POTENTIAL EVAPORATION ## 342 PE = PECONS * EE' EXPRESS THE RATIO OF POT. EVAP TO PAN EVAP ## 343 IF EVAP = 0 THEN EVAP = 1344 PECVAP = PE / EVAP345 IF PECVAP > 1.5 THEN PECVAP = 1.5' 0.1 PLANT COVERAGE IS 10% THROUGHOUT THE YEAR. AL/ ' 0.9 WHEN AL=3 THE REMAINING GROUND WILL BE COVERED AL/3 * ## ΤN ## ' GRASS NEVER ALLOW (0.1 + AL/3 * 0.9) TO BECOME GREATER ## ' THAN ONE WHEN THIS HAPPENS (AL GRATER THAN 3) ASSUME AE= PE ## 346 B = (.1 + ((AL) / 3) * .9)IF B >= 1 THEN B = 1347 348 IF $B \le 0$ THEN B = 0' CALCULATION OF LEAF WATER POTENTIAL ## 349 LWP = PE / HYCON350 LWP = GWP - LWP

1 CALCULATE THE LIMITATION OF WATER AVAILIBILITY ON PHOTOSYN-## THETIC EFFICIENCY ## COEF = .1 * (WPC - LWP)351 IF COEF $\langle = (WPC / -100)$ THEN GOTO 204 352 COEF = WPC / -100353 IF COEF >= WPC / 100 THEN GOTO 205 COEF = WPC / 100 204 354 355 205 A = EXP(COEF)356 357 FW = 1 / (1 + A)' EVAPORATE RAINFALL LESS THAN 3,0 MM AS PART OF ACTUAL EVAP ## 358 IF RAINF <= 3 THEN PE = PE - RAINF 359 IF PE $\langle = 0 \text{ THEN PE} = 0$ IF RAINF $\langle = 3 \rangle$ Then rainf = 0 360 ' IF RAINFALL EXCEED 5,0 MM THEN EVAP ACCORDING TO EQ. ## 361 IF RAINF > 5 THEN CNT(J) = -2CNT(J + 1) = CNT(J) + 1FG = EXP(-.5 * CNT(J)) 362 363 IF FG \rightarrow = 1 THEN FG = 1 364 ' CALCULATE RUNOFF ## RUNOF = 0365 INFIL = (RAINF + IRRIG) / 1000 IF INFIL <= .2 * RUNPAR THEN RUNOF = 0 ELSE RUNOF = (INFIL - .2 * RUNPAR) ^ 2 / (INFIL + .8 * RUNPAR) 366 367 INFIL = (INFIL - RUNOF) * 1000 TRUNOF = TRUNOF + RUNOF 368 369 ' CALCULATE SOIL VAPORATION IE SLVAP ## 370 SLVAP = (1 - B) * FG * PETRANS1 = B * FW * PE371 372 AED = SLVAP + TRANS1' SLVAP CAN ONLY OCCUR FROM THE FIRST LEVEL IF WATER IS AVLBL ## 373 IF SLVAP $\langle =$ SLWAT1(J) - PWP1 THEN GOTO 206 ELSE SLVAP = SLWAT1(J) - PWP1ENDIF 374 206 SLWAT1(J) = SLWAT1(J) - SLVAP' RECHARGE OF LEVEL 1 COMES FROM RAIN(J) AND THAT OF LEVEL 2 ## FROM DRAINAGE OF LEVEL 1 WHEN IT HAS REACHED FC ## 375 SLWAT1(J) = SLWAT1(J) + RAINF + IRRIG - TRANS1 376 IF SLWAT1(J) \rightarrow = FC1 THEN GOTO 207 377 IF SLWAT1(J) \rightarrow = PWP1 THEN GOTO 208 TRANSPIRATION HOWEVER CAN OCCUR FROM BOTH LEVELS AND IF WATER## IS NOT AVAILIBLE IN LEVEL 1 THE REST WILL BE DRAWN FROM ## ' LEVEL 2 ## 378 TRANS2 = TRANS1 - (SLWAT1(J) - PWP1)379 TRANS1 = 0380 SLWAT1(J) = PWP1381 GOTO 208

