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# PALEOPOLYPLOIDY AND MOLECULAR SYSTEMATICS OF SOUTHERN AFRICAN CHLORIDOIDEAE

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Dedicated, with love, to my parents  
Maarten and Lea



'Considering the widespread occurrence and ecological diversity of grasses, their dominance over vast regions of our globe, and their prime importance to humankind, we, the experts, may congratulate ourselves on having become authorities on the most important single family of organisms in the world of life, rivaled only by the human family itself.'

G.L. STEBBINS 1987

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# Abbreviations<sup>1</sup>

A	Adenine
ABI	Applied Biosystems Incorporated, Foster City, California, USA
<i>Adh</i>	Alcohol Dehydrogenase
<i>atpB</i>	ATP Synthase Beta Subunit
BEP	Bambusoideae-Ehrhartoideae-Pooideae
BOP	Bambusoideae-Oryzoideae-Pooideae
BLFU	Geo Potts Herbarium, Department of Plant Sciences, University of the Free State, Bloemfontein
bp	Basepair
<i>BglI</i>	<i>Bacillus globigii</i> II
BS	Bootstrap Index
C	Cytosine
cpDNA	Chloroplast DNA
CI	Consistency Index
CTAB	Hexadecyl-Trimethyl-Ammonium Bromide
DMSO	Dimethyl Sulfoxide
DNA	Deoxyribonucleic Acid
dNTP	Deoxynucleotide Triphosphate
EDTA	(Ethylenediamine) Tetra-Acetic Acid
Ethanol	Ethylalcohol
F	Coefficient of Similarity
G	Guanine
GBSS1	Granule Bound Starch Synthase Subunit 1
GPWG	Grass Phylogeny Working Group
HCl	Hydrochloric Acid
<i>HinfI</i>	<i>Haemophilus influenzae</i> RF I
IGS	Intergenic Spacer Region
ILD	Incongruence Length Difference Test
Indels	Insertions/Deletions
<i>ITS</i>	Internal Transcribed Spacer Region

<sup>1</sup> SI units not included in abbreviation list

kb	Kilobase
M	Molar
<i>matK</i>	Maturase K
MgCl <sub>2</sub>	Magnesium Chloride
mM	Millimolar
mmol	Millimoles
m/m	Mass per Mass
m/v	Mass per Volume
n	Gametic Chromosome Number
2n	Somatic Chromosome Number
NaCl	Sodium Chloride
NAD	Non-reduced form of Nicotine Amideadenine Dinucleotide
NADH	Reduced form of Nicotine Amideadenine Dinucleotide
NAD-ME	NAD-Malic Enzyme
<i>ndhF</i>	NADH Dehydrogenase, Subunit F
NNI	Nearest Neighbor Interchange Branch Swapping
nrDNA	Nuclear Ribosomal Deoxyribonucleic Acid
PACC	Panicoideae-Arundinoideae-Centothecoideae-Chloridoideae
PACCAD	Panicoideae-Arundinoideae-Centothecoideae-Chloridoideae-Aristidoideae-Danthonioideae
PAUP	Phylogenetic Analysis Using Parsimony
pBR328	<i>E. coli</i> Plasmid pBR328
PCK	Phosphoenol Pyruvate Carboxikinase
PCR	Polymerase Chain Reaction
<i>phyB</i>	Phytochrome B
pmol	Picomoles
PRE	National Herbarium, Pretoria
<i>psbA</i>	DII Protein of Photosystem II Reaction Center
PVP	Polyvinyl Pyrrolidone
<i>rbcl</i>	Ribulose -1,5- Bisphosphate Carboxylase/Oxygenase Large Subunit
rDNA	Ribosomal Deoxyribonucleic Acid
RI	Retention Index
RNA	Ribonucleic Acid

<i>rpl16</i>	Ribosomal Plastid Protein 16
<i>rpoC1</i>	RNA Polymerase Beta' Subunit
<i>rpoC2</i>	RNA Polymerase Beta" Subunit
<i>rps4</i>	Ribosomal Plastid Small Subunit Protein 4
rRNA	Ribosomal Ribonucleic Acid
SNL	Signal to Noise Ratio
T	Thymine
TAE	Tris-Acetic Acid-EDTA
<i>Taq</i> polymerase	<i>Thermus aquaticus</i> DNA Polymerase
TBR	Tree Bisection and Reconnection Branch Swapping
Tris	2-Amino-2-(Hydroxymethyl)-1,3-Propanediol
<i>trnL</i>	tRNA – Leu (UAA)
<i>trnL-F</i>	Region including <i>trnL</i> intron, <i>trnL</i> 3' exon, <i>trnL-F</i> spacer
<i>trnF</i>	tRNA – Phe (GAA)
<i>trnT</i>	tRNA – Thr (UGU)
u	Units
μl	Microlitre
μm	Micrometre
U	Uracil
UV	Ultraviolet
V	Volt
v/v	Volume per Volume
<i>waxy</i>	GBBS1
x	Basic Chromosome Number
3'	3 prime position
5'	5 prime position



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To my Father for His love and Heavenly Hands that carried me where others and my strength alone failed me.

## Noteworthy

Several chapters of this thesis dealing with the results obtained in this study have been submitted to various journals as manuscripts. Some of these (Chapter 3) have already been published, others are in press (Chapters 4 and 5) and Chapters 6 and 7 are still with the referees. As not to duplicate sections of the thesis, the materials and methods as well as references for each manuscript have been omitted from these five chapters. Two separate chapters (Chapter 2 and 9) are devoted to these subjects. Because the different journals require different formats, these chapters have been adapted to a universal format that is used throughout this thesis and these will differ from the format prescribed by each journal. Furthermore, due to the fact that each of these chapters is seen as separate manuscripts, taxonomic authorities are cited in each chapter. Throughout this thesis, American English was used.

# Summary

This study dealt with systematics of southern African representatives of the grass subfamily Chloridoideae. The group was studied on molecular and cytogenetic levels.

Two main basic chromosome numbers in the Chloridoideae, namely  $x = 9$  and  $10$ , were confirmed by this study. The basic chromosome number of  $x = 10$  is the most prevalent and is seen as the original number from which other aneuploid deviations in the group arose. The basic chromosome number of  $x = 10$  is, however, a paleopolyploid number as specimens with  $2n = 2x = 10$  have been found in the subfamily. Most of the chromosome numbers found in the Chloridoideae are derived from the original basic chromosome number,  $x = 5$ , or the paleopolyploid number,  $x = 10$ .

Polyploidy is frequent in the grasses and subfamily Chloridoideae. In this study more than 70% of the southern African Chloridoideae was found to be polyploid. This polyploidy is largely attributed to hybridization, as many of the studied specimens were segmental allopolyploids or allopolyploids. This is facilitated by an effective asexual reproduction system in the form of apomixis.

Two genomic regions were sequenced in this study, i.e. the nuclear *ITS* and chloroplast *trnL-F* regions. These two regions represent two different genomes and are inherited differently (maternal versus biparental), which have phylogenetic implications for studying hybridization, a frequent phenomenon in the Chloridoideae.

The regions studied differed in the amount of resolution they provided. The *ITS* phylogeny was well resolved, but the *trnL-F* region had less variation and less resolution, especially at species level. Despite this no hard incongruence was found between the two phylogenies and they could be combined.

The phylogenetic analyses indicated the monophyletic nature of the Chloridoideae. The two large tribes, Cynodonteae and Eragrostideae were polyphyletic, although a general division into two separate groups was evident. The monophyly of all the generic groups in the subfamily was well supported, except for the two largest genera in the study, *Eragrostis* and *Sporobolus*. These two genera are very variable and taxonomically difficult groups, probably related to interspecific and -generic hybridization.

The morphologically distinct tribe Pappophoreae was well supported in all analyses. The two genera *Entplocamia* and *Fingerhuthia* was found basal in the combined analysis, a finding that supports the derivation of the Chloridoideae from arundinoid ancestors as these two genera are seen as a link to *Spartochloa*, *Styppeiochloa* and *Tribolium* in the Arundinoideae and Danthonioideae.

Despite the frequency of hybridization in the subfamily, hybrids could not be positively identified based on sequence polymorphisms or their phylogenetic behavior. This is possibly related to the age of hybridization in the group or the close relationship of the groups between which hybridization occurs.

This study provides cytogenetic and molecular systematic support for paleopolyploidy in the Chloridoideae. This is based mainly on the occurrence of  $x = 5$  in the subfamily and the close relationship of the Chloridoideae to the Arundinoideae and Danthonioideae which have a main basic chromosome number of  $x = 6$  and from which  $x = 5$  in the Chloridoideae was derived. This chromosome number was probably highly unstable and subsequent polyploidization lead to the now frequent  $x = 10$  found in the majority of the subfamily.

**Keywords:** Chloridoideae, DNA sequencing, hybridization, *ITS* region, paleopolyploidy, phylogenetic relationships, *trnL-F* region

# Opsomming

Hierdie studie het gehandel oor die sistematiek van Suid-Afrikaanse verteenwoordigers van die gras subfamilie Chloridoideae. Die groep is bestudeer op molekulêre en sitogenetiese vlak.

Hoofsaaklik twee basiese chromosoomgetalle word in die Chloridoideae gevind, naamlik  $x = 9$  en  $10$  en is bevestig deur hierdie studie. Laasgenoemde basiese chromosoomgetal is die mees algemene en word gesien as die oorspronklike basiese chromosoomgetal waaruit ander aneuploïdiese afwykings in die groep ontstaan het. Die basiese chromosoomgetal van  $x = 10$  is egter 'n paleopoliploïede getal aangesien eksemplare met  $2n = 2x = 10$  al in die subfamilie gevind is. Meeste van die chromosoomgetalle wat in die Chloridoideae gevind word het ontstaan uit die oorspronklike basiese chromosoomgetal,  $x = 5$  of die paleopoliploïede basiese chromosoomgetal,  $x = 10$ .

Poliploïdie kom algemeen voor in die grasse en die subfamilie Chloridoideae. In hierdie studie was meer as 70% van die Suid-Afrikaanse Chloridoideae poliploïed. Hierdie poliploïdie word toegeskryf aan verbastering, aangesien meeste van die bestudeerde spesies segmenteel allopoliploïed of allopoliploïed was. Dit word bewerkstelling deur 'n effektiewe ongeslagtelike voortplantingsstelsel in die vorm van apomiksie.

Die nukleotiedvolgordes van twee genomiese gebiede, die kern *ITS* en chloroplas *trnL-F* gebiede, is bepaal. Hierdie twee gebiede verteenwoordig verskillende genome en word verskillend oorgeërf (moederlik teenoor oorerwing vanaf albei ouers) wat filogenetiese implikasies vir die bestudering van verbastering, wat 'n algemene verskynsel in die Chloridoideae is, inhou.

Die gebiede bestudeer verskil in die hoeveelheid resolusie wat hulle verskaf. Die *ITS* filogenie het goeie resolusie verskaf, maar die *trnL-F* gebied het minder variasie en swakker resolusie verskaf, veral op spesievlak. Ten spyte hiervan is geen sterk onverenigbaarheid tussen die twee filogenieë gevind nie en kon hulle gekombineer word.

Die filogenetiese analise toon aan dat die Chloridoideae monofileties is. Die twee groot tribusse, Cynodonteae en Eragrostideae, was polifileties, alhoewel 'n algemene skeiding in twee groepe sigbaar is. Die monofilie van al die genera in die

subfamilie word goed ondersteun, behalwe die twee grootste genera in die studie, *Eragrostis* en *Sporobolus*. Hierdie twee genera toon baie variasie en is taksonomiese moeilike groepe wat moontlik as gevolg van verbastering tussen spesies en genera is.

Die morfologiese kenmerkende tribus Pappophoreae is goed ondersteun in alle analises. Die twee genera *Entoplocamia* en *Fingerhuthia* was basaal in die gekombineerde analise. Dit ondersteun die ontwikkeling van die Chloridoideae vanaf arundinoid voorouers, aangesien hierdie twee genera gesien word as naverwant aan *Spartochloa*, *Styppeiochloa* en *Tribolium* in die Arundinoideae en Danthonioideae.

Ongeag die groot hoeveelheid verbastering wat in die subfamilie voorkom kon basters nie met sekerheid geïdentifiseer word op grond van nukleotiedvolgorde polimorfismes of hulle filogenetiese gedrag nie. Dit hou moontlik verband met die ouderdom van die verbastering in die groep of die noue verwantskap van die groepe waarin daar verbastering voorkom.

Hierdie studie lewer sitogenetiese en molekulêre sistematiese ondersteuning vir paleopoliploidie in die Chloridoideae. Dit word grootliks gegrond op die voorkoms van  $x = 5$  in die subfamilie en die noue verwantskap van die Chloridoideae met Arundinoideae en Danthonioideae met 'n basiese chromosoomgetal van  $x = 6$  en waaruit  $x = 5$ , wat in die Chloridoideae voorkom, kon ontwikkel het. Hierdie chromosoomgetal was moontlik hoogs onstabiel en poliploidisering het gelei tot die ontstaan van  $x = 10$  in die meeste genera en spesies in die subfamilie.

**Sleutelwoorde:** Chloridoideae, DNA nukleotiedvolgordebepaling, filogenetiese verwantskappe, ITS gebied, paleopoliploidie, *trnL-F* gebied, verbastering

---

**CHAPTER 1**  
**Introduction**

---



The grass family and in particular the subfamily Chloridoideae is well represented in southern Africa. Phenomena such as hybridization and polyploidization (>70%), as well as apomictic reproduction, have a high frequency in southern Africa. The evolution of the subfamily, as well as the grass family, is characterized by these events and nuclear and chloroplast DNA might tell different evolutionary stories, depending on the extent of these events.

## 1.1 Grasses

Poaceae (R.Br.) Barnhart inhabit the earth in greater magnitude than any other corresponding plant group (Gould 1968). According to Watson and Dallwitz (1992 onwards), there are about 700 genera and approximately 12 000 species of grasses in the world. They are the fourth largest flowering plant family (Liang and Hilu 1996), after Asteraceae Dumort., Orchidaceae Juss. and Fabaceae Lindl. (Tzvelev 1989) and are the second largest monocot family after Orchidaceae (Watson 1990). When comparatives are drawn between completeness of representation and percentage of the world's total vegetation, Poaceae far outnumbers any other family (Gould 1968). It is economically the most important and ecologically the most dominant plant family (Liang and Hilu 1996).

Grasses are the world's most important agricultural plants. This family includes (Chapman and Peat 1992; Van Oudtshoorn 1999):

- Cereals.
- Forage grasses.
- Grasses used as industrial raw materials.
- Bamboos with an abundance of uses.
- Grasses essential in soil conservation.
- Grasses utilized for the essential volatile oils they produce.
- A large group of ornamental grasses used in horticultural practices and used on, for example, sports grounds, lawns, parks, etc.

Direct utilization of grasses includes (Watson 1990):

- Modification of natural grasslands, along with improvement by the addition of management techniques and fertilizers.

- The creation of grasslands, lawns, etc.
- Intentional cultivation, together with selection, for genetic enhancement and the subsequent distribution and planting of pastures, cereal crops, etc.
- Cultivation of species such as sugarcane, culinary herbs, raw materials for the rope and paper industry.
- Exploitation of certain specialized forms as soil binders and stabilizers
- Applications of bambusoids as barrier plants.
- Use of various plant parts as construction material, etc.
- As a food source.

Except for their usefulness to man and livestock, they are also capable of serving as alternative hosts for pathogens and pests, which could affect crops. Some of the most invasive weeds belong to Poaceae. Seven of the top ten (May 1981) are species of the genera *Cynodon* Rich., *Echinochloa* P.Beauv., *Eleusine* Gaertn., *Imperata* L., *Panicum* L. and *Sorghum* Moench (Watson 1990).

The relative success of the grass family can be ascribed to features such as their morphology, anatomy, habit and reproductive cycles, which gives this family an advantage over its main enemies, namely man and herbivorous animals (Watson 1990). Furthermore the grasses have unusual adaptability. This allowed the first grasses to invade a wide range of habitats, and the ability to rapidly and effectively exploit changes in the environment (Watson 1990). This is exemplified by the evolution of the C<sub>4</sub> photosynthetic pathway, and its different types, more than once in the family (Clayton and Renvoize 1986; Sinha and Kellogg 1996; Kellogg 1999, 2000, 2001). Furthermore, the family is characterized by hybridization and polyploidization, along with great versatility in reproductive strategy (Watson 1990).

Grasses probably evolved during the Paleocene between 60 and 55 million years ago. This is based on grass pollen records from South America and Africa (Jacobs *et al.* 1999). The first grasses were probably mostly adapted to forest and shade habitats. The shift to drought tolerance and open habitats marked the major diversification of the family during the mid-Miocene, corroborated by the increase in the amount of pollen in the fossil record (Jacobs *et al.* 1999).

Currently 12 subfamilies are recognized: Anomochlooideae Pilg. ex Potzal, Aristidoideae Caro, Arundinoideae Burmeist., Bambusoideae Luerss., Centothecoideae Soderstr., Chloridoideae Kunth ex Beilschm., Danthonioideae Barker & H.P.Linder, Ehrhartoideae Link, Panicoideae Link, Pharoideae (Stapf) L.G.Clark & Judz., Pooideae Benth. and Puelioideae L.G.Clark, M.Kobay, S.Mathews, Spangler & E.A.Kellogg [Grass Phylogeny Working Group (GPWG) 2001].

## 1.2 Subfamily Chloridoideae

### 1.2.1 Classification

Brown (1810, 1814) divided the grasses into two major groups or tribes (now known as subfamilies): the 'Paniceae' (mostly tropical grasses, spikelets with two florets of which the lower is imperfect, generally comparable with the modern Panicoideae) and the 'Poaceae' (grasses with temperate distribution, spikelets with one to many florets, imperfect florets never basal, generally comparable with Festucoideae Link of Hitchcock (1971) or the modern Pooideae). Various taxonomists through the previous two centuries upheld this division.

The modern chloridoid grasses were located in the Pooideae. Zoysieae Benth. was only later moved from their initial position in Panicoideae to the Pooideae (Stapf 1900; Hitchcock 1936). In the early 1900's, Krause (1909, 1910, 1913) stated that the Chlorideae Rchb. was more closely related to the Panicoideae based on anatomical and epidermal studies, and advocated its removal from the Pooideae. Van Teighem (1897) drew the same conclusion based on his embryological studies in which he distinguished panicoid and festucoid embryo types.