382 207 DRN1P = 1DRAIN1 = (SLWAT1(J) - FC1) * DRN1P IF DRAIN1 <= 0 THEN DRAIN1 = 0 383 384 SLWAT1(J) = SLWAT1(J) - DRAIN1385 ' CALCULATE WATER CONTENT OF LEVEL 2 ## 386 208 SLWAT2(J) = SLWAT2(J) + DRAIN1 - TRANS2' EXPRESS THE RATIO OF ACT. EVAP TO PAN EVAP ## AECVAP = ae / EVAP 387 IF AECVAP > 1.5 THEN AECVAP = 1.5 388 ' IF SLWAT2 EXCEEDS FC2 DRAIN THE AMOUNT GREATER THAN FC2 ## 389 209 IF SLWAT2(J) \leq FC2 THEN GOTO 210 DRAIN2 = DRAINP * (SLWAT2(J) - FC2)390 IF DRAIN2 <= 0 THEN DRAIN2 = 0 391 392 SLWAT2(J) = SLWAT2(J) - DRAIN2' IF SLWAT2 GETS LOWER THAN PWP2..SLWAT2 IS EQUAL TO PWP2 ## IF SLWAT2(J) \rightarrow = PWP2 THEN GOTO 211 393 210 394 SLWAT2(J) = PWP2395 DRAIN2 = 0396 211 WAT = WJ' TO REDUCE SOIL WATER DURING WINTER I.E AFTER DAY 300 SOIL ' WATER IS REDUCED BY 0.5 MM/DAY 397 IF GRSTGE = 5 THEN SLWAT1(J) = SLWAT1(J) - .3IF GRSTGE = 5 AND SLWAT1(J) < PWP1 THEN SLWAT1(J) = PWP1 IF GRSTGE = 5 AND SLWAT1(J) = PWP1 THEN SLWAT2(J) = SLWAT2(J) - .1 398 399 IF GRSTGE = 5 AND SLWAT2(J) < PWP2 THEN SLWAT2(J) = PWP2 400 401 SLWAT(J + 1) = (SLWAT1(J) + SLWAT2(J)) / SDPSLWAT1(J + 1) = SLWAT1(J)SLWAT2(J + 1) = SLWAT2(J)402 403 404 WJ = SLWAT(J + 1) * 100405 STRESS = 1 - FWIF TRANS2 <= 0 THEN TRANS2 = .001 406 407 ALAI = AL408 TDRAIN = TDRAIN + DRAIN2 409 TDRN = DRAIN1 + DRAIN2 410 TSLW = SLWAT1(J) + SLWAT2(J)411 RETURN 412 300 1* SUBROUTINE TRANSLOCATION 413 IF GRSTGE \leftrightarrow 1 THEN DBL = 0 414 R(1) = DBL415 R(2) = DPROS416 R(3) = DPROR417 R(4) = DPROC418 R(5) = DPROG

TEST FOR THE AVAILABILTY OF RESERVES

,

419	IF RES > $R(1) + R(2) + R(3) + R(4) + R(5)$ THEN GOTO 317
,	RESET THE MASS FLOW VARIABLES TO ZERO
420 421 422	FOR IR = 1 TO 5 R(IR) = 0 NEXT IR
423 424 425 426 427 428 429 430 431 432 433	317 DRES = DMG - R(1) - R(2) - R(3) - R(4) - R(5) BL = BL + R(1) - X(2) BD = BD + X(2) - X(7) SL = SL + R(2) - X(1) SD = SD + X(1) - X(6) CL = CL + R(4) - X(9) CD = CD + X(9) - X(5) GL = GL + R(5) - X(8) GD = GD + X(8) - X(4) RL = RL + R(3) RD = RD + X(3)
434 435 436 437	RES = RES + DRES TRD = TR + $X(4)$ + $X(5)$ + $X(6)$ + $X(7)$ C = GL + GD + CL + CD + BL + BD + SL + SD + RL + RD CLIVE = GL + CL + BL + SL + RL
	COMPUTE FOR SOILN
438 439 440 441 442 443	TOTMASS(1) = BRL + BRD V(1) = SLWAT1(J) * 10: V(2) = SLWAT2(J) * 10 V01(1) = FC1 * 10: V01(2) = FC2 * 10 V15(1) = PWP1 * 10: V15(2) = PWP2 * 10 W(1) = V(1) * SDP1 / 1000: W(2) = V(2) * SDP2 / 1000 W01(1) = V01(1) * SDP1 / 1000: W01(2) = V01(2) * SDP2 / 1000
444 445 446 447	<pre>FOM(1) = TRD * SDP1 / SDP: FOM(2) = TRD * SDP2 / SDP T(1) = ATEMP: T(2) = ATEMP + 1.2 TAVE = ATEMP PERC(1) = DRAIN1: PERC(2) = DRAIN2</pre>
448	CALL SOILN(J, BRL, BCL, BBL, BGL, BRD, BCD, BBD, BGD, TRANS1, TRANS2, V(), VO1(), V15(), ANC(), NO3(), NH4(), WO1(), W(), IFOM(), FOM(), T(), INFIL, PERC(), DoyFert(), FERTO3(), FERTH4(), TAVE, GWP, N(), BO, FACN, GRSTGE, TOTMASS(), NFERT, FTMIN,FWMIN, CNR, FCNR, DECR, RDECR(), NMINFOM(), NFOM(), NIMMFOM(), NMINHUM(), NHUM(), DMINR, NMINNET(), HUM(), NIT(), NOUTFLO(), NLEACH, AN(), NDEM(), NCMAX(), NC(), TOTAN, TPAN, TNFLO, TNMASFLO)
449	RETURN
450	500 /***********************************
	' Fresh Organic Matter
451 452 453 454	FROM = 3000: FRHUM = 30000 IFOM(1) = FROM * SDP1 / SDP: IFOM(2) = FROM * SDP2 / SDP HUM(1) = FRHUM * SDP1 / SDP: HUM(2) = FRHUM * SDP2 / SDP NO3(1) = 1: NO3(2) = 1: NH4(1) = .5: NH4(2) = .5: N(1) = 20
455 456	<pre>FertSerie1(1) = 30: DoyFert(1) = 93: NFERT = 1 FOR i = 1 TO NFERT</pre>

457		FERT(i) = FertSerie1(i)
458		DoyToFert(i) = DoyFert(i)
459		FERTO3(i) = (FERT(i) * .8) * (62 / 80)
460		FERTH4(i) = (FERT(i) * .8) * (18 / 80)
461		PRINT FERT(i)
462		NEXT i
463	TO T	

463 RETURN

APPENDIX B

Source code for the soil nitrogen balance used in the PUTU 14 model.