In 1936, Prat recognized the Chloridoideae on the bases of grass leaf anatomy and epidermal structure. Tateoka *et al.* (1959) described the inflated spherical (egg-shaped) microhairs as the chloridoid type. Reeder (1957) described the characteristic chloridoid embryo features (P+PF).

Details of embryo structure are believed to be of great importance in the classification of grass subfamilies and are still in use at the genus level or higher (Hilu and Wright 1982; Watson *et al.* 1985). The formula that Reeder (1957, 1961,

1962) proposed is based on the embryo types described by Van Teighem (1897) and uses the following four features:

- Elongation of the vascular system in the embryo, with a mesocotyl present (P) or absent (F).
- The presence (+) or absence (-) of an epiblast.
- The presence (P) or absence (F) of a groove or cleft between the lower part of the scutellum and the coleorhiza.
- The position of the margins of the first embryonic leaves either rolled (P) or folded (F).

The concept of the chloridooid subfamily as now perceived became widely accepted when Pilger (1956) presented the group as the Eragrostoideae.

Over the last fifty years, various classifications for the Chloridoideae have been proposed (Table 1.1). These differences center on whether the Eragrosteae Benth (or Eragrostideae Stapf), Sporoboleae Stapf or the other smaller tribes should be included in the Cynodonteae Dumort. (or Chlorideae) (Jacobs 1987). Campbell (1985) included the two larger tribes Eragrosteae and Sporoboleae in the Cynodonteae. The recognition of two main tribes, which are based on the large genera *Eragrostis* Wolf and *Chloris* Sw., remains an important theme in all the proposed classifications. This concept was inherited from Pilger (1956) and is a division between paniculate inflorescences with two or more floreted spikelets on the one hand, versus inflorescences with spike-like main branches and one-floreted spikelets, on the other. The other subtribes and additional groups are usually segregated from one of these two groups (Table 1.1).

Only a few of the above mentioned classifications are worldwide treatments (Pilger 1956; Prat 1960; Clayton and Renvoize 1986; Watson and Dallwitz 1992 onwards). However, even within this context, the contents of similarly named tribes, subtribes and even generic composition differ greatly between different classifications (Van den Borre and Watson 1997).

In 1997, Van den Borre and Watson proposed a new classification for the Chloridoideae. In their study, cladistic and phenetic analyses were done on 166 genera in the subfamily, by using 120 selected leaf anatomical and morphological characters. This analysis did not give support for the Eragrosteae and Chlorideae,

**Table 1.1.** A summary of 16 classifications for the subfamily Chloridoideae (1956 onwards) in which tribes, subtribes or equivalent groups in the subfamily are indicated [Adapted from Van den Borre and Watson (1997)].

Author	Pilger (1956)	Tateoka (1957)	Prat (1960)	Stebbins and Crampton (1961)	Parodi (1961)	Jacques-Felix (1962)	Hubbard (1966)	Tutin (1980)
Subfamily	Eragrostoideae	Eragrostoideae	Chloridoideae	Eragrostoideae	Eragrostoideae	No subfamilies	Group XVIII	No subfamily
	Eragrosteae Eragrostinae Scleropogoninae Lycurinae Gainotiinae Sporobolinae Muhlenbergiinae Jouveae Phaenospermeae	Pappophoreae  Chlorideae    Spartineae	Group 3 Pappophoreae  Eragrosteae   Sporobolinae	Pappophoreae  Eragrosteae   Aeluropodeae  Chlorideae  Spartineae	Pappophoreae  Eragrosteae   Sporoboleae  Chlorideae  Spartineae	Pappophoreae  Eragrosteae Eragrostinae Aeluropinae  Sporoboleae  Chlorideae  Spartineae Leptureae Tripogoninae Lepturinae Zoysieae	Pappophoreae Orcuttieae Eragrostideae   Sporoboleae  Aeluropodeae  Chlorideae  Spartineae Leptureae  Zoysieae Pommereulleae Sphaerocarpeae	Pappophoreae  Eragrostideae       Spartineae Zoysieae
Excluded from the Chloridoideae				(Arundinoideae) Unioleae Aristideae	(Danthonieae) Uniola		(Centothecaeae) Uniolae (Danthonieae) Triodia	

Gould and Shaw (1983)	Dahlgren, Clifford and Yeo (1985)	Campbell (1985)	Clayton and Renvoize (1986)	Tzvelev (1989)	Wheeler, Jacobs and Norton (1990)	Watson and Dallwitz (1992 onwards)	Van den Borre and Watson (1997)	GPWG (2001)
	Chloridoideae	Chloridoideae	Chloridoideae	Within the Poaeae	Eragrostoideae	Chloridoideae	Chloridoideae	Chloridoideae
Pappophoreae	<i>Pappophorum</i> Group		Pappophoreae	Pappophoreae	Pappophoreae	Pappophoreae	Pappophoreae	Pappophoreae
Orcuttieae			Orcuttieae			Orcuttieae	Orcuttieae	Orcuttieae
Eragrosteae	<i>Eragrostis</i> group	Cynodonteae	Eragrostideae	Cynodonteae	Eragrostideae	Chlorideae s.l.	<i>Eragrostis</i> group I	Eragrostideae
			Eleusininae	Eleusinae		Triodeae	<i>Eragrostis</i> group II	
			Triodiinae	Eragrostinae			Triodiinae	
Sporoboleae			Monanthochloinae	Muhlenbergiinae				
			Uniolinae	Chloridinae				
Aeluropodeae	<i>Muhlenbergia</i> group	Aeluropodeae	Sporobolinae		Sporoboleae			
				Aeluropodeae				
Chlorideae	<i>Chloris</i> group		Cynodonteae		Chlorideae		<i>Chloris</i> group	Cynodonteae
			Pommereullinae				<i>Muhlenbergia</i> group	
			Chloridinae					
			Boutelouinae				<i>Zoysia</i> group	
			Zoysiinae					
Zoysieae		Zoysieae	Leptureae	Zoysieae	Zoysieae			Leptureae
					Triodieae			
Unioleae		Unioleae		Aristideae	Aristideae			
Aristideae								
		(Arundinoideae) Aristideae						

but instead indicate five different high-level groups, namely the *Zoysia* group, the *Chloris* group, the *Eragrostis* I and *Eragrostis* II groups and the *Muhlenbergia* group. Their analyses also showed strong support for the smaller tribes Pappophoreae Kunth, Orcuttieae Reeder and Triodeae Benth.

They, however, stated that many of the species need more thorough investigation and, therefore, their system is not a formal taxonomic classification (Van den Borre and Watson 1997).

The Grass Phylogeny Working Group (2001) has recently proposed a new classification for the grasses. Their results are based on the combined data of six molecular sequence data sets, morphological data and chloroplast restriction site data. The most significant changes in this new classification are the division of the traditional Bambusoideae and Arundinoideae and the expansion of the Pooideae. They divide the grasses into eleven previously published subfamilies and one new subfamily, Danthonioideae. Changes in the circumscription of traditionally recognized subfamilies have also been included. The circumscription of the Chloridoideae remains basically the same with the recognition of the tribes Cynodonteae, Eragrostideae, Leptureae Dumort., Orcuttieae and Pappophoreae, with the exception that the danthonioid genus *Centropodia* Rchb. and the danthonioid species *Merxmuellera rangei* (Pilg.) Conert are now also included in the Chloridoideae (GPWG 2001). The GPWG did, however, note that the current tribal classification conflicts with molecular data and could be modified in the future (see for example Hilu *et al.* 1999).

In this study the latest classification of the subfamily (GPWG 2001) will be followed, in that *Centropodia* and *Merxmuellera rangei* are included in the Chloridoideae (*incertae cedis*), but the tribal and subtribal delimitations will be according to Clayton and Renvoize (1986), as GPWG (2001) did not revise any tribal classifications (Table 1.2).



**Table 1.2.** Southern African representatives of the subfamily Chloridoideae, according to the classification of GPWG (2001) and tribes and subtribes according to Clayton and Renvoize (1986). Genera included in this study are highlighted in bold.

**Cynodonteae**

**Pommereullinae**

*Lintonia* Stapf

**Chloridinae**

*Brachyachne* (Benth.) Stapf

***Chloris* Sw.**

*Ctenium* Panz.

*Craspedorachis* Benth.

***Cynodon* Rich.**

***Enteropogon* Nees**

***Eustachys* Desv.**

***Harpochloa* Kunth**

***Microchloa* R.Br.**

***Polevansia* DeWinter**

***Rendlia* Chiov.**

*Schoenefeldia* Kunth

*Spartina* Schreb.

*Tetrapogon* Desf.

*Willkommia* Hack.

**Boutelouinae**

—

**Zoysiinae**

***Catalepis* Stapf & Stent**

*Monelytrum* Hack. ex Schinz

*Mosdenia* Stent

***Perotis* Aiton-**

***Tragus* Haller**

**Eragrostideae**

**Triodiinae**

—

**Uniolinae**

***Entoplocamia* Stapf**

***Fingerhuthia* Nees**

*Tetrachne* Nees

**Monanthochloinae**

—

**Eleusininae**

*Acrachne* Wright & Arn. ex Chiov.

***Bewsia* Gooss.**

*Brachychloa* S.M.Phillips

***Cladoraphis* Franch.**

*Coelachyrum* Hochst. & Nees

***Dactyloctenium* Willd.**

*Diandrochloa* DeWinter

*Dinebra* Jacq.

***Eleusine* Gaertn.**

***Eragrostis* Wolf**

*Leptocarydion* Stapf

***Leptochloa* P.Beauv.**

*Lophacme* Stapf

***Odyssea* Stapf**

*Oropetium* Trin.

***Pogonarthria* Stapf**

***Stiburus* Stapf**

***Trichoneura* Andersson**

*Tripogon* Roem. & Schult.

*Triraphis* R.Br.

**Sporobolinae**

***Sporobolus* R.Br.**

**Leptureae**

*Lepturus* R.Br.

**Orcuttieae**

—

**Pappophoreae**

***Enneapogon* Desv. ex P.Beauv.**

*Kaokochloa* DeWinter

***Schmidtia* Steud. ex J.A.Schmidt**

***Incertae cedis***

***Centropodia* Rchb.**

***Merxmuellera rangei* (Pilg.) Conert**

## 1.2.2 Morphology and Anatomy

Chloridoideae are anatomically distinct in the grass family. This subfamily is characterized by unspecialized, usually many-flowered spikelets with 1-(3)-nerved lemmas [although Jacobs (1987) found that more than 50% of the 152 species examined had more than three lemma nerves and, therefore, this feature might be too inconsistent to use in the delimitation of the subfamily], Kranz syndrome and associated anatomy, distinctive leaf-blade anatomy and equidimensional silica bodies. Within the subfamily little anatomical variation exists (Ellis 1987; Renvoize and Clayton 1992).

According to Linder *et al.* (1990), glandular structures may be bi- or multicellular. These bicellular structures are referred to as bicellular trichomes, microhairs (by anatomists) or salt glands (by physiologists). The trichomes present in Poaceae, usually referred to as microhairs, are a feature of the upper and lower epidermis of leaf blades, paleas, lodicules and lemmas (Terrell and Wergin 1981). They are relatively thin walled and small in size (25-70  $\mu\text{m}$ ). This distinguishes them from grass macrohairs and 'prickles' that are unicellular, thick walled and larger (Amarashinge and Watson 1988). These microhairs are present throughout Poaceae, with the exception of the subfamily Pooideae. The occurrence of multicellular glands in Poaceae is rare (Johnston and Watson 1976; Linder *et al.* 1990).

Chloridoid microhairs are unique in their shape, with relatively short, broad and thick walled cap cells (Johnston and Watson 1976). They are the only type of microhairs for which the function has been determined, namely the secretion of salt from two-celled salt glands (Oross and Thomson 1984). The secretion of ions by such specialized salt glands, is a mechanism for regulating the mineral content of various halophytic plants (Lipschitz and Waisel 1974), and enables the rehabilitation of soils damaged by poor irrigation practice (Chapman and Peat 1992). Salt glands have a concentrated occurrence in the subfamily Chloridoideae and have been observed in various species (Lipschitz and Waisel 1974). It is proposed that salinity tolerance is highly correlated to leaf  $\text{Na}^+$  and  $\text{Cl}^-$  exclusion, which is in turn correlated to leaf salt gland ion secretion rates. The tolerance of high salt levels is also associated with accumulation of the proposed compatible solute glycinebetaine,

which reaches effective levels for cytoplasmic osmotic adjustment in salt tolerant grasses only (Marcum 1999).

The Kranz syndrome is an example of structure related to function with the features of the photosynthetic carbon fixation process being correlated with leaf anatomy (Ellis 1977).  $C_4$  plants are grouped into two distinct groups, depending on the type of reaction for  $C_4$  acid decarboxylation that occurs in the bundle sheath cells of the leaf blades. The groups are aspartate and malate formers (Downton 1970; Brown and Gracen 1972; Hatch *et al.* 1975). Chloridoideae are aspartate formers. This implies mesophyll cells that are arranged radially and a double bundle sheath with large, specialized chloroplasts localized in the outer bundle sheath (Ellis 1984a). Two subtypes (structural/biochemical variants) are recognized for the aspartate pathway, namely NAD-ME (nicotinic amide adenine dinucleotide-malic enzyme) and PCK (phosphoenol pyruvate carboxylase) (Ellis 1977) (Table 1.3). Ellis *et al.* (1980) found NAD-ME grasses to dominate in arid areas with low and unpredictable rainfall and PCK grasses were found in generally moist habitats, with less than 350mm rainfall per year. In areas where the rainfall exceeds this amount, the PCK grasses are found primarily in more specialized habitats. Therefore, NAD-ME species would probably be more successful in dry areas, whereas PCK has intermediate moisture requirements (Ellis *et al.* 1980).

Chloridoideae are accepted to be a  $C_4$  subfamily. This enables these plants to photosynthesize more efficiently in high temperatures and high light intensities (Clayton 1983). The only exceptions are *Eragrostis walterii* Pilg. and *Merxmüllera rangei*, which have been found to be  $C_3$  plants (Ellis 1982, 1984a).

A small minority of chloridoideae is known as 'resurrection grasses' (Chapman and Peat 1992). This implies that these grasses have foliage that is able to revive after dehydration (Chapman 1992). Resurrection plants in southern Africa include the following: *Brachyachne patentiflora* (Stent & Rattray) C.E.Hubb., *Eragrostis hispida* K.Schum., *E. nindensis* Ficalho & Hiern, *E. paradoxa* Launert, *Microchloa caffra* Nees, *M. kunthii* Desv., *Oropetium capense* Stapf, *Sporobolus festivus* Hochst. ex A.Rich., *S. lampranthus* Pilg., *S. stapfianus* Gand. and *Tripogon minimus* (A.Rich.) Hochst. ex Steud. (Gaff and Ellis 1974). Other grass groups containing resurrection grasses are the Arundinoideae (*Micraira* F.Muell.) and Pooideae (*Poa*

**Table 1.3.** Characteristics of the C<sub>4</sub> aspartate subtypes (Ellis 1977).

NAD-ME	PCK
Centripetal chloroplasts	Centrifugal chloroplasts
Granal chloroplast membranes	Distinct grana on the chloroplast membranes
Mitochondria centripetal	Centrifugally arranged mitochondria
Malate decarboxylate as decarboxylating enzyme	Cytosolic phosphoenol pyruvate carboxylase as decarboxylating enzyme

*bulbosa* L.) (Lazarides 1992). Other plant groups in which this phenomenon occurs include some ferns and sedges (Chapman 1992).

### 1.2.3 Biogeography

The subfamilies and tribes of Poaceae are rather uniformly distributed across the continents in broad regions corresponding to climatic zones. The genera, however, being younger in age, are usually restricted to single continents (Gibbs Russell *et al.* 1990).

Chloridoideae and Panicoideae are the most diverse subfamilies in the open environments of the warm tropical and subtropical regions, which includes southern Africa (Renvoize and Clayton 1992; Davis and Soreng 1993).

Hartley and Slater (1960) and Hartley (1964) concluded that Chloridoideae occur mainly in arid regions that are characterized by high winter temperatures and summer or sporadic rainfall. This centers their distribution in the seasonally dry, or arid to semi-arid areas of the southern Hemisphere tropics, most notably southern Africa and Australia (Jacobs 1987). In these areas the photosynthetic system and associated morphological and anatomical adaptations it provides, presents these grasses with a competitive advantage (Jacobs 1987).

The close relationship between climatic factors and the distribution of the subfamily Chloridoideae, suggests that the subfamily is an old one, and that the present distribution of the subfamily is not greatly affected by historical factors (Hartley and Slater 1960). Particular tribes and subtribes and especially certain

genera have restricted distributions, but the subfamily as a whole appears to be spread throughout the world in those parts to which they are physiologically and climatically adapted. Furthermore, many of the smaller genera, as well as individual species, have markedly distinct distributions. These factors support the antiquity of the subfamily (Hartley and Slater 1960).

The great number of chloridoids (number of taxa and number of endemics) in tropical Africa led Hartley and Slater (1960) to suggest an African origin for the chloridoids. From Africa the subfamily then spread to other regions of the world.

Factors supporting an African origin for Chloridoideae include the following:

- The subfamily is taxonomically related to Panicoideae, which are believed to have originated in the east Africa-Madagascar region (Hartley 1958a).
- As is the case with Panicoideae, there appears to be a major center of endemism of the subfamily in Madagascar, with certain genera and species being confined to this area.
- Many of the larger tribes and subtribes have centers of high specific differentiation on the African continent.
- The subtribe Eragrostinae Ohwi [according to Pilger's classification (1956)] exhibits many characters, which are regarded as primitive in Poaceae. This is believed to be the progenitor of the other tribes and subtribes in the subfamily. Factors corroborating this hypothesis are the strong representation of the subtribe on the African continent, monographs on some of the genera suggesting an African origin, the primitive characters of some species in the tribe, as well as apparent affinities to other grass tribes found in Africa (Hartley and Slater 1960).

The subfamily Chloridoideae comprises approximately 1360 species, in  $\pm 150$  genera of which 100 are mono- or ditypic (Van den Borre and Watson 1997; Hilu and Alice 2001), which according to Clayton and Renvoize (1986) indicates that the subfamily has been dominated by strong adaptive radiation into specialized, often stressful habitats.