SUB SOILN (J, BRL, BCL, BBL, BGL, BRD, BCD, BBD, BGD, TRANS1, TRANS2, V(), VO1(), V15(), ANC(), NO3(), NH4(), WO1(), W(), IFOM(), FOM(), T(), INFIL, PERC(), DoyToFert(), FERTO3(), FERTH4(), TAVE, FW, N(), BO, FACN, GRSTGE, TOTMASS(), NFERT, FTMIN,FWMIN, CNR, FCNR, DECR, RDECR(), NMINFOM(), NFOM(), NIMMFOM(), NMINHUM(), NHUM(), DMINR, NMINNET(), HUM(), NIT(), NOUTFLO(), NLEACH, AN(), NDEM(), NCMAX(), NC(), TOTAN, TPAN, TNFLO, TNMASFLO) STATIC 1 SUB SOIL NITROGEN TO CALCULATE NITRATE IN SOIL LAYERS (2) 'IFOM(L) 'Initial amount of fresh organic matter 'FOM(L) 'Fresh organic matter 'HUM(L) 'Stable organic matter 'T(L) 'Soil temperature 'V(L) 'Soil water content 'Ammonium in layer L (kg/ha) 'NH4(L) 'NO3(L) 'Nitrate in layer L (kg/ha) 'NFOM(L) 'Nitrogen in FOM 'Relative decay rate of humus DMINR = 8.3×10^{-5} 2 3 CNRFOM = 40'C:N Ratio of FOM 'C:N Ratio OF HUM 4 CNRHUM = 105 FLAG = FLAG + 16 FOR L = 1 TO 2 IF FLAG = 1 THEN NFOM(L) = .01 * FOM(L) IF FLAG = 1 THEN NHUM(L) = .04 * HUM(L) 7 8 IF FLAG = 1 THEN NHUM(L) = $.04 \times HUM(L)$ IF FOM(L) / IFOM(L) > .8 THEN RDECR(L) = .8IF FOM(L) / IFOM(L) < .8 THEN RDECR(L) = .05IF FOM(L) / IFOM(L) < .1 THEN RDECR(L) = .0095FTMIN = 1 / (1 + EXP(-.15 × (T(L) - 16))) FWMIN = 1 / (1* EXP(-GWP/1500)) 9 10 11 12 13 IF FWMIN < 0 THEN FWMIN = 0 14. $\begin{array}{l} \text{IF FWHIN} < \text{O IIIEN FWHIN} = 0 \\ \text{CNR} = (.4 * \text{FOM}(L)) / (\text{NFOM}(L) + \text{NO3}(L) + \text{NH4}(L)) \\ \text{FCNR} = \text{EXP}(-.693 * (\text{CNR} - 25) / 25) \\ \text{IF FCNR} > 1 \text{ THEN FCNR} = 1 \\ \text{DECR} = \text{RDECR}(L) * \text{FTMIN} * \text{FWMIN} * \text{FCNR} \\ \end{array}$ 15 16 17 18 19 NMINFOM(L) = NFOM(L) * DECRNIMMFOM(L) = DECR * FOM(L) * .0220 IF NIMMFOM(L) \rightarrow AN(L) THEN NIMMFOM(L) = .4 * AN(L) 21 **'UPDATE FOM & NFOM** FOM(L) = FOM(L) - DECR * FOM(L)22 23 NFOM(L) = NFOM(L) + NIMMFOM(L) - NMINFOM(L)'MINERALIZATION FROM HUMUS NMINHUM(L) = NHUM(L) * DMINR * FWMIN * FTMIN NMINNET(L) = .8 * NMINFOM(L) + NMINHUM(L) - NIMMFOM(L) 24 25 'Net mineralization 26 IF NMINNET(L) > 0 THEN NH4(L) = NH4(L) + NMINNET(L) 27 IF NMINNET(L) $\langle = 0$ THEN NH4(L) = NH4(L) + NMINNET(L) * NH4(L) / (NH4(L) + NO3(L)) 28 IF NMINNET(L) $\langle = 0$ THEN NO3(L) = NO3(L) + NMINNET(L) * NO3(L) / (NH4(L) + NO3(L)) 'UPDATE HUM & NHUM 29 NHUM(L) = NHUM(L) - NMINHUM(L) + .2 * NMINFOM(L)

```
30
         HUM(L) = HUM(L) - NMINHUM(L) * 10 + .2 * NMINFOM(L) / .04
31
       NEXT L
       'SIMPLE NITRIFICATION ROUTINE(25 % OF NH4 NITRIFIES PER DAY)
       FOR L = 1 TO 2
32
            NIT(L) = .25 * (NH4(L) - 1)
IF NIT(L) < 0 THEN NIT(L) = 0
33
34
            NH4(L) = NH4(L) - NIT(L)
35
            NO3(L) = NO3(L) + NIT(L)
36
37
       NEXT L
       'NITROGEN LEACHING & INFLUX BY RAIN
38
       IF INFIL > 0 THEN NINFLO(1) = 0 \times INFIL ELSE NINFLO(1) = 0
       FOR L = 1 TO 2
39
40
            NINFLO(L + 1) = (PERC(L) * NO3(L) / (WO1(L) + PERC(L))) * .2
41
            NOUTFLO(L) = NINFLO(L + 1)
42
            NO3(L) = NO3(L) + NINFLO(L) - NOUTFLO(L)
       NEXT L
43
44
       NLEACH = NLEACH + NOUTFLO(2)
       'NITROGEN FERTILIZER
       IF J = DoyToFert(NFERT) THEN
45
          NO3(1) = NO3(1) + FERTO3(NFERT)
46
47
          NH4(1) = NH4(1) + FERTH4(NFERT)
48
          NFERT = 1 + NFERT
49
       END IF
       'UPDATE MINERAL N IN SOLUTION & SOIL
50
       FOR L = 1 TO 2
51
            AN(L) = ((NO3(L) - 1) + (NH4(L) - 1)) * .9
            IF AN(L) < 0 THEN AN(L) = 0
52
53
            ANC(L) = AN(L) / W(L)
       NEXT L
54
     CALL PLANTN(J, BRL, BCL, BBL, BGL, BRD, BCD, BBD, BGD, TRANS1, TRANS2,
ANC(), AN(), V(), V15(), NO3(), NH4(), TAVE, FW, N(),
BO, FACN, GRSTGE, TOTMASS(), NDEM(), NCMAX(), NC(),
TOTAN, TPAN, TNFLO, TNMASFLO, TOTNUP)
55
```

56 END SUB

APPENDIX C

Source code for the plant nitrogen balance used in the PUTU 14 model.

SUB PLANTN (J, BRL, BCL, BBL, BGL, BRD, BCD, BBD, BGD, TRANS1, TRANS2, ANC(), AN(), V(), V15(), NO3(), NH4(), TAVE, FW, N(), BO, FACN, GRSTGE, TOTMASS(), NDEM(), NCMAX(), NC(), [:] 1 TOTAN, TPAN, TNFLO, TNMASFLO, TOTNUP) STATIC 'SUB TO CALCULATE N DEMAND, TRANSLOCATION & LIMITATION IN THE PLANT 2 MASS(2) = BCL + BCD: MASS(3) = BBL + BBD: MASS(4) = BGL + BGDFOR COMP = 2 TO 4 3 IF MASS(COMP) > TOTMASS(COMP) THEN 4 5 TOTMASS(COMP) = MASS(COMP)END IF 6 7 NEXT COMP 'MINIMUM AND MAXIMUM NITRATE CONTENT IN PLANT FOR (ROOTS, CULMS, LEAVES & GRAIN) 8 9 10 NCMIN(1) = .0047511 NCMIN(2) = (J / 365 * 5 * -.00133) + .011731NCMIN(3) = (J / 365 * 5 * -.00296) + .0330712 13 14 LNCL = .0047515 TS(1) = TRANS1: TS(2) = TRANS2**'DEMAND FOR NITRATE** TAU2 = 2: TNDEM = 0 16 IF GRSTGE = 5 THEN EOSF = 0 ELSE EOSF = 117 FOR COMP = 1 TO 318 NDEM(COMP) = (TOTMASS(COMP) * (NCMAX(COMP) - NC(COMP))) / TAU2 IF NDEM(COMP) < 0 THEN NDEM(COMP) = 0 TNDEM = (TNDEM + NDEM(COMP)) * EOSF 19 20 21 22 NEXT COMP 'MAXIMUM NITROGEN UPTAKE 23 NUPO = 6 * (1 - EXP(-.005 * (TOTMASS(2) + TOTMASS(3))))24 'FLOW BY MASS 25 TNMASFLO = 0FOR L = 1 TO 2 26 27 IF TS(L) < 0 THEN TS(L) = 028 NMASFLO(L) = TS(L) * ANC(L)29 TNMASFLO = TNMASFLO + NMASFLO(L) 30 NEXT L IF (GRSTGE<=4) THEN NDIFFLOO=(TNDEM-TNMASFLO) / 1.5 ELSE NDIFFLOO = 31 0 32 IF NDIFFLOO < 0 THEN NDIFFLOO = 0 33 TOTAN = 0: TNDIFFLO = 034 FOR L = 1 TO 2 IF $V(L) \rightarrow = V15(L)$ THEN TOTAN = TOTAN + AN(L) 35 36 NEXT L 37 FOR L = 1 TO 2 38 IF TOTAN AND $V(L) \ge V15(L)$ THEN NDIFFLO(L) = NDIFFLOO * AN(L) / TOTAN ELSE NDIFFLO(L) = 039 MASDIFLO(L) = NMASFLO(L) + NDIFFLO(L)