The largest genus in the subfamily is the genus *Eragrostis*. This is also one of the most widespread genera in the world (Carnahan and Hill 1961). The genus

exhibits almost a full range of anatomical and morphological variation found in the subfamily, of which it contains approximately 25% of the species (Van den Borre and Watson 1994).

The grass family includes 194 genera and 912 species in southern Africa and the subfamily Chloridoideae is represented by 50 genera and 230 species in southern Africa (Fish 2000) (Table 1.4).

## 1.3 Cytogenetics

Any data that can differentiate species are of taxonomic significance (Stace 1991). For this reason, Raven (1975) regarded cytogenetics as an important element in the evaluation of relationships and in the determination of phylogenetic evolution in the angiosperms.

Cytogenetics includes studies dealing with observations of chromosomal pairing or meiotic behavior. Cytotaxonomy refers to the use of these characteristics and others, such as chromosome number and chromosome morphology, as data for classification (Jones and Luchsinger 1987).

Cytogenetically, the grasses are so diverse that it raises many problems for those attempting to divide them into discrete species. Some 80% of them have a polyploid chromosome number (Clayton 1978), and the occurrence of polyhaploidy, the reversion of polyploids back to the diploid state, has been demonstrated for various genera, the best known example being the *Dicanthium* Willem.-*Bothriochloa* Kuntze complex (Harlan and De Wet 1963; De Wet 1968; De Wet and Harlan 1970). Apomictic swarms are common and over 4000 hybrids have been recorded (Freeling 2001).

In 1931, Avdulov led the way with the use of cytogenetic features in establishing phylogenetic and systematic relationships among species and genera of grasses. By using cytogenetic information, Avdulov (1931) proposed a phylogenetic subdivision of Poaceae and the publication, *Karyo-systematic investigations in the family Gramineae* (originally published in Russian), was the start of a new age in the classification of the grasses (Jauhar 1993). Avdulov indicated that the basic chromosome number,  $x$ , as well as chromosome morphology could serve as a basis



**Table 1.4.** Chloridoid genera present in southern Africa and their worldwide occurrence (Adapted from Fish 2000).<sup>1</sup>

GENUS	SPECIES IN SOUTHERN AFRICA	SPECIES WORLDWIDE
<i>Acrachne</i>	1	3
<i>Bewsia</i>	1	1 <sup>♣</sup>
<i>Brachyachne</i>	1	10
<i>Brachychloa</i>	2	2 <sup>•</sup>
<i>Catalepis</i>	1	1 <sup>◇</sup>
<i>Centropodia</i>	2	4
<i>Chloris</i>	8	55
<i>Cladoraphis</i>	2	2 <sup>◇</sup>
<i>Coelachyrum</i>	1	6
<i>Craspedorachis</i>	2	2-5
<i>Ctenium</i>	1	20
<i>Cynodon</i>	8	10
<i>Dactyloctenium</i>	4	13
<i>Diandrochloa</i>	2	7
<i>Dinebra</i>	1	3
<i>Eleusine</i>	4	9
<i>Enneapogon</i>	7	30
<i>Enteropogon</i>	4	6-17
<i>Entoplocamia</i>	1	1 <sup>♣</sup>
<i>Eragrostis</i>	90	350
<i>Eustachys</i>	1	10
<i>Fingerhuthia</i>	2	2

<sup>1</sup> ♣ Occur only in Africa

• Occur only in southern Africa

◇ Occur only in South Africa

□ This single species of the previously danthonioid genus *Merxmuellera* is now included in the Chloridoideae (GPWG 2001)

<i>Harpochloa</i>	1	2
<i>Kaokochloa</i>	1	1°
<i>Leptocarydion</i>	1	1°
<i>Leptochloa</i>	7	30
<i>Lepturus</i>	1	15
<i>Lintonia</i>	1	2*
<i>Lophacme</i>	1	2°
<i>Merxmuellera rangeri</i> <sup>p</sup>	1	1°
<i>Microchloa</i>	3	4
<i>Monelytrum</i>	1	1°
<i>Mosdenia</i>	1	1°
<i>Odyssea</i>	1	2*
<i>Oropetium</i>	1	3-6
<i>Perotis</i>	3	10
<i>Pogonarthria</i>	3	4*
<i>Polevansia</i>	1	1°
<i>Rendlia</i>	1	1*
<i>Schmidtia</i>	2	2
<i>Schoenefeldia</i>	1	2
<i>Spartina</i>	1	16
<i>Sporobolus</i>	38	160
<i>Stiburus</i>	2	2°
<i>Tetrachne</i>	1	1
<i>Tetrapogon</i>	1	5
<i>Tragus</i>	4	7
<i>Trichoneura</i>	2	7
<i>Tripogon</i>	1	30
<i>Triraphis</i>	5	7
<i>Willkommia</i>	3	4

for the broad divisions of Poaceae and for the grouping of genera into tribes and subtribes, and species into genera and sections. He also regarded the increase in absolute size of chromosomes as a major trend in grass evolution (Jauhar 1993).

This classification has been found to be consistent with anatomy and geographical distribution (Jauhar 1993).

The basic chromosome number of the Poaceae is the subject of large disputes. Mehra *et al.* (1968), as well as Sharma (1979), support  $x = 6$  as the basic ancestral chromosome number. Flovik (1938) is of the opinion that  $x = 5$  represent the basic chromosome number from which higher numbers were derived through gain aneuploidy and/or polyploidization. Stebbins (1985) suggested that  $x = 5, 6,$  and  $7$  could all have been present in the now extinct complex that was the ancestor of the present day Poaceae.

Basic chromosome numbers in the Chloridoideae are  $x = 9$  and  $10$ . Cytogenetically the group has been investigated to a great extent, and especially in southern Africa, with its large concentration of Chloridoideae, there have been various cytogenetic studies (Appendix A and B). The basic chromosome number of  $x = 9$  occurs in the lowest frequency of the two main basic chromosome numbers and is proposed to have originated by aneuploid or dysploid reduction from  $x = 10$ . Polyploidy is frequent in the group with more than 70% of southern African Chloridoideae reported having somatic chromosome numbers of more than  $2x$ .

## 1.4 Apomixis

Plants reproduce sexually, asexually by means of seed (agamospermy) or vegetatively (McWilliam 1964). Apomixis includes the various developmental pathways by which a plant reproduces asexually by means of seed formation (Nogler 1984; Asker and Jerling 1992). Apomictic reproduction can vary from obligate (strictly asexual) to facultative (partially sexual) with differing degrees of apomixis between and within species and genotypes (Hanna and Bashaw 1987).

Apomixis is known from more than 300 species in more than 35 different plant families (Hanna and Bashaw 1987). The phenomenon is predominant in the families Asteraceae, Rosaceae L. and Poaceae, which together comprise approximately 10% of the angiosperm species (Nogler 1984).

In Poaceae, the phenomenon has been reported in more than 125 species representative of most of the tribes (Bashaw and Hanna 1990). The replacement of

a sexual reproduction system by an asexual reproduction system has been known in the grass family since 1932 when Müntzing first identified apomixis in *Poa* L. Apospory, the development of unreduced embryo sacs from somatic cells of the ovule (Asker 1979), is the most common mechanism of apomixis in grasses and accounts for more than 95% of apomictic grasses (Bashaw and Hanna 1990).

Apomixis has been observed in the following chloridoid genera: *Bouteloua* Lag. (Harlan 1949; Freter and Brown 1955; Mohammed and Gould 1966; Bierzychudek 1985); *Chloris* (Brown and Emery 1958; Hutton 1961); *Eragrostis* (Streetman 1963a, b; Voight 1971; Voight and Bashaw 1972, 1976; Brix 1974; Vorster and Liebenberg 1984; Voight *et al.* 1992); *Eustachys* (Strydom and Spies 1994a); *Fingerhuthia* (Brown and Emery 1958); *Harpochloa* (Strydom and Spies 1994a); *Hilaria* Kunth (Brown and Emery 1958); *Rendlia* (Strydom and Spies 1994a) and *Schmidtia* (Brown and Emery 1958).

## 1.5 Hybridization

Great importance is attached to hybridization in the evolution of Poaceae (Tzvelev 1972, 1975). The primitive species of many genera, as well as tribes, could have originated in association with the descent of grasses from various mountain systems to the plain, followed by subsequent cross-migrations. Taxa that have undergone hybridization have a much greater possibility for genetic recombination as opposed to their primary diploids and also exhibit a greater degree of despecialization, resulting in greater evolutionary possibilities (Tzvelev 1975). Natural hybridization is common in the grasses and in populations where hybrids occur the variability is increased. The high levels of genetic variability allow grasses to colonize new habitats (Gibbs Russell and Spies 1988; Spies and Gibbs Russell 1988).

The diversity level in hybrid taxa will be affected by factors such as the number of parental species involved in their origin, the degree of genetic variation between the parental taxa, type of mating system, species age and historical circumstances (Morrell and Rieseberg 1998).

The process of natural hybridization between closely related taxa results in the production of a genetic genotype with a new genetic composition, which might or

might not be favorable (Rieseberg *et al.* 1996). This implies that hybridization between lineages, along with the process of mutation, provides the basis for adaptive evolution (Anderson 1949). The process of hybrid speciation has the potential of fixing adaptive genetic combinations in the population (Grant 1981).

Polyploidy is nearly always associated with hybridization (De Wet 1987).

## 1.6 Polyploidy

Polyploidy and the process of plant domestication are two general features of plant evolution. Chromosome doubling renders the resultant plants with greater stress tolerance, delayed reproduction, a longer life span, greater defense against herbivores and pathogens, lower reproductive effort, larger seeds, etc. Many of these properties are important in the domestication of plants (Hilu 1993). Polyploids are also known to exhibit a greater colonizing ability than their diploid progenitors (Ehrendorfer 1980) and, thus, have a wider geographical distribution. The high incidence of polyploidy in the grass family can be attributed to the success it presents under various ecological conditions and can be explained by genomic hybridity and chromosome multiplicity (Tal 1980; Levin 1983).

Polyploids can be classified into two basic groups in terms of time of origin (Grant 1963; Goldblatt 1980):

- Neopolyploid plants have chromosome numbers that are multiples of the basic chromosome numbers found in their diploid ancestors.
- Paleopolyploid species have re-diploidized and have high secondary basic chromosome numbers.

Polyploidy has a higher frequency in perennial plants than in annuals (Stebbins 1971) due to the fact that the perennial habitat provides a selective advantage to polyploidy. In a perennial habitat, auto- and allopolyploids have a better chance to recover from sterility after polyploidization, because there is no need for immediate fitness (Hilu 1993).

Polyploid plants have higher genetic variability and higher heterozygosity than their diploid parents. There is increasing evidence that polyploids may have higher levels of self-compatibility than their related diploid ancestors. Asexual reproduction,

usually absent in diploid taxa, shows an increase in consequent polyploids (Thompson and Lumaret 1992). Furthermore, inbreeding depression, traditionally counteracting the evolution of selfing, may be reduced by polyploidy. The reason for the success of polyploids may be sought in greater enzyme diversity, wider physiological tolerances and increased flexibility in the reproductive system (Thompson and Lumaret 1992).

Polyploidy has a very high frequency in the Chloridoideae (more than 60%) and especially in the southern African Chloridoideae where more than 70% of the known chromosome reports are polyploid (Appendix A, B).

## 1.7 Molecular markers

It is widely accepted that molecular phylogenetic studies should include multiple markers to guarantee that the resultant gene tree is an accurate representation of the species phylogeny. Relying upon a single gene for phylogenetic reconstruction can be problematic due to phenomena such as introgression, concerted evolution and mistaken orthology (Doyle 1992).

The markers that are most widely used in plants, are the *ITS* (internal transcribed spacer) regions of nuclear ribosomal DNA, and various regions in the chloroplast genome (Kim *et al.* 1999).

The *ITS* nuclear ribosomal DNA region has proven useful for elucidation of relationships at various taxonomic levels (e.g. Baldwin *et al.* 1995; Kim *et al.* 1996; Cox *et al.* 1997; Kelly 1998; Freudenstein 1999; Alan and Porter 2000; Hao *et al.* 2000; Torrecilla and Catalan 2002). This region consists of components that evolve at different rates (Ainouche and Bayer 1997) and the intergenic spacer regions are particularly suitable (Baldwin *et al.* 1995) being flanked by conserved regions that make amplification with a set of universal primers possible (White *et al.* 1990). Furthermore, the region is repeated many times in the nuclear genome (Rogers and Bendich 1987).

Chloroplast DNA sequencing has been widely used in plant phylogeny and genes such as *atpB*, *matK*, *ndhF* and *rbcl* are popular choices (for example Chase *et al.* 1993; Kim and Jansen 1995; Gaut *et al.* 1997; Hoot and Douglas 1998; Hilu

and Alice 1999; Oxelman *et al.* 1999; Savolainen *et al.* 2000; Smith 2000; Cameron *et al.* 2001). Most chloroplast coding regions do, however, not evolve rapidly enough to resolve relationships at lower taxonomic levels (Doebley *et al.* 1990; Gaut *et al.* 1992). Non-coding chloroplast regions (introns and intergenic spacers) evolve more rapidly than their coding regions by accumulating insertion/deletions (indels) at a rate almost equal to that of nucleotide substitutions (Curtis and Clegg 1984; Wolfe *et al.* 1987; Zurawski and Clegg 1987; Clegg and Zurawski 1992). This is because these regions are less functionally constrained (Clegg *et al.* 1994) and, therefore, useful in studies below family level (Gielly and Taberlet 1994). Examples of such non-coding regions include *rpoC1* (intron) (Liston 1992; Wallace and Cota 1996; Downie *et al.* 1998); *rpl16* (intron) (Kelchner and Clark 1997; Renner 1999; Applequist and Wallace 2000); *atpB-rbcL* spacer (Manen and Natalie 1995; Hoot and Douglas 1998; Schwarzbach and Ricklefs 2000) and the *trnL-F* intron, spacer region (Bayer and Starr 1998; Bakker *et al.* 1999; Bellstedt *et al.* 2001; Liede and Täuber 2002).

Although the need for multiple data sets to reliably determine phylogenies is apparent, it is also becoming clear that different data sets may in fact possess different phylogenetic histories (Wendel and Doyle 1998). This incongruence may be caused by various phenomena such as hybridization, introgression, conversion, evolutionary rate heterogeneity, lineage sorting, reticulation, concerted evolution or mistaken orthology (Rieseberg and Soltis 1991; Doyle 1992; Maddison 1997; Wendel and Doyle 1998; Sang and Zhong 2000).

Various methods have been developed to test whether the observed incongruence between different data sets is statistically robust (Larson 1994; de Queiroz *et al.* 1995; Mason-Gamer and Kellogg 1996). Some of the significance tests of heterogeneity are the following:

- 'The homogeneity test (Farris *et al.* 1995a, b) tests the null hypothesis that characters are randomly distributed between data sets with regard to their phylogenetic informativeness. If two data sets are highly incongruent then the sum of their minimal trees should be significantly shorter than that of the sum of tree lengths from random partitions of the combined data and the null hypothesis will be rejected'. This is known as the ILD (incongruence length difference) test (Mason-Gamer

and Kellogg 1996) and implemented as partition homogeneity testing in PAUP\*.

- 'The Wilcoxon signed-rank test (Wilcoxon *et al.* 1970; Templeton 1983) can be used to assess whether data provide significantly less support for a specified alternative topology compared to the most parsimonious topology. This method tests whether either data set, when reanalyzed under a constraint that accommodates the topological conflict presented by the other data set, produces a set of changes in the lengths of individual characters whose directionality is greater than expected by chance alone'.

Whether or not data sets are congruent is usually taken as an indication of whether they can be combined or not. This is the methodology on which the conditional combination approach is based with both separate and combined analyses being conducted. Homogeneity tests are conducted to measure the significance of incongruence levels. If the null hypothesis of data set homogeneity cannot be rejected then the data sets can be combined. If the null hypothesis is rejected, the source of heterogeneity needs to be investigated (Bull *et al.* 1993; Bremer 1996; Huelsenbeck *et al.* 1996; Mason-Gamer and Kellogg 1996; Johnson and Soltis 1998; Wiens 1998).

Two other approaches to analyzing multiple data sets are the total evidence (Kluge 1989; Kluge and Wolf 1993) and congruence (Miyamoto 1985; Miyamoto and Fitch 1995) approaches. With the total evidence approach phylogenetic analysis is done of all available evidence in combination. With the congruence approach, data sets are analyzed separately and after the separate analyses, consensus methods are used to identify points of agreement between the data sets.

### **1.7.1 Chloroplast genome**

Chloroplast DNA is independent of polyploidy, an event which has its highest known incidence in the grass family. Autopolyploidy, where genomes are duplicated, does not affect the chloroplast genome (Ogihara and Tsunewaki 1982). With allopolyploids and segmental allopolyploids, however, where hybridization occurs, the mode of chloroplast inheritance would play a role. Chloroplasts and



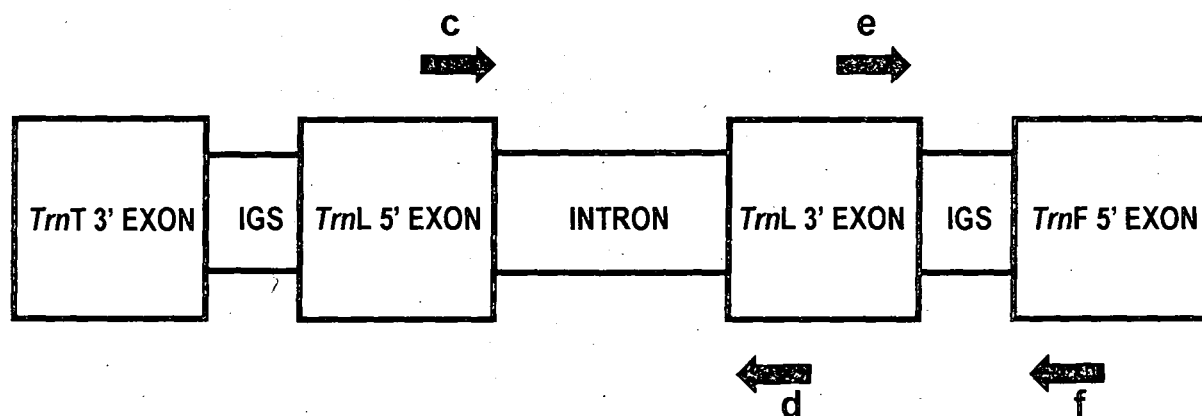
chloroplast DNA are inherited maternally (reviewed in Kirk and Tilney-Bassett 1978; Sears 1980; Whatley 1982; Mogensen 1996). In some angiosperm families paternal inheritance does occur and, in the case of biparental inheritance, the maternal parent usually donates most of the cytoplasm (Hilu 1987).