40 TNDIFFLO = TNDIFFLO + NDIFFLO(L)NEXT L 41 TNFLO = TNMASFLO + TNDIFFLO 42 'CALCULATE UPTAKE IF NUPO <= TNFLO THEN NUP = NUPO ELSE NUP = TNFLO 43 44 IF NUP > TNDEM THEN NUP = TNDEM 45 IF NUP > TOTAN THEN NUP = TOTAN * .9 IF NUP < 0 THEN NUP = 0 46 47 TOTNUP = TOTNUP + NUP'UPDATE NO3 & NH4 IN SOIL FOR L = 1 TO 2 48 IF TNFLO > 0 THEN DNO3(L) = MASDIFLO(L) / TNFLO * NUP * NO3(L) / 49 (NO3(L) + NH4(L)) ELSE DNO3(L) = 0 IF TNFLO > 0 THEN DNH4(L) = MASDIFLO(L) / TNFLO * NUP * NH4(L) / 50 (NO3(L) + NH4(L)) ELSE DNH4(L) = 0 51 NO3(L) = NO3(L) - DNO3(L)NH4(L) = NH4(L) - DNH4(L)52 53 NEXT L 'NITROGEN DISTRIBUTION & TURNOVER RFNS = 1 - (1 - FACN ^ 2) 54 ONC = .0755 56 DN(4) = TOTMASS(4) * (ONC - NC(4)) * RFNSIF DN(4) < 0 THEN DN(4) = 057 N(4) = N(4) + DN(4)58 59 IF TOTMASS(4) > 0 THEN NC(4) = N(4) / TOTMASS(4) 60 TPAN = 0FOR COMP = 1 to 361 62 IF TNDEM * NUP > 0 THEN DN(COMP) = NDEM(COMP) / TNDEM * NUP 63 N(COMP) = N(COMP) + DN(COMP)64 IF TOTMASS(COMP) > 0 THEN NC(COMP) = N(COMP) / TOTMASS(COMP) 65 IF NC(COMP) <= NCMIN(COMP) THEN PAN(COMP) = 0 ELSE PAN(COMP) = (NC(COMP) - NCMIN(COMP)) * TOTMASS(COMP) 66 TPAN = TPAN + PAN(COMP)67 NEXT COMP 68 FOR COMP = 1 TO 369 IF TPAN > 0 THEN DN(COMP) = DN(4) * PAN(COMP)/TPAN ELSE DN(COMP)=070 N(COMP) = N(COMP) - DN(COMP)IF TOTMASS(COMP) > 0 THEN NC(COMP) = N(COMP) / TOTMASS(COMP) 71 NEXT COMP 72 'EFFECT OF N STATUS ON PROCESSES 73 NCOPT(3) = NCMAX(3) * .874 IF NCOPT(3) > 0 THEN FACN(3) = (NC(3) - NCMIN(3)) / (NCOPT(3) -NCMIN(3)) 75 IF FACN(3) \lt 0 THEN FACN(3) = 0 76 FACN = FACN(3)77 IF FACN > 1 THEN FACN = 1 78 END SUB

1	08

APPENDIX D

D.1 Table of high, low and average yield values (t ha^{-1}) simulated for a bad year.

FACTOR		High	Low	Average	
TIME	1	High 0.9 1.1 0.9 0.5 0.7	0.5	0.7	
	2	1.1	0.5	0.8	
	3	0.9	0.5	0.7	
RATE	5	0.5	0.5	0.5	
	10	0.7	0.6	0.7	
	15	0.9	0.8	0.8	
	20	1.1	0.8	0.9	
	25	1.1	0.8	0.9	

D.2 Table of high, low and average yield values (t ha⁻¹) simulated for a poor year.

FACTOR		High	Low	Average
TIME	1	2.3 2.1	0.8	1.7
	2	2.1	0.9	1.7
	3	1.7	0.9	1.5
RATE	15	0.9	0.8	0.9
	30	1.4	1.3	1.4
	45	1.8	1.7	1.7
	60	2.2	1.7	2.0
	75	2.2	1.7	2.0

D.3 Table of high, low and average yield values (t ha⁻¹) simulated for a moderate year.

FACTOR		High	Low	Average	
TIME	1	3.3	1.3	2.5	
	2	3.6	1.3	2.7	
	3	2.3	1.4	2.0	
RATE	25	1.4	1.3	1.3	
	50	2.2	2.1	2.1	
	75	2.9	2.3	2.6	
	100	3.6	2.3	3.0	
	125	3.6	2.3	3.0	

FACTOR		High	Low	Average
TIME	1	4.7	1.7	3.5
	2	5.5	1.9	3.8
	3	4.2	1.9	3.5
RATE	35	1.9	1.7	1.8
	70	3.1	2.7	2.9
	105	4.2	3.8	3.9
	140	5.1	4.2	4.7
	175	5.5	4.1	4.8

D.4 Table of high, low and average yield values (t ha⁻¹) simulated for a good year.

D.5 Table of high, low and average yield values (t ha⁻¹) simulated for a wet year.

FACTOR		High	Low	Average	
TIME	1	7.7 7.3 5.7	2.8	5.8	
	2	7.3	2.7	5.7	
	3	5.7	2.8	4.9	
RATE	60	2.8	2.7	2.8	
	120	4.7	4.5	4.6	
	180	6.4	5.7	6.2	
	240	7.7	5.7	6.9	
	300	7.7	5.7	6.9	

D.6 Simulated seasonal yield kg ha⁻¹ from 1916-1991 for different N-levels

Year	N(0)	N(30)	N(60)	N(90)	N(120)	N(150)
1916	249.3	268.0	268.0	268.0	268.0	268.0
1917	400.3	1599.1	2464.2	3432.2	4322.6	5159.4
1918	529.2	1699.9	2561.6	3436.0	4361.3	5315.8
1919	429.8	1276.2	1755.2	1755.2	1755.2	1755.2
1920	243.9	411.1	411.1	411.1	411.1	411.1
1921	488.6	1544.9	2424.4	3355.3	3820.8	3820.8
1922	380.4	1535.4	2375.1	3575.5	3575.5	3575.5
1923	452.3	1093.5	1093.5	1093.5	1093.5	1093.5
1924	547.9	1390.2	2284.3	3248.0	3527.4	3527.4
1925	418.0	1210.6	1210.6	1210.6	1210.6	1210.6
1926	535.8	1347.4	2354.3	2354.3	2354.3	2354.3
1927	470.2	1363.3	2041.2	2041.2	2041.2	2041.2
1928	491.0	1379.0	2333.6	2622.3	2622.5	2622.5
1929	422.8	1643.0	2487.7	3404.7	4301.9	5420.1
1930	393.0	1292.6	1947.3	1947.3	1947.3	1947.3
1931	423.9	1220.8	1294.3	1294.3	1294.3	1294.3
1932	372.3	1246.2	1246.2	1246.2	1246.2	1246.2
1933	434.8	1281.6	2311.1	2311.1	2311.1	2311.1
1934	417.7	1336.4	2272.5	2828.0	2828.0	2828.0
1935	414.3	665.1	665.1	665.1	665.1	665.1
1936	328.9	731.7	731.7	731.7	731.7	731.7