In most respects the molecular evolution of the chloroplast genes reflects that of the nuclear genes. Chloroplast protein-coding genes, however, evolve approximately five times slower than the nuclear genes (Wolfe *et al.* 1987, 1989). The non-coding regions of the chloroplast genome tend to evolve more rapidly than the coding region (Wolfe and Sharp 1988). Two major factors in distinguishing between chloroplast and nuclear genome evolution are the lack of transposon activity associated with the chloroplast genome and the apparent lack of recombinational potential. Because biparental inheritance is rare for the chloroplasts and intraspecific variance is low, recombinational processes do not play an important role in chloroplast sequence evolution (Clegg and Zurawski 1992).

The mutations in the chloroplast genome include point mutations (substitutions) and rearrangements. Major rearrangements include inversions and indels of genes and introns. Minor rearrangements consist of smaller indels (1-1000bp). These minor rearrangements are the most common mutation and occur mostly in the non-coding regions of the intergenic spacers and introns (Palmer 1985). According to Soltis *et al.* (1989, 1990a), these mutations may provide essential information in studies of closely related taxa.

The combination of adjacent regions that evolve at different rates may increase the phylogenetic range over which the sequences can be useful. The more slowly evolving regions provide support for older divergences, while the more rapidly evolving regions, usually non-coding chloroplast DNA sequences, which exhibit higher mutation rates (for example Clegg and Zurawski 1992), provide resolution at intrageneric as well as intraspecific levels (Taberlet *et al.* 1991; McDade and Moody 1999).

The non-coding chloroplast region investigated in this study comprises the *trnL* intron (the intron of the transfer RNA leucine-UAA gene), *trnL* 3' exon, and the *trnL-F* spacer (the intergenic spacer between *trnL* exon and the transfer RNA phenylalanine-GAA gene) (hereafter referred to as the *trnL-F* region) (Figure 1.1).



**Figure 1.1.** *TrnL-F* region in the chloroplast genome. IGS indicates the intergenic spacer region between the *trn* genes. The primers used are designated as c–f and the positions in which they approximately anneal are also indicated. Regions are not drawn to scale.

This region has been investigated at various levels:

- Generic – *Disa* (Bellstedt *et al.* 2001); *Miscanthus* (Hodkinson *et al.* 2002a); *Pelargonium* (Bakker *et al.* 1999).
- Tribal – Andropogoneae (Hodkinson *et al.* 2002b); Gnaphalieae (Bayer *et al.* 2000); Melanthieae (Zomlefer *et al.* 2001).
- Subfamily – Zygophylloideae (Sheahan and Chase 2000).
- Family – Acanthaceae (McDade and Moody 1999); Palmae (Baker *et al.* 1999); Restionaceae (Eldenas and Linder 2000).

## 1.7.2 Nuclear genome

*ITS* sequence analysis has proven to be an effective tool for testing phylogenetic relationships within and among closely related genera (e.g. Baldwin *et al.* 1995; Kim *et al.* 1996). Variation in the internal transcribed spacer region has also been used in higher-level relationships, for example within tribes in the Poaceae (Hsiao *et al.* 1995a, b), at familial level (e.g. Downie and Katz-Downie 1996) and even among families of flowering plants (Hershkovitz and Zimmer 1996).

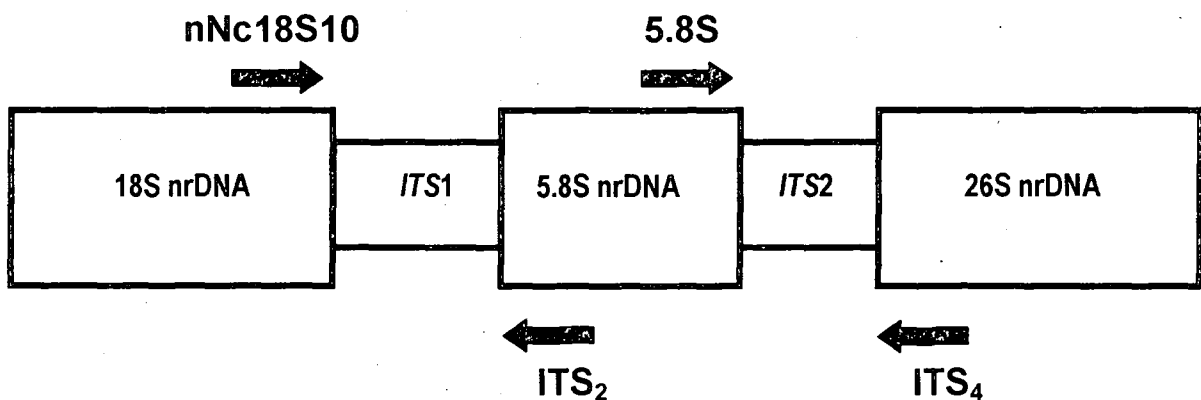
The predominance of rRNA genes in nature and the considerable evidence that the repeat unit consists of regions that have different rates of sequence

divergence explains the phylogenetic utility of *ITS* (Wojciechowski *et al.* 1993).

Ribosomal repeats exhibit a pattern of concerted evolution in which the repeats present in one array are more closely related to one another than to repeats in arrays on other chromosomes (Copenhaver *et al.* 1995) or repeats in other taxa (Arnheim 1983). Processes such as unequal crossing over (Smith 1976), gene conversion (Arnheim 1983) and biased gene conversion (Hillis *et al.* 1991) drive this concerted evolution.

Nucleotide differences arising from mutation are usually corrected by homogenization (Leskinen and Alström-Rapaport 1999). *ITS* region polymorphisms may, however, occur when concerted evolution among paralogous genes does not homogenize the sequences fast enough for them to behave as a single orthologous unit, thereby impairing the ability to infer a species phylogeny from the gene phylogeny (Sanderson and Doyle 1992). In angiosperms this can be caused by polyploidy, hybridization, agamospermy and slow rates of concerted evolution across nuclear ribosomal DNA loci on non-homologous chromosomes (Buckler *et al.* 1997).

The nuclear *ITS* region includes the internal transcribed spacer regions *ITS*1 and *ITS*2 and the 5.8S rDNA (Figure 1.2).



**Figure 1.2.** The internal transcribed spacer regions (*ITS*) of nuclear ribosomal DNA. The primers used are designated as nNc18S10, *ITS*<sub>2</sub>, *ITS*<sub>4</sub> and 5.8S and the positions in which they approximately anneal are also indicated. Regions are not drawn to scale.

## 1.8 Phylogeny

Understanding evolutionary processes involves a detailed knowledge of evolutionary pattern. Phylogeny reconstruction is, therefore, essential to all evolutionary biology. Especially over the last 20 years, more and more methods have been developed for phylogenetic analysis, which is in large part attributed to the increasing availability of computing power (Kellogg and Watson 1993).

In recent years phylogenetic studies of the grass family, especially at the molecular level, have greatly increased in number [Hamby and Zimmer 1988 (18S ribosomal DNA); Doebley *et al.* 1990 (plastid gene *rbcL*); Doyle *et al.* 1992a (chloroplast DNA inversions); Hamby and Zimmer 1992 (nuclear ribosomal 18S and 26S RNA); Davis and Soreng 1993 (plastid DNA restriction site data); Cummings *et al.* 1994 (plastid gene *rpoC2*); Nadot *et al.* 1994 (plastid gene *rps4*); Barker *et al.* 1995 (plastid gene *rbcL*); Clark *et al.* 1995 (plastid gene *ndhF*); Duvall and Morton 1996 (plastid gene *rbcL*); Liang and Hilu 1996 (plastid gene *matK*); Mathews and Sharrock 1996 (nuclear phytochrome gene family); Morton *et al.* 1996 (nuclear gene *Adh*); Gaut *et al.* 1997 (nuclear gene *Adh* and plastid gene *rbcL*); Kellogg 1998a (combination of published results on anatomy, morphology, plastid genes *rbcL*, *ndhF*, *rpoC2* and *rps4*, chloroplast restriction site data, nuclear ribosomal RNA and the nuclear genes *phyB* and *GBSSI*); Mason-Gamer *et al.* 1998 (nuclear gene *GBSSI*); Soreng and Davis 1998 (combination of morphological, anatomical, chromosomal, biochemical and structural chloroplast features with chloroplast restriction site data); Barker *et al.* 1999 (grass-specific insert in plastid gene *rpoC2*); Gaut *et al.* 1999 (nuclear gene *Adh*); Hilu and Alice 1999 (plastid gene *matK*); Hilu *et al.* 1999 (plastid gene *matK*); Hsiao *et al.* 1999 (nuclear *ITS* regions); Clark *et al.* 2000 (combination of plastid genes *ndhF* and *rbcL* with nuclear gene *phyB*); Mathews *et al.* 2000 (nuclear gene *phyB*); Zhang 2000 (plastid gene *rp16* intron)].

These studies mostly support a monophyletic PACC (Panicoideae-Arundinoideae-Chloridoideae-Centothecoideae) assemblage *sensu* Davis and Soreng (1993). Hilu and Wright (1982) first resolved this monophyletic group in a phenetic analysis of 85 taxa with morphological and anatomical data. Further morphological and anatomical evidence for this grouping includes immunological and prolamin studies (Hilu and Esen 1988, 1990, 1993; Esen and Hilu 1989). Very

few of these studies have, however, been able to resolve relationships within the PACC assemblage.

The study done by Hsiao *et al.* (1999) resolved relationships within the PACC clade and placed Panicoideae as the sister to Chloridoideae. This supports their sharing of an arundinoid-like common ancestor as proposed by Clayton (1981). Further evidence indicating a close relationship between these two subfamilies is characteristics such as lodicule structure, length of the embryo, nature of the first leaf of the seedling, C<sub>4</sub> photosynthetic pathways and characteristic anatomy, as well as chromosome number and size (Hilu and Johnson 1991).

Studies done by the Grass Phylogeny Working Group (2001), involved the combined data of eight data sets: morphological [a varied set of characters representing variation in macromorphology, biochemistry and anatomy, as well as structural variants for example inversions and deletions in the plastid and nuclear genes), restriction site variation throughout the plastid genome, sequences of *ndhF*, *rbcL*, *rpoC2* (plastid genome) and *ITS*, *phyB* and *waxy* (nuclear genome)]. Representatives were sampled from all recognized subfamilies, constituting 57 exemplar grass genera (GPWG 2001). In this analysis the two clades PACC and BOP (Bambusoideae-Oryzoideae-Pooideae) were resolved as sister groups. Within PACC, Chloridoideae, Centothecoideae and Panicoideae were resolved as monophyletic, with Arundinoideae being paraphyletic. According to this, Chloridoideae are nested in the Arundinoideae (GPWG 2001). Subsequently with the new subfamilial classification of the GPWG (2001) the BOP clade has become the BEP (Bambusoideae-Ehrhartoideae-Pooideae) clade and the PACC clade has become the PACCAD (Panicoideae-Arundinoideae-Centothecoideae-Chloridoideae-Aristidoideae-Danthonioideae) clade.

## 1.9 Aim of the study

Hybridization and subsequent polyploidy have a high frequency in the grasses. These phenomena, which lead to reticulate evolution, may be partly responsible for large-scale homoplasy in Poaceae (Van den Borre and Watson 1997).

The main aim of this study was to investigate southern African Chloridoideae on cytogenetic and molecular levels.

Chromosome numbers and meiotic behavior of southern African Chloridoideae will be determined. The incidence of polyploidy will also be investigated. By making a worldwide and southern African comparison of known chromosome number reports, the incidence of ancestral polyploidization (paleopolyploidy) in the subfamily will be examined. By studying the types of polyploids present in the subfamily, the frequency of hybridization can be assessed.

The phylogeny of southern African Chloridoideae will be investigated based on two genomic regions, namely nuclear ribosomal *ITS* and chloroplast *trnL-F*. The monophyly of the subfamily, constituent tribes and genera in the subfamily will be investigated. Due to the different modes of inheritance of the two regions investigated, incongruent phylogenies may indicate the existence of putative hybrids.

Lastly it is the aim of this study to examine whether groups with a high known occurrence of polyploidy and hybridization can be successfully examined on a phylogenetic level and whether adequate resolution can be obtained.

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**CHAPTER 2**

**Materials and Methods**

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One hundred and thirty specimens, representing 27 genera and 58 species of southern African Chloridoideae, were investigated during this study. Studies done on these specimens include cytogenetic investigation by means of squashes, where anthers were dissected and meiotic cells were investigated, and DNA sequencing of the *ITS* regions in the nuclear genome and the *trnL-F* region in the chloroplast genome. These results were used in a phylogenetic study to determine relationships, to investigate phenomena such as polyploidy and hybridization, and the effects of these phenomena on phylogenetic reconstruction and resolution. The computer program PAUP\* (version 4.0b10) was used for phylogenetic analyses.

## 2.1 Materials

Voucher herbarium specimens were collected in the veld and stored in the Geo Potts Herbarium Bloemfontein (BLFU) or in the National Herbarium, Pretoria (PRE). The plants collected are listed in Table 2.1.

DNA Molecular Marker VI (pBR328 DNA cleaved with a mixture of *Bgl*I and *Hinf*I) (Boehringer Mannheim cat. no. 1062590); *Thermus aquaticus* (*Taq*) Super Therm DNA polymerase with 10X Buffer (Southern Life Biotechnology LPI-801, LPI-455); sequencing primers (Integrated DNA Technology, custom synthesized) and the ABI (Applied Biosystems Inc.) PRISM BigDye Terminator v3.0 Ready Reaction Cycle Sequencing Kit and 5X [Tris-HCl (pH9.0) + MgCl<sub>2</sub>] dilution solution (ABI, product no. 4390242, 4305605) were used during this study. All other chemicals used during the study were of either analytical or electrophoretic grade.

Data on some of the *ITS* and *trnL-F* sequences were obtained from Genbank. These taxa are listed in Table 2.2.



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**Table 2.1.** List of localities and voucher herbarium numbers of specimens investigated in this study. (C) indicates chromosome studies only, (S) indicates molecular studies only (DNA sequencing) and (B) indicates both cytogenetic and molecular studies. Where AS follows a voucher number, the cytogenetic results have been published already in Strydom and Spies (1994b) but additional molecular work was done on these specimens during this study. Grid references are presented using the degree reference system (Edwards and Leistner 1971). Spies refers to material collected by J.J. Spies, Davidse to G. Davidse, HJTV to H.J.T. Venter, Swart to E. Swart and HdP to H. du Plessis.

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### ***Bewsia* Gooss.**

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#### ***B. biflora* (Hack.) Gooss.**

MPUMALANGA.—2530 (Lydenburg): Steenkamp's Mountains (–AA), Spies 1531 (C).

### ***Catalepis* Stapf & Stent**

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#### ***C. gracilis* Stapf & Stent**

FREE STATE.—2828 (Bethlehem): 10 km from Clarens to Fouriesburg (–CB), Spies 6653 (S).

EASTERN CAPE.—3028 (Matatiele): 26 km from Rhodes to Maclear (–DA), Spies 6962 (S).

### ***Centropodia* Rchb.**

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#### ***C. glauca* (Nees) Cope**

NORTHERN CAPE.—2816 (Oranjemund): 46 km from Bloeddrift to Alexander Bay (–DA), Spies 5706 (C).

### ***Chloris* Sw.**

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#### ***C. virgata* Sw.**

FREE STATE.—2827 (Senekal): 20 km from Senekal to Rosendal (–BD), Spies 6616 (B). 2926 (Bloemfontein): UFS library, Bloemfontein (–AA), Spies 7498 (S).

## ***Cladoraphis* Franch.**

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### ***C. cyperoides* (Thunb.) S.M.Phillips**

NORTHERN CAPE.—2816 (Oranjemund): 46 km from Bloeddrift to Alexander Bay (–CB), *Spies* 5704 (B). 2917 (Springbok): 84 km from Springbok to Kleinsee (–CA), *Spies* 4889 (C); 135 km from Springbok to Kleinsee (–CA), *Spies* 4894 (B).

WESTERN CAPE.—3118 (Vanrhynsdorp): 21 km from Doorn Bay to Donkin Bay (–CD), *Spies* 5356 (C).

### ***C. spinosa* (L.f.) S.M.Phillips**

NORTHERN CAPE.—2917 (Springbok): 82 km from Springbok to Kleinsee (–CA), *Spies* 4885 (B).

WESTERN CAPE.—3118 (Vanrhynsdorp): 7 km from Vanrhynsdorp to Nuwerus (–DA), *Spies* 6335 (S).

## ***Cynodon* Rich.**

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### ***C. dactylon* (L.) Pers.**

SWAZILAND.—2631 (Mbabane): 18 km north east of Mbabane (–BD), *Spies* 2549 (C).

NORTHERN CAPE.—2816 (Oranjemund): Alexander Bay (–DA), *Spies* 2966 (C).

FREE STATE.—2925 (Jagersfontein): 33 km from Bloemfontein to Petrusburg (–AB), *Spies* 7480 (S).

EASTERN CAPE.—3026 (Aliwal North): Aliwal North (–DA), *Spies* 5248 (B) (AS).

## ***Dactyloctenium* Willd.**

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### ***D. aegyptium* (L.) Willd.**

KWAZULU-NATAL.—2832 (Mtubatuba): Richard's Bay (–CC), *HJTV* 9326 (S); 22 km from Cape Vidal to St Lucia (–AD), *Spies* 2403 (C).

## ***Eleusine* Gaertn.**

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### ***E. coracana* (L.) Gaertn. subsp. *africana* (Kenn.-O'Byrne) Hilu & DeWet**

KWAZULU-NATAL.—2832 (Mtubatuba): Cape Vidal (–BA), *Spies* 2365 (C), *Spies* 2366 (C).

WESTERN CAPE.—3118 (Vanrhynsdorp): Koekenaap, irrigation scheme (–CB), *Spies* 2783 (C).

## ***Enneapogon* P.Beauv.**

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### ***E. cenchroides* (Roem. & Schult.) C.E.Hubb**

LIMPOPO.—2428 (Nylstroom): Soutpan at the crater (–CD), *Spies* 3288 (C).

NORTHERN CAPE.—2924 (Hopetown): 31 km from Hopetown to Britstown (–CA), *Spies* 2709 (C).

EASTERN CAPE.—3125 (Steynsburg): Cradock, on rocky outcrop outside town (–DD), *Spies* 7506 (S). 3323 (Willowmore): 110 km from Willowmore to Patensie (–DB), *Spies* 5213 (S).

### ***E. pretoriensis* Stent**

LIMPOPO.—2627 (Potchefstroom): 7 km from Hartbeeshoek turn-off between Muldersdrift and Hekpoort (–BB), *Spies* 3716 (C).

### ***E. scoparius* Stapf**

NORTHERN CAPE.—2922 (Prieska): Nelspoortjie / Vogelstruisbult (–CB), *Swart* 18 (S).

FREE STATE.—2925 (Jagersfontein): 54 km from Petrusburg to Kimberley (–AB), *Spies* 7487 (S)

### ***Enneapogon* sp.**

NORTH-WEST.—2725 (Bloemhof): 31 km from Vryburg to Schweizer-Reineke (–AB), *Spies* 5532 (C).

## ***Enteropogon* Nees**

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### ***E. macrostachyus* (A.Rich.) Benth.**

NAMIBIA.—1913 (Sesfontein): Sesfontein, Kaokoland (–BB), *HJTV* 9339 (B).

## ***Entoplocamia* Stapf**

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### ***E. aristulata* (Hack. & Rendle) Stapf**

NAMIBIA.—2415 (Sossusvlei): Sesriem (–DB), *HJTV* 9353 (S).

## ***Eragrostis* Wolf**

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### ***E. barbinodes* Hack.**

LIMPOPO.—2428 (Nylstroom): Soutpan (–CD), *Spies* 3317 (C).

***E. biflora* Hack. ex Schinz**

FREE STATE.—2926 (Bloemfontein): Bloemfontein (–DA), *Spies* 6705 (S).

***E. capensis* (Thunb.) Trin.**

MPUMALANGA.—2530 (Lydenburg): 41 km from Lydenburg to Roosenekal (–AB), *Spies* 1595 (C). 2729 (Volksrust): Hartbeesfontein, Volksrust (–BB), *Spies* 6532 (S).

FREE STATE.—2729 (Volksrust): 92 km from Harrismith to Normandien Pass (–DC), *Spies* 5069 (C); Giddies Home, Memel (–DC), *Spies* 6551 (S).

KWAZULU-NATAL.—2931 (Stanger): Balito Bay to Umhlali at turnoff before Stanger/Tongaat connection (–AC), *HdP* 110 (C).

EASTERN CAPE.—3028 (Matatiele): 65 km from Rhodes via Naude's Neck (–CC), *Spies* 4696 (C). 3325 (Port Elizabeth): 2 km from Rocklands to Elands River (–CD), *Spies* 3483 (C). 3424 (Humansdorp): 4 km from Humansdorp to Cape St Frances (–BB), *Spies* 3498 (C).

LOCALITY UNKNOWN. *Davidse* 33552 (C).

***E. chloromelas* Steud.**

FREE STATE.—2925 (Jagersfontein): 12 km from Bloemfontein to Petrusburg (–AB), *Spies* 7474 (S).

EASTERN CAPE.—3027 (Lady Grey): 35 km from Aliwal North to Lady Grey (–CA), *Spies* 6947 (B).

***E. ciliaris* (L.) R.Br.**

KWAZULU-NATAL.—2931 (Stanger): On route from Balito Bay to Umhlali at turnoff before Stanger / Tongaat connection (–AC), *HdP* 111 (C).

***E. curvula* (Schrad.) Nees**

MPUMALANGA.—2729 (Volksrust): Hartbeesfontein, Volksrust (–BB), *Spies* 6529 (S).

EASTERN CAPE.—3225 (Somerset East): 35 km from Somerset East to Pearston (–CB), *Spies* 1137 (C). 3324 (Steytlerville): 25 km from Patensie to Willowmore (–DA), *Spies* 5191 (S).

***E. echinochloidea* Stapf**

FREE STATE.—2925 (Jagersfontein): 60 km from Bloemfontein to Petrusburg (–AB), *Spies* 7485 (S).

NORTHERN CAPE.—2917 (Springbok): 36 km from Springbok to Hondeklip Bay (–DB), *Spies* 5933 (S). 3018 (Kamiesberg): 3 km from Bitterfontein to Garies (–DB), *Spies* 2799 (C), *Spies* 2800 (C).

***E. heteromera* Stapf**

SWAZILAND.—2631 (Mbabane): 80 km from Manzini to Lomahasha (–BA), *Spies* 2634 (C).

***E. inamoena* K.Schum.**

KWAZULU-NATAL.—2832 (Mtubatuba): 12 km from Cape Vidal to St Lucia (–AD), *Spies* 2392 (C).

***E. lehmanniana* Nees var. *lehmanniana***

NORTHERN CAPE.—2922 (Prieska): Nelspoortjie / Vogelstruisbult (–CB), *Swart* 20 (S).

EASTERN CAPE.—3125 (Steynsburg): 4 km from turnoff to Graaff-Reinet from Cradock (–DD), *Spies* 7514 (S).

***E. obtusa* Munro ex Ficalho & Hiern.**

FREE STATE.—2827 (Senekal): 20 km from Senekal to Rosendal (–BD), *Spies* 6623 (S). 2924 (Hopetown): 16 km from Griekwastad to Kimberley (–CD), *Spies* 2886 (C).

NORTHERN CAPE.—3123 (Victoria West): 140 km from Beaufort West to Colesberg (–DA), *Spies* 6732 (S).

***E. planiculmis* Nees**

EASTERN CAPE.—3225 (Somerset East): Daggaboer's Neck Pass, Cookhouse to Cradock (–DB), *Spies* 1116 (C).

***E. pseudobtusa* DeWinter**

EASTERN CAPE.—3125 (Steynsburg): Cradock, on rocky outcrop outside town (–DD), *Spies* 7508 (S).

***E. racemosa* (Thunb.) Steud.**

MPUMALANGA.—2729 (Volksrust): Hartbeesfontein, Volksrust (–BB), *Spies* 6533 (S).

FREE STATE.—2729 (Volksrust): 92 km from Harrismith to Normandien Pass (–DC), *Spies* 5066 (C).

KWAZULU-NATAL.—2829 (Harrismith): Windy Corner, Van Reenen (–AD), *Spies* 3279 (C).

EASTERN CAPE.—3027 (Lady Grey): 15 km from Barkly East to Lady Grey (–CD), *Spies* 4743 (C).

***E. sclerantha* Nees subsp. *sclerantha***

FREE STATE.—2828 (Bethlehem): 19 km from Fouriesburg to Clarens (–CA), *Spies* 4844 (C).

***E. superba* Peyr.**

LIMPOPO.—2428 (Nylstroom): Soutpan (–CD), *Spies* 3326 (C).

KWAZULU-NATAL.—2830 (Dundee): 5 km from Muden to Greytown (–DC), *HdP* 136 (C).

FREE STATE.—2925 (Jagersfontein): 12 km from Bloemfontein to Petrusburg (–AB), *Spies* 7470 (S).

***E. tef* (Zucc.) Trotter**

KWAZULU-NATAL.—2832 (Mtubatuba): 22 km from Cape Vidal to St Lucia (–AD), *Spies* 2405 (C).

***E. tenuifolia* (A.Rich.) Steud.**

SWAZILAND.—2631 (Mbabane): Siteki (–BD), *Spies* 2595 (C).

***E. trichophora* Coss. & Dur.**

WESTERN CAPE.—3118 (Vanrhynsdorp): 3 km from Lutzville to Koekenaap (–CB), *Spies* 2774 (C).

***Eustachys* Desv.**

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***E. paspaloides* Lanza & Mattei**

EASTERN CAPE.—3228 (Somerset East): Cookhouse (–DB), *Spies* 5597 (S).

***Fingerhuthia* Nees**

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***F. africana* Lehm.**

NAMIBIA.—2618 (Keetmanshoop): Remshoogte, Annisfontein (–BD), *Spies* 2947 (C).

NORTHERN CAPE.—2917 (Springbok): 21 km from Springbok to Kleinsee (–DB), *Spies* 4879 (S).

WESTERN CAPE.—3118 (Vanrhynsdorp): 35 km from Vanrhynsdorp to Nieuwoudtville (–DA), *Spies* 4349 (B).

EASTERN CAPE.—3125 (Steynsburg): Cradock, on rocky outcrop outside town (–DD), *Spies* 7507 (S).

## ***Harpochoa* Kunth**

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### ***H. falx* (L.f.) Kuntze**

MPUMALANGA.—2530 (Lydenburg): Nederhorst turnoff on Lydenburg-Roosenekal road (–AA), *Spies* 5128 (C); 11 km from Dullstroom to Lydenburg via Frischgewaagd (–AC), *Spies* 5118 (S); 16 km from Dullstroom to Lydenburg via Frischgewaagd (–AC), *Spies* 5125 (C); 5 km from Belfast to Dullstroom (–CA), *Spies* 5113 (C). 2729 (Volksrust): Hartbeesfontein, Volksrust (–BB), *Spies* 6530 (S).

FREE STATE.—2729 (Volksrust): 92 km from Harrismith to Normandien Pass (–DC), *Spies* 5065 (C); 97 km from Harrismith to Normandien Pass (–DC), *Spies* 5078 (C). 2827 (Senekal): 33 km from Senekal to Rosendal (–BD), *Spies* 6629 (C). 2828 (Bethlehem): 38 km from Clarens to Phudatitjhaba (–DA), *Spies* 6670 (S).

EASTERN CAPE.—3027 (Lady Grey): 39 km from Barkly East to Rhodes (–DD), *Spies* 6955 (C); 45 km from Barkly East to Rhodes (–DD), *Spies* 3986 (C); 52 km from Rhodes via Lundean's Neck (–DD), *Spies* 4729 (C). 3028 (Matatiele): 47 km from Rhodes via Naude's Neck (–CC), *Spies* 4691 (C); 65 km from Rhodes to Maclear via Naude's Neck (–CC), *Spies* 4695 (C). 3128 (Umtata): 38 km from Maclear to Elliott (–AC), *Spies* 4712 (C).

## ***Leptochloa* P.Beauv.**

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### ***L. fusca* (L.) Kunth**

NORTHERN CAPE.—2917 (Springbok): 7 km from Springbok to Kleinsee (–DB), *Spies* 4875 (C). 3017 (Hondeklip Bay): 12 km east of Hondeklip Bay (–AD), *Spies* 3037 (C); 5 km east of Kamieskroon (–BB), *Spies* 2991A (C). 3018 (Kamiesberg): 35 km from Leliefontein to Garies (–AB), *Spies* 4316 (C).

FREE STATE.—2826 (Brandfort): Soetdoring nature reserve (–CC), *Spies* 7622 (S).

WESTERN CAPE.—3118 (Vanrhynsdorp): Gifberg, FM tower gate (–CB), *Spies* 3794 (C). 3419 (Caledon): McGregor FM tower, Riviersonderend Mountains (–BA), *Spies* 3932 (C).

EASTERN CAPE.—3324 (Steytlerville): 34 km from Patensie to Willowmore (–DD), *Spies* 5200 (C).

LOCALITY UNKNOWN. *Davidse* 33407 (C).

## ***Microchloa* R.Br.**

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### ***M. caffra* Nees**

MPUMALANGA.—2530 (Lydenburg): 49 km from Lydenburg to Machadadorp (–CB), *Spies* 5146 (B) (AS).

## ***Odyssea* Stapf**

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### ***O. paucinervis* (Nees) Stapf**

NORTHERN CAPE.—3017 (Hondeklip Bay): Groen River mouth, on dunes and rocks (–DC), *Spies* 3384 (C).

## ***Perotis* Aiton**

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### ***P. patens* Gand.**

KWAZULU-NATAL.—2832 (Mtubatuba): Richard's Bay (–CC), *HJTV* 9323 (S).

## ***Pogonarthria* Stapf**

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### ***P. squarrosa* (Roem. & Schult.) Pilg.**

FREE STATE.—2925 (Jagersfontein): 46 km from Bloemfontein to Petrusburg (–AB), *Spies* 7483 (S). 2926 (Bloemfontein): Bloemfontein (–AA), *Spies* 7504 (S).

## ***Polevansia* DeWinter**

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### ***P. rigida* DeWinter**

EASTERN CAPE.—3027 (Lady Grey): Between Lady Grey and Barkly East (–CD), *Spies* 7621 (S).

## ***Rendlia* Chiov.**

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### ***R. altera* (Rendle) Chiov.**

MPUMALANGA.—2530 (Lydenburg): 4 km from Pilgrim's Rest to Graskop (–BA), *Spies* 5133 (S).

## ***Schmidtia* Steud.**

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### ***S. kalihariensis* Stent**

NORTHERN CAPE.—2824 (Kimberley): 24 km from Kimberley to Barkly West (–DA), *Spies* 7490 (S).

FREE STATE.—2925 (Jagersfontein): 37 km from Petrusburg to Kimberley (–AB), *Spies* 7486 (S).



***S. pappophoroides* Steud.**

LIMPOPO.—2329 (Pietersburg): 66 km from Pietersburg to Louis Trichardt (–BB), *HdP* 186 (C).

WESTERN CAPE.—3118 (Vanrhynsdorp): 7 km from Vanrhynsdorp to Nuwerus (–DA), *Spies* 6334 (S).

***Schmidtia* sp.**

NORTHWEST.—2624 (Vryburg): 36 km from Vryburg to Amalia (–DC), *Spies* 5536 (C).

***Sporobolus* R.Br.**

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***S. africanus* (Poir.) Robyns & Tournay**

KWAZULU-NATAL.—2832 (Mtubatuba): Cape Vidal (–BA), *Spies* 2369 (C).

FREE STATE.—2925 (Jagersfontein): 46 km from Bloemfontein to Petrusburg (–AB), *Spies* 7476 (S).

WESTERN CAPE.—3420 (Bredasdorp): 1.4 km from De Hoop Nature Reserve (–AD), *Spies* 4508 (C).

***S. albicans* Nees**

WESTERN CAPE.—3220 (Sutherland): 15 km from Sutherland to Matjiesfontein, Verlatenkloof (–BC), *Spies* 3141 (C).

***S. consimilis* Fresen.**

NAMIBIA.—2316 (Nauchas): Gaub River bridge, on road between Namib park and Solitaire (–CC), *HJTV* 9354 (S).

***S. fimbriatus* (Trin.) Nees**

KWAZULU-NATAL.—2832 (Mtubatuba): Richard's Bay (–CC), *HJTV* 9328 (S).

EASTERN CAPE.—3125 (Steynsburg): Cradock, next to rocky outcrop outside town (–DD), *Spies* 7509 (S).

***S. ioclados* (Trin.) Nees**

NORTHERN CAPE.—2923 (Douglas): 65 km from Hope Town to Strydenburg (–DC), *Spies* 2717(C).

***S. pyramidalis* P.Beauv.**

EASTERN CAPE.—3028 (Matatiele): 31 km from Rhodes to Maclear in Naude's Neck (–DA), *Spies* 6966 (S).

***S. virginicus* (L.) Kunth**

KWAZULU-NATAL.—2931 (Stanger): Tongaat River, on beach near bridge (–CA),  
HdP 122 (C).

***Stiburus* Stapf**

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***S. alopecuroides* (Hack.) Stapf**

MPUMALANGA.—2530 (Lydenburg): Steenkamp's Mountains, 18 km from  
Dullstroom to Goede Hoop (–AC), Spies 1470 (C).

***S. conrathii* Hack.**

MPUMALANGA.—2530 (Lydenburg): 19 km from Lydenburg to Weltevreden (–AB),  
HdP 19 (C).

***Tragus* Haller**

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***T. berteronianus* Schult.**

FREE STATE.—2827 (Senekal): 20 km from Senekal to Rosendal (–BD), Spies  
6620 (S).

***T. koelerioides* Asch.**

EASTERN CAPE.—3123 (Victoria West): 140 km from Beaufort West to Colesberg  
(–DA), Spies 6729 (S).

***T. racemosus* (L.) All.**

FREE STATE.—2727 (Kroonstad): Kroonstad (–CA), Spies 7108 (S).

***Tragus* sp.**

FREE STATE.—2827 (Senekal): 6 km from Clocolan to Peka bridge (–DC), Spies  
4803 (C).

***Trichoneura* Anderss.**

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***T. grandiglumes* (Nees) Ekman**

FREE STATE.—2827 (Senekal): 6 km from Nebo to Fouriesburg via Generaal's  
Neck (–DB), Spies 4833 (C).

## OUTGROUPS

### ***Aristida* L. (Aristidoideae)**

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#### ***A. adscensionis* L.**

MPUMALANGA.—2729 (Volksrust): Hartbeesfontein, Volksrust (–BB), *Spies* 6540 (S).

### ***Chaetobromus* Nees (Danthonioideae)**

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#### ***Chaetobromus involubrates* (Schrad.) Nees subsp. *dregeanus* (Nees) Verboom**

NORTHERN CAPE.—3119 (Calvinia): 83 km from Clanwilliam to Nieuwoudtville (–AC), *Spies* 6284 (S).

### ***Karoochloa* Conert & Türpe (Danthonioideae)**

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#### ***Karoochloa purpurea* (L.f.) Conert & Türpe**

WESTERN CAPE.—3320 (Montagu): 48 km from Montagu to Touwsrivier (–CD), *Spies* 6241 (S).

### ***Phragmites* Adans. (Arundinoideae)**

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#### ***Phragmites australis* (Cav.) Steud.**

FREE STATE.—2729 (Volksrust): Giddies Home, Memel (–DC), *Spies* 6557 (S).

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**Table 2.2.** List of Genbank sequences used in this study.