Year	N(Ö)	N(30)	N(60)	N(90)	N(120)	N(150)
1937	331.6	922.0	922.0	922.0	922.0	922.0
1938	441.9	1290.7	2083.9	2083.9	2083.9	2083.9
1939	530.0	1336.1	2152.0	2152.0	2152.0	2152.0
1940	425.7	1333.6	2367.6	2367.6	2367.6	2367.6
1941	391.3	1093.3	1093.3	1093.3	1093.3	1093.3
1942	734.8	1355.5	2258.0	2658.6	2659.8	2660.9
1943	456.6	1917.6	2635.8	3458.9	4427.3	5237.8
1944	459.9	1285.1	1792.4	1792.4	1792.4	1792.4
1945	445.0	1022.6	1022.6	1022.6	1022.6	1022.6
1946	267.9	286.8	286.8	286.8	286.8	286.8
1947	410.4	1271.8	2032.3	2032.3	2032.3	2032.3
1948	394.0	1284.1	1284.1	1284.1	1284.1	1284.1
1949	937.2	1457.1	2369.3	3253.8	3907.4	3907.4
1950	400.3	1282.8	2034.1	2034.1	2034_1	2034.1
1951	433.8	1314.9	1815.2	1815.2	1815.2	1815.2
1952	646.9	1438.5	2368.3	3262.5	3848.2	3848.2
1953	384.1	1312.1	2117.9	2117.9	2117.9	2117.9
1954	504.0	1318.4	2625.9	2625.9	2625.9	2625.9
1955	289.0	529.4	529.4	529.4	529.4	529.4
1956	722.7	1387.3	2253.6	3269.2	3269.2	3269.2
1957	918.9	2104.3	2819.5	4367.8	4367.9	4368.0
1958	442.4	1408.2	2291.1	3072.9	3072.9	3072.9
1959	412.6	782.1	782.1	782.1	782.1	782.1
1960	665.7	1389.5	2293.5	3189.4	3397.7	3397.7
1961	387.2	728.8	728.8	728.8	728.8	728.8
1962	514.0	1146.4	1146.4	1146.4	1146.4	1146.4
1963	329.1	917.8	917.8	917.8	917.8	917.8
1964	327.8	908.4	908.4	908.4	908.4	908.4
1965	423.1	1305.8	2370.7	2370.7	2370.7	2370.7
1966	532.0	1381.5	2339.3	3246.5	3246.5	3246.5
1967	509.4	1224.6	1336.2	1336.2	1336.2	1336.2
1968	555.0	1320.8	2236.8	2749.1	2749.1	2749.1
1969	376.3	1277.6	1622.7	1623.4	1624.1	1624.8
1970	568.4	1412.0	2352.3	3259.2	3706.2	3706.2
1971	586.2	1393.7	2365.5	3484.3	3499.3	3499.3
1972	302.2	365.4	365.4	365.4	365.4	365.4
1973	461.1	1302.3	2365.1	2365.1	2365.1	2365.1
1974	489.9	1255.9	1255.9	1255.9	1255.9	1255.9
1975	518.6	1319.3	2230.9	2895.1	2895.1	2895.1
1976	366.2	812.1	812.1	812.1	812.1	812.1
1977	434.9	832.6	832.6	832.6	832.6	832.6
1978	263.3	1269.5	2264.5	2325.5	2325.5	2325.5
1979	502.3	1484.6	2643.2	3437.7	4222.0	4930.3
1980	777.6	1999.7	2619.7	3498.8	4389.1	5373.0
1981	455.3	1296.7	1877.6	1877.7	1877.7	1877.8
1982	471.4	1365.2	2237.3	2533.1	2533.1	2533.1
1983	487.1	1362.9	2289.5	2728.5	2728.5	2728.5
1984	355.8	1467.8	2315.5	3242.2	3577.7	3577.7
1985	396.3	1118.6	1118.6	1118.6	1118.6	1118.6
1986	573.4	1510.3	2377.1	3340.3	4863.3	4924.3
1987	587.03	2173.1	2653.6	3495.4	4395.4	5356.0
1988	1041.48	1463.1	2379.4	3277.4	4397.6	4397.6
1989	413.36	1244.8	1777.6	1777.6	1777.6	1777.6
1990	435.66	1407.6	1799.6	1799.6	1799.6	1799.6
1991	367 90	936.2	936.2	936.2	936 2	936 2