Species	(Subfamily, Tribe)	Genbank accession number	Region investigated	Reference
<i>Astreblla lappacea</i>	Chloridoideae, Cynodonteae	AF019838	ITS1-ITS2	Hsiao <i>et al.</i> 1999
<i>Centropodia glauca</i>	Chloridoideae, <i>incertae cedis</i>	AF019861	ITS1-ITS2	Hsiao <i>et al.</i> 1998
<i>Cynodon dactylon</i>	Chloridoideae, Cynodonteae	AJ489601	ITS1-ITS2	Ferris <i>et al.</i> (unpubl.)
<i>Cynodon dactylon</i>	Chloridoideae, Cynodonteae	AJ489466	TrnL intron	Ferris <i>et al.</i> (unpubl.)
<i>Eleusine coracana</i>	Chloridoideae, Eragrostideae	AJ489602	ITS1-ITS2	Ferris <i>et al.</i> (unpubl.)
<i>Eleusine coracana</i>	Chloridoideae, Eragrostideae	AJ489464	TrnL intron	Ferris <i>et al.</i> (unpubl.)
<i>Eleusine indica</i>	Chloridoideae, Eragrostideae	AJ489603	ITS1-ITS2	Ferris <i>et al.</i> (unpubl.)
<i>Eleusine indica</i>	Chloridoideae, Eragrostideae	AJ489465	TrnL intron	Ferris <i>et al.</i> (unpubl.)
<i>Imperata cylindrica</i>	Panicoideae, Andropogoneae	AY116297	ITS1-ITS2	Hodkinson <i>et al.</i> 2002b
<i>Imperata cylindrica</i>	Panicoideae, Andropogoneae	AY116262	TrnL-F spacer	Hodkinson <i>et al.</i> 2002b
<i>Merxmuellera rangei</i>	Chloridoideae, <i>incertae cedis</i>	AF019862	ITS1-ITS2	Hsiao <i>et al.</i> 1998
<i>Muhlenbergia richardsonis</i>	Chloridoideae, Eragrostideae	AF019837	ITS1-ITS2	Hsiao <i>et al.</i> 1999
<i>Panicum lanipes</i>	Panicoideae, Paniceae	AY129710	ITS1-ITS2	Missaoui <i>et al.</i> (unpubl.)
<i>Panicum lanipes</i>	Panicoideae, Paniceae	AY142732	TrnL intron	Missaoui <i>et al.</i> (unpubl.)
<i>Panicum stapfianum</i>	Panicoideae, Paniceae	AY129724	ITS1-ITS2	Missaoui <i>et al.</i> (unpubl.)
<i>Panicum stapfianum</i>	Panicoideae, Paniceae	AY142745	TrnL intron	Missaoui <i>et al.</i> (unpubl.)
<i>Sorghum halepense</i>	Panicoideae, Andropogoneae	AY116293	ITS1-ITS2	Hodkinson <i>et al.</i> 2002b
<i>Sorghum halepense</i>	Panicoideae, Andropogoneae	AY116244	TrnL-F spacer	Hodkinson <i>et al.</i> 2002b
<i>Triodia stenostachya</i>	Chloridoideae, Eragrostideae	AF019836	ITS1-ITS2	Hsiao <i>et al.</i> 1999

## **2.2 Methods**

### **2.2.1 Cytogenetics**

#### **2.2.1.1 Meiotic analysis**

Young inflorescences were fixed in Carnoy's fixative (ethanol: chloroform: acetic acid::6:3:1) (Carnoy 1886). The fixative was replaced by 70% (v/v) ethanol 24-48 hours after fixation. Anthers of the inflorescences were squashed in 2% (m/v) aceto-carmine (Darlington and La Cour 1976) on a microscope slide. Contrast between cytoplasm and chromosomes was enhanced by adding a droplet of 45% (v/v) acetic acid, saturated with iron acetate, to the stain immediately before making the squash (Thomas 1940) and then gently heating the slide over a spirit flame. Squashes were made according to Darlington and La Cour's (1976) method. The slides were made permanent by freezing them with liquid carbon dioxide (Bowen 1956), followed by dehydration in ethanol and mounting in Euparal.

Whenever possible, at least twenty cells each of diakinesis, metaphase I, anaphase I and telophase I were examined in each specimen. The gametic chromosome numbers, the presence of B chromosomes as well as the percentage rod and ring bivalents and multivalents were recorded. In the case of metaphase I, anaphase I and telophase I, the number of chromosomal abnormalities (univalents, chromosome laggards and micronuclei) were recorded as well.

Genome relationships in tetraploid specimens were determined with the model of Kimber and Alonso (1981).

#### **2.2.1.2 Microphotography**

Microphotography was done using a Nikon Microphot-FXA photomicroscope with Pan-F 35 mm (ASA 50) black and white films. The films were developed for twelve minutes in Agfa Rodinol film developer and then rinsed in water for approximately 5 minutes. After fixation in Ilford rapid fixer for 10 minutes, the films were rinsed in running water for 20 minutes. The films were then left to dry overnight.

Ilfospeed developer was used to develop the photographs and development was then stopped in water to which some acetic acid was added. The photographs were fixed with Ilford Hypam fixative where after the photographs were rinsed in water for 5 minutes and left face up, to dry. Ilford Multigrade IV RC DE LUXE paper was used for the photographs.

A digital camera, Nikon Coolpix 990 was used to photograph meiotic stadia during the latter part of the study. Images were directly transferred to the computer.

Microphotographs, depicting meiotic stages, abnormalities or certain behavioral trends during meiosis, were mounted on herbarium sheets and are stored in the Geo Potts Herbarium, Bloemfontein. Selections of these photographs, which depict certain of these phenomena the best, are included in this thesis.

## **2.2.2 Molecular studies**

The leaves of the different specimens were collected in the veld and stored in a saturated sodium chloride and hexadecyl trimethyl ammonium bromide (CTAB) solution (Rogstad 1992).

### **2.2.2.1 DNA extraction**

The CTAB method (Doyle and Doyle 1987) was used to extract DNA from  $\pm$  0.5 g of leaf material. The leaves were rinsed with distilled water and blotted with paper before the extractions were carried out in eppendorf tubes. The material was ground to a fine powder with liquid nitrogen. The ground tissue was then immediately incubated at 65°C for one hour in 600  $\mu$ l CTAB extraction buffer [2% (m/v) CTAB, 100mM Tris-HCl (pH 8.0), 25mM EDTA (pH 8.0), 1.4M NaCl, to which 0.2% (v/v) 2-mercapto-ethanol had been added just before use]. After one hour 600  $\mu$ l chloroform:iso-amylalcohol (24:1) was added, mixed thoroughly and the resultant mixture centrifuged for five minutes at 5000 g. The supernatant was transferred to a clean tube and to this 600  $\mu$ l cold (-20°C) absolute ethanol, containing 3M sodium acetate (25:1) was added to precipitate the DNA. After one hour of incubation at 4°C, the mixture was centrifuged at 7000 g. for eight minutes. The supernatant was discarded and the DNA pellet washed twice with 70% (v/v) ethanol containing 10mM ammonium acetate. After decanting the ethanol and evaporating any excess

ethanol left, the DNA was dissolved in sterilized, distilled water (20-50  $\mu$ l, depending on the size of the pellet).

A modification of the CTAB method, adapted for extraction from herbarium specimens (Loockerman and Jansen 1996), was also used. This method differs in the following aspects from the above mentioned:

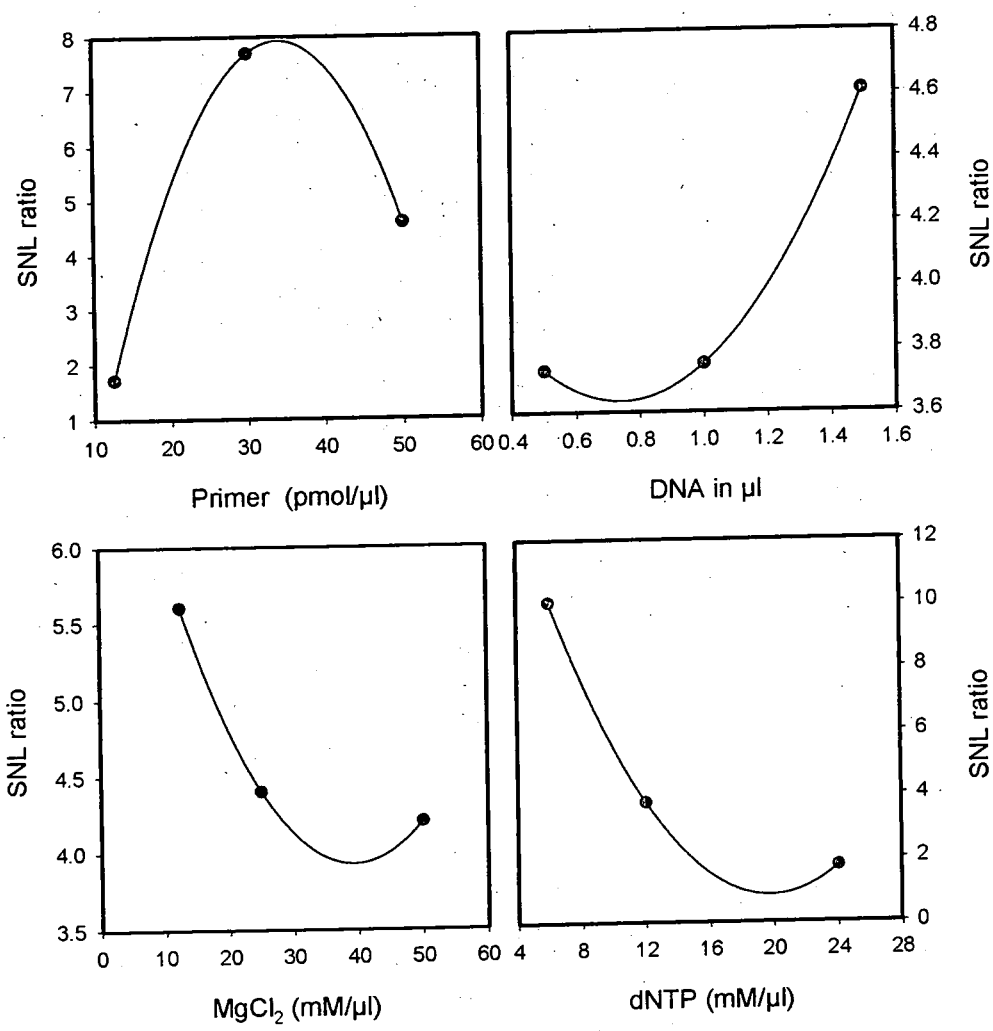
- The ground tissue was incubated at 60°C for only half an hour.
- The CTAB extraction buffer contains 4% (m/v) CTAB and an additional 1% (m/v) PVP.
- The mixture was centrifuged at 7000 g. for two minutes after the chloroform:iso-amylalcohol (24:1) was added.
- The mixture was incubated overnight at 4°C to precipitate the DNA.
- After the mixture was incubated overnight, it was centrifuged at 7000 g. for five minutes.

### 2.2.2.2 Taguchi optimization

The sequencing reactions were optimized according to a modified Taguchi method (Cobb and Clarkson 1994). With this optimization, the optimal conditions for four reaction variables can be achieved, by using only nine reactions. The number of experimental reactions (E) required (nine), can be calculated by using the equation  $E = 2k + 1$ , where k is the number of factors to be tested (four) (Cobb and Clarkson 1994). These variables are the primer, magnesium chloride, DNA and dNTP concentrations. In this optimization, three concentrations of each reaction component are varied in an orthogonal array. With this Taguchi method, the product yield for each reaction is used to estimate the effects that the individual components have on the amplification products (Cobb and Clarkson 1994). This yield can be calculated by using quadratic loss functions, which are referred to as signal to noise (SNL) ratios by Taguchi (Taguchi and Wu 1980; Taguchi 1986):

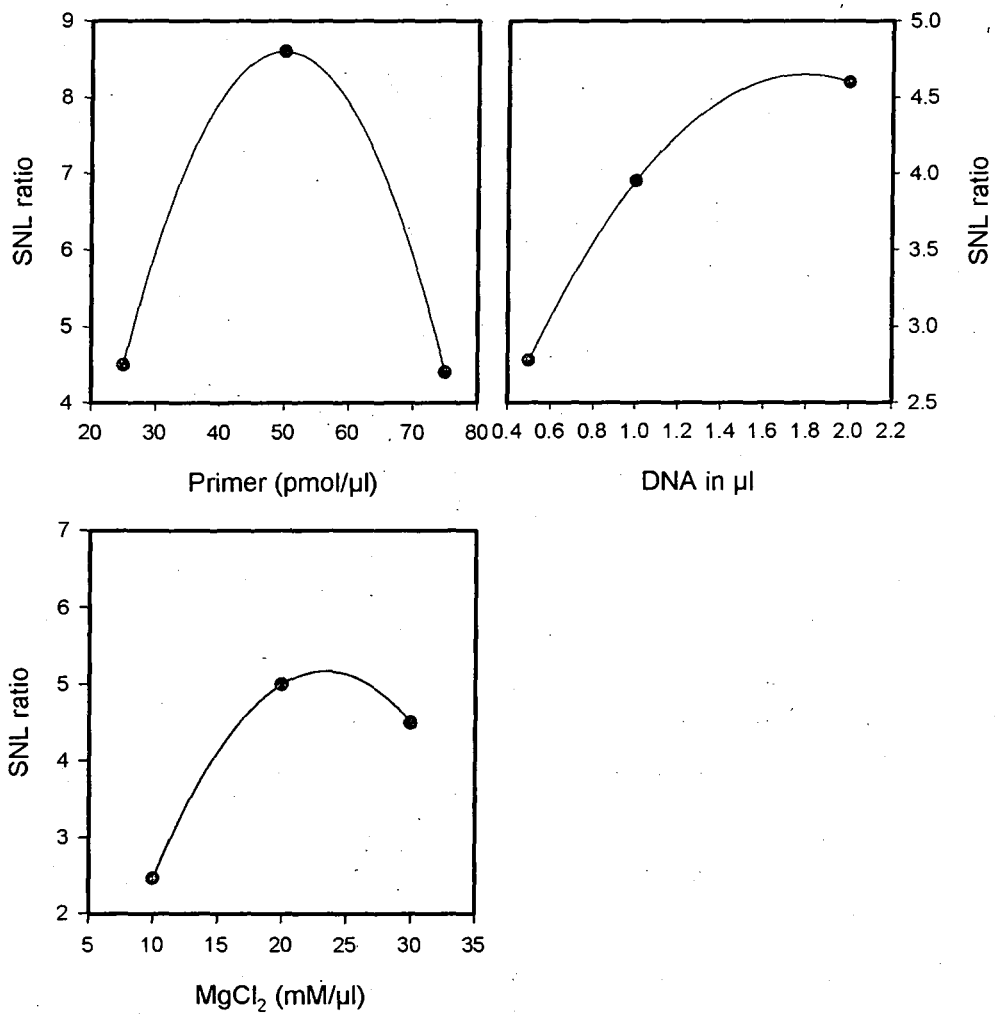
$$SNL = -10 \log [1/n \sum 1/y^2],$$

where SNL is the signal to noise ratio, y is the yield for each amplification reaction and n is the number of levels. For each of the reaction components the largest SNL value represents the optimal condition.



**Figure 2.1.** Taguchi optimization values for reaction conditions for *ITS* fragment amplification. Largest SNL values were taken as reaction optima.





**Figure 2.2.** Taguchi optimization values for reaction conditions for *trnL-F* fragment amplification. Largest SNL values were taken as reaction optima. dNTP optima were not determined as this was included in the 5X buffer.

## 2.2.2.3 Sequencing

### 2.2.2.3.1 *ITS* fragment amplification

Genomic DNA was used to amplify the DNA region between the 18S and 5.8S nrDNA genes (the *ITS1* region), as well as between the 5.8S and 26S nrDNA genes (the *ITS2* region). In both cases a part of the 5.8S gene was also amplified as a result of the annealing sites of the primers. The primers used for PCR amplification of the regions were nNc18S10 (Wen and Zimmer 1996) and *ITS*<sub>2</sub> (White *et al.* 1990) [for *ITS1*] and 5.8<sub>s</sub> (Sun *et al.* 1994) and *ITS*<sub>4</sub> (White *et al.* 1990) [for *ITS2*]. The regions were also sequenced using these primers.

<b>nNc18S10</b>	5' – AGGAGAAGTCGTAACAAG – 3'
<b><i>ITS</i><sub>2</sub></b>	5' – GCTGCGTTCTTCATCGATCG – 3'
<b>5.8<sub>s</sub></b>	5' – ACGACTCTCGGCAAC – 3'
<b><i>ITS</i><sub>4</sub></b>	5' – TCCTCCGCTTATTGATATGC – 3'

The PCR reactions were performed in a total volume of 25 µl. The reactions were optimized according to Taguchi (2.2.2.2) (Figure 2.1) and each reaction consisted of 1.5 µl DNA, 0.6 µl dNTP (10mM), 0.5 µl MgCl<sub>2</sub> (20mM), 2.8 µl of each primer (12.5pmol), 2.5 µl 10X Buffer, 2 µl DMSO, and 0.16 µl *Taq* polymerase (5u).

The reactions were briefly centrifuged and placed in the Perkin Elmer GeneAmp PCR system 9600. An initial denaturation step at 94°C for 3 minutes was followed by 40 amplification cycles, each consisting of 30 sec. at 94°C, 30 sec. at 50°C and 90 sec. at 72°C. Amplification was completed by a final step at 72°C for 7 minutes (modification of Baldwin 1992). Strong denaturing reaction conditions were needed because of the high GC content of the *ITS* region and the presence of secondary structure (Buckler and Holtsford 1996a). This was achieved by the addition of DMSO, which reduces strand reannealing (Winship 1989; Vadaraj and Skinner 1994). Buckler *et al.* (1997) also suggested that the addition of DMSO would prevent the preferential amplification of pseudogenes. These might be preferentially amplified due to low secondary structure stability.

The amplification products were run on 1% (m/v) agarose gels with 1X TAE running buffer (40mM Tris-acetate, 1mM EDTA, pH 8.0) at 60V, intercalated with ethidium bromide and visualized by illumination with ultraviolet (UV) light.

### 2.2.2.3.2 *TrnL-F* fragment amplification

The regions amplified in the chloroplast genome were the intron in the *trnL* gene (between the 5' and 3' exons of the *trnL* gene), the 3' exon of the *trnL* intron, as well as the intergenic spacer between the 3' exon of the *trnL* gene and the 5' exon of the *trnF* gene. The primers used for the PCR were c and f for amplification (Taberlet *et al.* 1991) and primers PS1, PS2, PS3 and PS4 were used for sequencing (Pfosser and Speta 1999).

<b>c</b>	5' – TCGTAACAAGGTTTCCGTAGGTG – 3'
<b>f</b>	5' – TCCTCCGCTTATTGATATGC – 3'
<b>PS1</b>	5' – CTACGGACTTAATTGGATTGAGC – 3'
<b>PS2</b>	5' – GGGGATAGAGGGACTTGAAC – 3'
<b>PS3</b>	5' – GGTTC AAGTCCCTCTATCCC – 3'
<b>PS4</b>	5' – AGGATTTTCAGTCCTCTGCTC – 3'

The PCR reactions were performed in a total volume of 10  $\mu$ l. The reactions were optimized according to Taguchi (2.2.2.2) (Figure 2.2) and consisted of 2  $\mu$ l DNA, 1  $\mu$ l MgCl<sub>2</sub> (20mM), 1  $\mu$ l primer combination (50 pmol), 2  $\mu$ l 5X Buffer (containing 10mM dNTP's) and 0.16  $\mu$ l *Taq* polymerase (5u).

The reactions were briefly centrifuged and placed in the Perkin Elmer GeneAmp PCR system 9600. A touchdown PCR procedure was followed: An initial denaturation at step 94°C for 3 minutes was followed by 2 cycles each of 40 sec. at 94°C, 40 sec. at 58°C and 90 sec. at 72°C. The annealing temperature was decreased with 2°C in every two consecutive cycles until an annealing temperature of 49°C was reached and maintained for 25 cycles. This was followed by a final extension step at 72°C for 7 minutes (modification of Don *et al.* 1991).

The amplification products were visualized as described in 2.2.2.3.1

### 2.2.2.3.3 Sequencing

Sequencing reactions were carried out by using the system based on Sanger *et al.*'s dideoxynucleotide method (1977).

For each template to be sequenced the following were combined:

Sequence reagent pre-mix	1 $\mu$ l
Sequencing buffer	1.5 $\mu$ l
Primer (1 pmol)	1.6 $\mu$ l
DNA template (30ng)	3 $\mu$ l
distilled water	2.9 $\mu$ l
Total volume	10 $\mu$ l

These reactions were placed in the Perkin Elmer thermal cycler with 25 amplification cycles, each consisting of 94°C for 10 sec., 50°C for 5 sec. and 60°C for four minutes, according to the manufacturer's protocol.

After amplification 16  $\mu$ l deionised water was added to each reaction, as well as 64  $\mu$ l 95% (v/v) ethanol. These reactions were vortexed briefly and left for a period of 20 minutes. After 20 minutes the reactions were centrifuged for 20 minutes at 10 000 g., where after the supernatant was aspirated and 250  $\mu$ l 70% (v/v) ethanol was added to wash the pellet. The mixtures were centrifuged at 10 000 g. for 10 minutes and after the supernatant was drawn off the pellets were left to air dry. The pellets were stored in this dry state at 4°C until they were loaded on the gel. Prior to gel loading each pellet was resuspended in 4  $\mu$ l formamide loading buffer and vortexed vigorously for 10-20 sec. to ensure complete resuspension. The samples were briefly centrifuged to collect each sample at the bottom of the tube. Approximately 1.5-2  $\mu$ l of the samples was loaded on a 6% polyacrylamide gel run for 4-6 hours on an ABI Prism<sup>TM</sup> 377 fluorescent sequencing system.

### 2.2.2.3.4 Sequence alignment

The boundaries of the *ITS* and *trnL-F* regions were identified by comparison to known sequences. For sequence editing and assembly of complimentary strands, AutoAssembler version 1.4.0. (Applied Biosystems Inc.) was used. Sequences were aligned in ClustalG version 1.0 (Wilson *et al.* 1999). ClustalG default parameters were used and include a gap opening cost of 1 and gap extension cost of 0.1 with

the slow-accurate pairwise alignment parameter and a gap opening cost of 1 and gap extension penalty of 0.05 for the multiple alignment parameters. Final alignment was visually inspected and manually optimized for phylogenetic analysis. Nucleotide ambiguities were coded as N (uncertain) and when two peaks were definitely present in two strands with the same resolution these were coded as polymorphic according to the IUPAC system.

### 2.2.3 Phylogenetic Analyses

Data were analyzed with the computer program PAUP\* (version 4.0b10) (Swofford 2003). All characters were weighted equally and unordered (Fitch parsimony, Fitch 1971). Uninformative (autapomorphic and constant) characters were removed from all analyses. The heuristic search comprised of an initial search of 10 000 random additions, nearest neighbor interchange (NNI) branch swapping and mulpars not in effect, to search for islands of most parsimonious trees (Maddison 1991). The trees resulting from this search were then subjected to more thorough tree bisection and reconnection (TBR) branch swapping with mulpars on in the absence of a maxtrees limit (with *trnL-F* a maxtree limit of 20 000 was set). The amount of phylogenetic information in the parsimony analysis was estimated using the consistency index (Kluge and Farris 1969) and retention index (Farris 1989) as calculated in PAUP\*. Gaps were treated as missing data, but their presence/absence was coded as a separate binary matrix appended to the initial data matrix (Wojciechowski *et al.* 1993; Oxelman and Liden 1995; Simmons and Ochoterena 2000). Gaps were coded by the program GapCoder (Young and Healy 2003), but where nucleotides were missing due to sequencing failure (N), these were coded as uncertain (?) in the gap matrix. For each heuristic analysis a strict consensus tree (Sokal and Rohlf 1981) was generated. Branch support was determined by means of bootstrap (Felsenstein 1985), jackknife (Lanyon 1985) and decay values (Donoghue *et al.* 1992; Bremer 1994).

Bootstrap searches were performed with 200 bootstrap replicates using simple taxon addition, TBR branch swapping, mulpars selected and saving no more than 500 trees per replicate. The same settings were selected for jackknife searches, but emulate Jac was additionally selected. With emulate Jac the

conditions of the program Parsimony Jackknifer (Farris 1995) is emulated and user specified weighting schemes are not allowed. Therefore the deletion percentage was 36.79%. The following categories were used for bootstrap and jackknife support: weak, 50-74%; moderate, 75-84% and strong support 85-100% as proposed by Crespo *et al.* (2000). Bremer support was determined by 10 random addition sequences per converse constraint, TBR branch swapping, mulpars selected and maxtrees set at 500. Decay of clades was assessed using the reverse constraint approach implemented in Autodecay 4.02 (Eriksson 1998).

Congruence between the individual data sets was assessed by visual inspection of the Strict consensus trees in a conditional combination approach (Huelsenbeck *et al.* 1996; Kellogg *et al.* 1996; Mason-Gamer and Kellogg 1996; Seelanan *et al.* 1997; Johnson and Soltis 1998; Wiens 1998; Whitten *et al.* 2000; Mort *et al.* 2002). Separate analyses were conducted and the results compared for hard incongruence (well supported alternative placement of taxa). The 70% cutoff percentage proposed by Mason-Gamer and Kellogg (1996) was used. For the combined analysis all taxa with either *trnL-F* or *ITS* data missing were removed.

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## CHAPTER 3

# Chromosome numbers

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**PUBLISHED AS:**

R. ROODT AND J.J. SPIES. 2002. POACEAE: CHROMOSOME STUDIES ON AFRICAN PLANTS. 18. THE SUBFAMILY CHLORIDOIDEAE. *BOTHALIA* 32 (2): 240–249.



Chromosome numbers are reported for 79 specimens representing 42 species in 19 genera. Basic chromosome numbers of 6, 9 and 10 were observed. Meiosis was normal in most instances. B chromosomes, which have a high incidence in the grass family, were only observed in 2 specimens. Polyploidy is reported for 52 specimens and 37 new or additional counts are reported.

## 3.1 Introduction

The subfamily Chloridoideae comprises  $\pm$  150 genera and 1 360 species and occurs mainly in arid regions (Hilu and Alice 2001). The plants probably originated in Africa (Hartley 1964), hence the great representation of the subfamily in Africa and especially southern Africa, with  $\pm$  51 genera and 235 species (Gibbs Russell *et al.* 1990). It is currently divided into five tribes by the Grass Phylogeny Working Group (2001): Cynodonteae Dumort., Eragrostideae Stapf, Leptureae Dumort., Orcuttieae Reeder and Pappophoreae Kunth. The genus *Centropodia* Rchb. and the species *Merxmullera rangei* (Pilg.) Conert, previously included in the Arundinoideae, are now included in the Chloridoideae. They have not previously been included in any of the recognized tribes (GPWG 2001).

The aim of this study is to investigate chromosome numbers, meiotic chromosome behavior and polyploid levels of some southern African representatives of this subfamily.

## 3.2 Results and Discussion

Seventy-nine plants, representing 42 species and 19 genera, were studied (Appendix A). They represent three of the recognized tribes, namely Cynodonteae, Eragrostideae and Pappophoreae, as well as the unplaced genus *Centropodia*.

### Tribe Cynodonteae

In the genus *Chloris* Sw., a single *C. virgata* Sw. specimen was investigated and found to be diploid ( $n = x = 10$ ) (Figure 3.1A). This confirms the basic chromosome number of 10 for this genus (Darlington and Wylie 1955; Pienaar 1955; Ornduff 1967, 1968, 1969; Federov 1969; Moore 1970, 1971, 1972, 1973, 1974, 1977; Goldblatt 1981, 1983, 1985, 1988; Goldblatt and Johnson 1990, 1991, 1998,

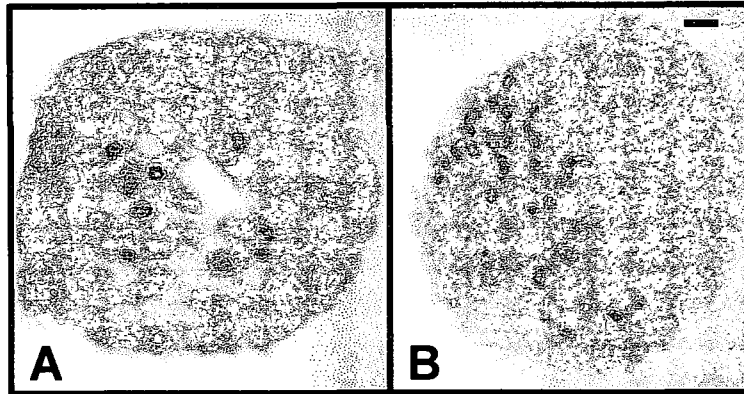


2000). Polyploid levels do occur in the genus and triploid and tetraploid numbers have been observed in South African specimens by previous authors (Moffet and Hurcombe 1949; De Wet 1954a; Spies and Du Plessis 1987). The genus is known to contain many aneuploid deviations from the basic chromosome number of ten (Fish 2000), but none have been observed in South African specimens before (Hunter 1934; Moffet and Hurcombe 1949; De Wet 1954a; Spies and Du Plessis 1987; Spies and Jonker 1987; Strydom and Spies 1994b).

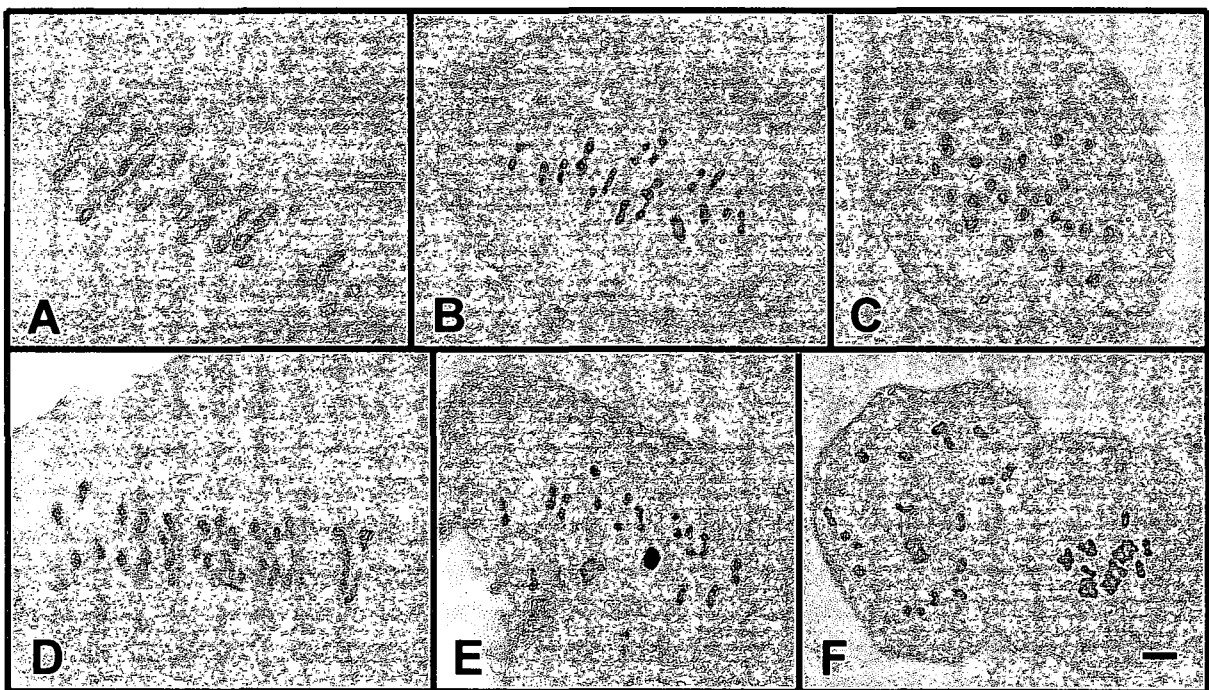
*Cynodon dactylon* (L.) Pers is an introduced species in tropical and warm temperate areas throughout the world, but is indigenous to southern Africa (Gibbs Russell and Spies 1988). Two specimens were investigated, both with  $2n = 4x = 36$  (Figure 3.1B). Tetraploidy is by far the most numerous polyploid level present in the genus (Darlington and Wylie 1955; Pienaar 1955; Ornduff 1968, 1969; Federov 1969; Moore 1970, 1972, 1973, 1974, 1977; Goldblatt 1981, 1983, 1985; Goldblatt and Johnson 1990, 1994, 1996, 1998). De Silva and Snaydon (1995) concluded that the differences in the polyploid level in *C. dactylon* could be related both to different climatic regions and different ecological habitats. The populations growing in arid, dry and intermediate regions were tetraploid and those from wetter regions consisted entirely of diploid plants. De Silva and Snaydon (1995) also related these polyploid levels to soil acidity and alkalinity (tetraploid = pH > 6.5; diploid = pH < 5.0). These findings might explain the predominance of tetraploidy in South Africa with its dryer, more arid regions. A basic chromosome number of  $x = 9$  is confirmed for this species and genus, although 10 has also been reported in a few instances (Ornduff 1968, 1969; Moore 1970, 1972, 1974, 1977; Goldblatt 1981, 1983, 1985; Goldblatt and Johnson 1990, 1994, 1996, 1998).

Chromosome numbers for the genus *Enteropogon* Nees have only been reported once and a basic chromosome number of 10 published for the genus (Darlington and Wylie 1955). One *E. macrostachyus* (A.Rich.) Benth. specimen investigated in this study was found to be tetraploid ( $n = 2x = 20$ ). This is the first report on a South African specimen in the genus, the previous one being from India.

Twelve specimens representing the species *Harpochloa falx* (L.f.) Kuntze were studied. Nine of these were tetraploid and three were hexaploid. All specimens had



**Figure 3.1.** Photomicrographs of meiotic chromosomes in the genera *Chloris* and *Cynodon*. **A.** *Chloris virgata*, Spies 6616,  $2n = 2x = 20$ , diakinesis with  $20_{II}$ . **B.** *Cynodon dactylon*, Spies 2549,  $2n = 4x = 36$ , metaphase I. Scale bar: 5  $\mu$ m.



**Figure 3.2.** Photomicrographs of meiotic chromosomes in the genus *Harpochloa*. **A–F.** *H. falx*. **A.** Spies 3986,  $2n = 4x = 36$ , metaphase I. **B, C.** Spies 4695,  $2n = 4x = 36$ , metaphase I (**B**) and desynapsis of bivalents during metaphase I (**C**). **D.** Spies 4729,  $2n = 4x = 36$ , metaphase I. **E.** Spies 5113,  $2n = 4x = 36$ , metaphase I. **F.** Spies 6955,  $2n = 4x = 36$ , diakinesis with  $18_{II}$ . Scale bar: C, E = 8  $\mu$ m; A, B, D, F = 10  $\mu$ m.

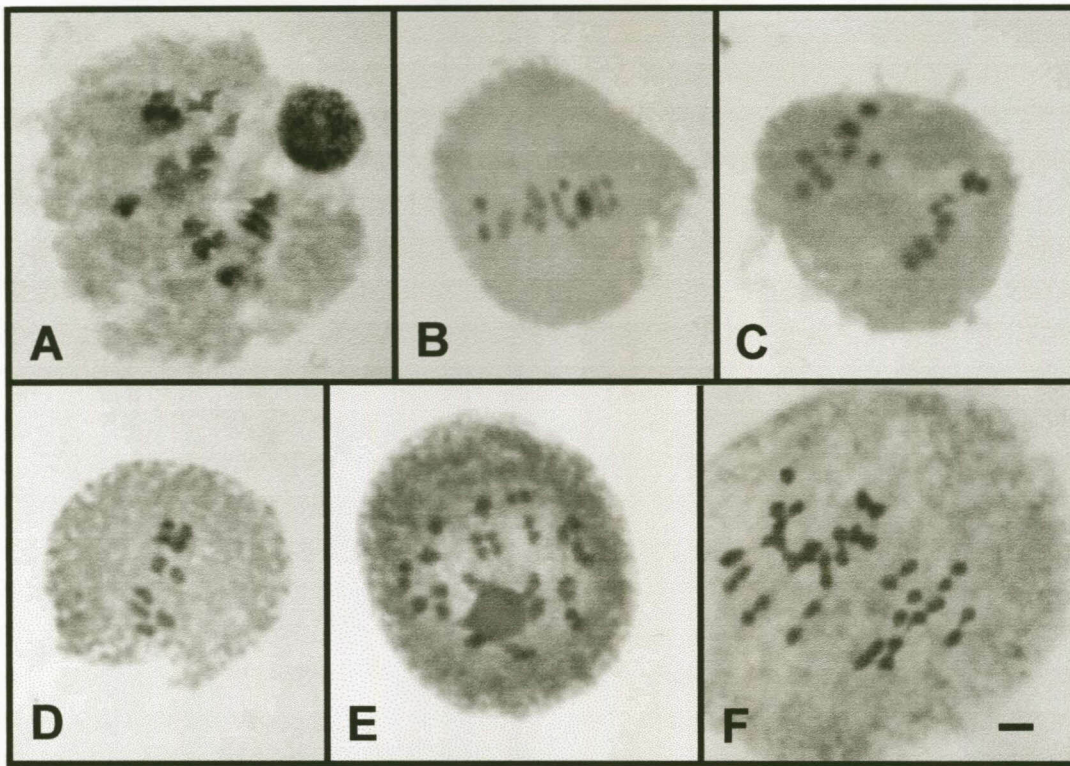
multiples of nine ( $n = 2x = 18$ ;  $n = 3x = 27$ ) (Figure 3.2A–F). This represents a new basic chromosome number for this genus and could imply that *Harpochloa* Kunth also has two basic chromosome numbers (De Wet 1958; Spies and Du Plessis 1986a; Spies *et al.* 1991; Strydom and Spies 1994b). Furthermore, no known reports of any diploid specimens in the genus exist, which could indicate the existence of an older polyploid complex.

The genus *Tragus* Haller is widespread throughout the tropics, but mainly in Africa. It is especially common in disturbed areas (Clayton and Renvoize 1986; Fish 2000). One *Tragus* specimen was investigated. It was diploid with  $n = x = 10$ . Only diploid, as in this study, or tetraploid chromosome numbers are known for this genus, based on a basic chromosome number of 10 (Darlington and Wylie 1955; Ornduff 1967, 1968, 1969; Federov 1969; Moore 1970, 1972, 1973, 1974, 1977; Goldblatt 1981, 1983, 1988; Goldblatt and Johnson 1990, 1994, 1998).

### Tribe Eragrostideae

The genus *Bewsia* Gooss. is monotypic and a single *Bewsia biflora* (Hack.) Gooss. specimen was investigated. This specimen was tetraploid ( $n = 2x = 20$ ) (Figure 3.3A), which confirms a basic chromosome number of ten, based on previous reports by De Wet and Anderson (1956) of  $2n = 3x = 30$  and Davidse *et al.* (1986) of  $2n = 45$  from Zimbabwe. Davidse *et al.* (1986) reported on very irregular meiosis in the particular specimen. Results presented in this study are the third known report for this genus.

*Cladoraphis* Franch. comprises two species *C. cyperoides* (Thunb.) S.M. Phillips and *C. spinosa* (L.f.) S.M. Phillips. De Winter (1955) included this genus in *Eragrostis* Wolf, but later authors (Phillips 1982; Clayton and Renvoize 1986; Gibbs Russell *et al.* 1990; Watson and Dallwitz 1992 onwards) retained its separate generic status. It has a very specific habitat and occurs in sandy desert (*C. spinosa*) and coastal dunes (*C. cyperoides*) (Clayton and Renvoize 1986), mainly in the western regions of Namibia and Northern and Western Cape (Fish 2000). Five specimens were investigated representing both species. All four *C. cyperoides* specimens were diploid, with *C. spinosa* being tetraploid ( $n = 2x = 20$ ) (Figure 3.3B–E). This is, to the best of our knowledge, the first reports for chromosome numbers in this genus.



**Figure 3.3.** Photomicrographs of meiotic chromosomes in the genera *Bewsia*, *Cladoraphis* and *Dactyloctenium*. **A.** *Bewsia biflora*, Spies 1531,  $2n = 4x = 40$ , diakinesis with  $20_{II}$ . Three chromosomes are not in focus on this photograph. **B,** **C.** *Cladoraphis cyperoides*, Spies 4894,  $2n = 2x = 20$ , metaphase I (**B**) and anaphase I, 10-10 segregation (**C**). **D.** *C. cyperoides*, Spies 5356,  $2n = 2x = 20$ , diakinesis. **E.** *C. spinosa*, Spies 4885,  $2n = 4x = 40$ , diakinesis with  $20_{II}$ . **F.** *Dactyloctenium aegyptium*, Spies 2403,  $2n = 4x = 40$ , metaphase I. Scale bar: 5  $\mu\text{m}$ .

One specimen of the widespread tropical weed (Gould and Soderstrom 1974) *Dactyloctenium aegyptium* (L.) Willd. was investigated. It was found to be tetraploid ( $n = 2x = 20$ ) (Figure 3.3F). Three basic chromosome numbers are recorded for this genus,  $x = 9, 10, 12$  [Darlington and Wylie 1955 ( $x = 10, 12$ ); Pienaar 1955 ( $x = 9, 12$ ); Ornduff 1967 ( $x = 10$ ), 1968 ( $x = 12$ ); Moore 1971 ( $x = 9$ ), 1972 ( $x = 9$ ), 1977 ( $x = 12$ ); Goldblatt 1981 ( $x = 9, 10, 12$ ), 1983 ( $x = 9, 10, 12$ ), 1985 ( $x = 12$ ), 1988 ( $x = 9, 10$ ); Goldblatt and Johnson 1990 ( $x = 9, 10, 12$ ), 1991 ( $x = 10$ ), 1994 ( $x = 12$ ), 1998



( $x = 10, 12$ )]. This is one of the genera in the Chloridoideae (as is *Sporobolus*), with the most variation in basic chromosome number.

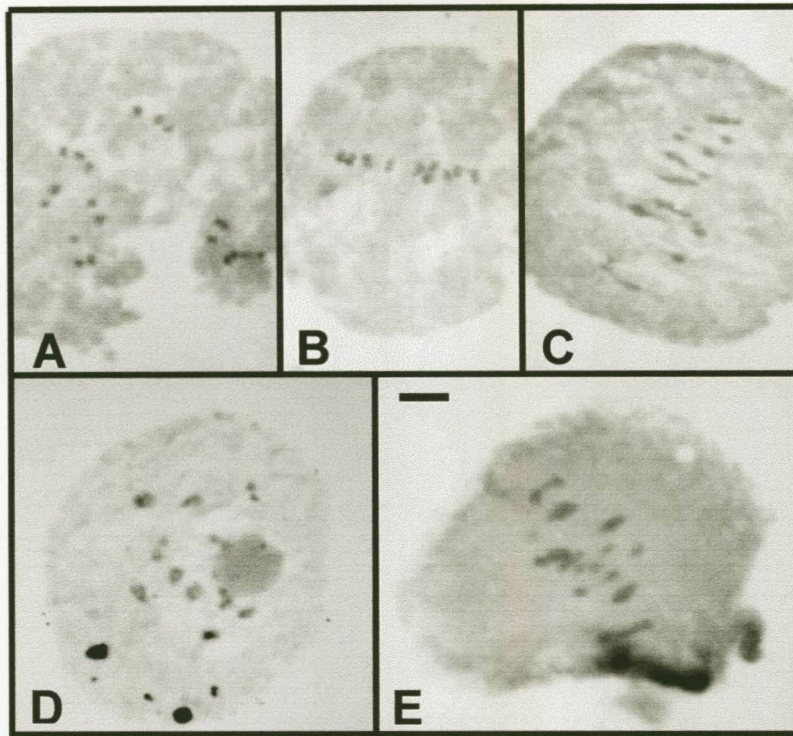
The generic status of *Diplachne* P.Beauv. has long been in doubt (McVaugh 1983; Peterson *et al.* 1997), with some authors preferring to unite this genus with the closely related genus *Leptochloa* P.Beauv. (McNeill 1979; Phillips 1982). These two genera have traditionally been kept distinct by Old World taxonomists where these genera are quite distinct, whereas the position of the genera from the Americas are very confused with intergrading taking place (Phillips 1982). For this study, the accepted name *Leptochloa* will be used.

Eight *Leptochloa fusca* (L.) Kunth specimens were investigated and all were diploid ( $n = x = 10$ ) (Figure 3.4A–E), which confirms the basic chromosome number of ten for this genus (Darlington and Wylie 1955; Ornduff 1968; Federov 1969; Moore 1977; Goldblatt and Johnson 1990, 1991, 1994, 1998). Previous studies have mostly reported tetraploids and this is the first study with such a large number of diploids. The specimens investigated in this study were largely from Northern and Western Cape, and due to the widespread distribution of this species, the total variation present might not be represented in this study.

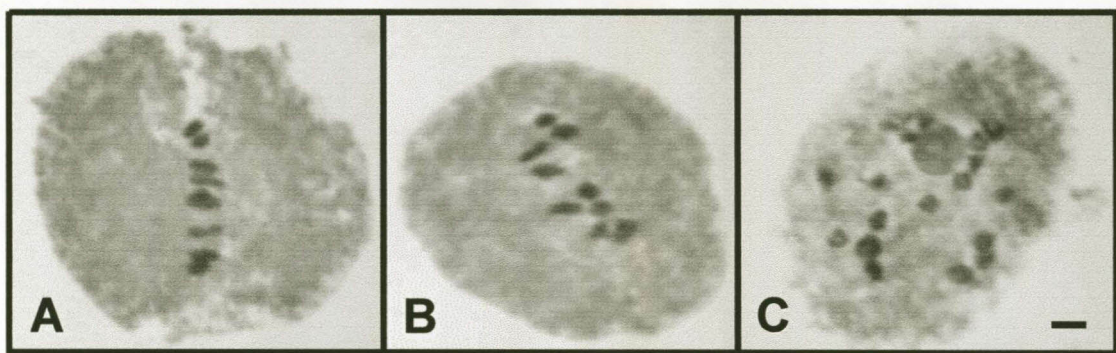
*Eleusine* Gaertn. is predominantly an African genus, with six of the nine species confined to tropical and subtropical Africa (Phillips 1972). *Eleusine coracana* (L.) Gaertn. is widely grown in Africa, India and China, and used as a cereal. It is derived from *E. indica* (L.) Gaertn., a diploid cosmopolitan weed (subsp. *indica*,  $2n = 18$ ), which has a tetraploid race in Africa (subsp. *africana*). The morphological characters of the two races overlap greatly, and this leads to their inclusion in a single species (Clayton and Renvoize 1986). *Eleusine coracana* (L.) Gaertn. subsp. *africana* (Kenn.-O'Byrne) Hilu & DeWet (= *E. indica* subsp. *africana*) is native to Africa, where it is widespread along the eastern highlands and the highlands of the southern African plateau (Phillips 1972).

Three *E. coracana* subsp. *africana* specimens were investigated. The two specimens from Cape Vidal were diploid (Figure 3.5A, B) and the specimen from Koekenaap in Western Cape was tetraploid ( $n = 2x = 18$ ) (Figure 3.5C).

This indicates that diploid and tetraploid forms of this species are present in South Africa and confirm a basic chromosome number of nine for this genus



**Figure 3.4.** Photomicrographs of meiotic chromosomes in the genus *Leptochloa*. **A–E.** *L. fusca*. **A, B.** *Spies* 3794,  $2n = 2x = 20$ , anaphase I with 10-10 segregation (**A**) and metaphase I (**B**). **C.** *Spies* 3932,  $2n = 2x = 20$ , metaphase I. **D.** *Spies* 4316,  $2n = 2x = 20$ , diakinesis with  $10_{II}$ . **E.** *Spies* 5200,  $2n = 2x = 20$ , metaphase I. Scale bar: 5  $\mu$ m.



**Figure 3.5.** Photomicrographs of meiotic chromosomes in the genus *Eleusine*. **A–C.** *E. africana* subsp. *africana*. **A, B.** *Spies* 2365,  $2n = 2x = 18$ , metaphase I. **C.** *Spies* 2783,  $2n = 4x = 36$ , diakinesis with  $18_{II}$ . Scale bar: 5  $\mu$ m.

(Darlington and Wylie 1955; Pienaar 1955; Ornduff 1967, 1968, 1969; Federov 1969; Moore 1970, 1971, 1972, 1974, 1977; Goldblatt 1981, 1983, 1985, 1988; Goldblatt and Johnson 1990, 1991, 1994, 1998, 2000).

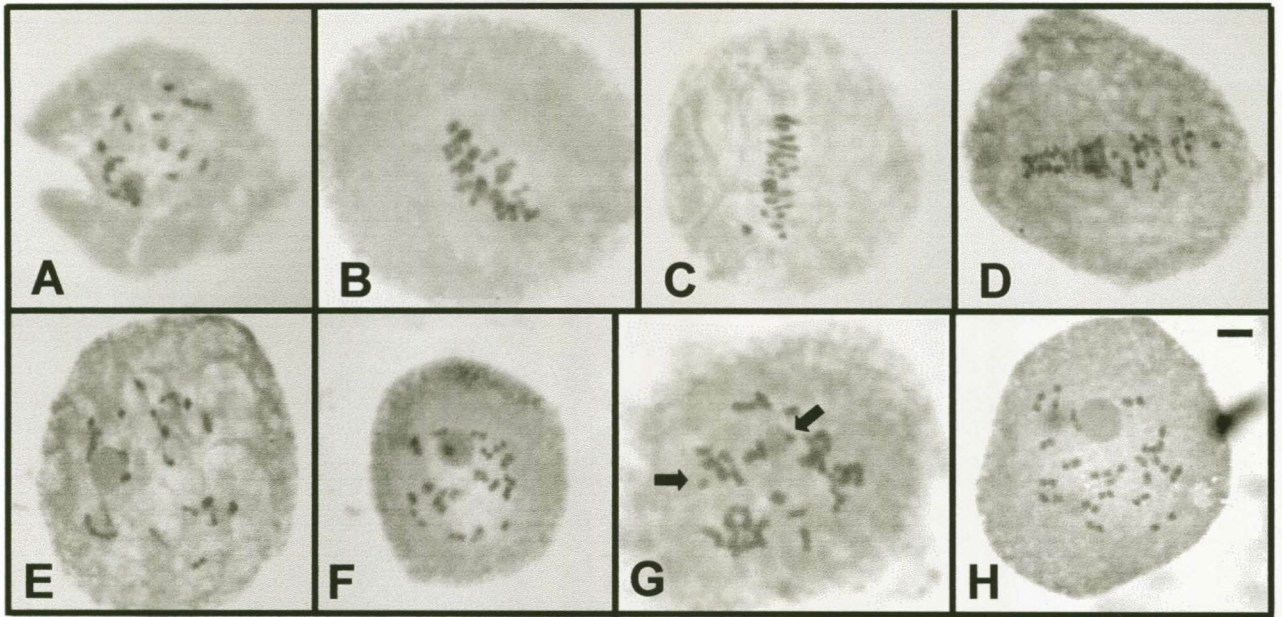
*Eragrostis* Wolf is the largest genus in the subfamily Chloridoideae. It has a worldwide occurrence in the warmer regions where it is found in most habitats, showing a preference for open sites, poor dry soil and weedy places (Clayton and Renvoize 1986). The genus exhibits the full range of morphological and anatomical variation found in the subfamily (Van den Borre and Watson 1994).

*Eragrostis* is the largest grass genus in southern Africa with  $\pm$  90 species. In this study 26 specimens were investigated, representing 17 species. Twenty of the specimens were tetraploid (Figure 3.6A–C, E, F, H). Only four specimens investigated were diploid and two were hexaploid (Figure 3.6D, G). Polyploidy is frequent in this genus as can be seen from the results presented. Tetraploidy, as in this study, is the most frequent polyploid level observed, followed by diploidy (Darlington and Wylie 1955; Pienaar 1955; Ornduff 1967, 1968, 1969; Moore 1970, 1971, 1972, 1973, 1974, 1977; Goldblatt 1981, 1983, 1985, 1988; Goldblatt and Johnson 1990, 1991, 1994, 1998, 2000).

Seven specimens were investigated for *E. capensis* (Thunb.) Trin. All but one was tetraploid (Figure 3.6A–C). *Spies* 4696 was diploid and is the second report for this species (De Wet 1958) where tetraploidy (Avdulov 1931; Pienaar 1953; Davidse *et al.* 1986; Spies and Du Plessis 1986a; Spies *et al.* 1991) and hexaploidy (Moffet and Hurcombe 1949; Spies and Voges 1988) have previously been observed.

De Winter (1955) regards *E. curvula* (Schrad.) Nees as the most variable species in the genus in southern Africa, with a great many morphological forms. This was corroborated by large-scale cytogenetic studies by Vorster and Liebenberg (1977). Several species are known to intergrade with *E. curvula* through hybridization: *E. barbinodes* Hack., *E. caesia* Stapf, *E. chloromelas* Steud., *E. lehmanniana* Nees, *E. planiculmis* Nees and *E. rigidior* Pilg. (Smook 1990). Proof of hybridization in this species indicates a collapse of isolating mechanisms between different species in the *Eragrostis curvula* complex, resulting in a large-scale hybrid swarm, with continuous variation of characters between parental extremes. The variation in morphological characters is an indication of the extent of hybridization





**Figure 3.6.** Photomicrographs of meiotic chromosomes in the genus *Eragrostis*. **A–C.** *E. capensis*. **A.** Spies 3483,  $2n = 4x = 40$ , diakinesis with  $20_{II}$ . **B.** Spies 3498,  $2n = 4x = 40$ , metaphase I. **C.** Spies 5069,  $2n = 4x = 40$ , metaphase I. **D.** *E. curvula*, Spies 1137,  $2n = 6x = 60$ . **E.** *E. echinochloidea*, Spies 2799,  $2n = 4x = 40$ , diakinesis with  $20_{II}$ . **F.** *E. inamoena*, Spies 2392,  $2n = 4x = 40$ , diakinesis with  $20_{II}$ . **G.** *E. planiculmis*, Du Plessis 116,  $2n = 6x = 60+0-4B$ , diakinesis with  $30_{II}$ . Two B chromosomes are indicated. **H.** *E. superba*, Du Plessis 136,  $2n = 4x = 40$ , diakinesis with  $20_{II}$ . Scale bar: B, G = 10  $\mu$ m; A, C–F, H = 12  $\mu$ m.

(Spies 1984). Four specimens in this complex were cytogenetically investigated and tetraploid (*E. barbinodes* and *E. chloromelas*) and hexaploid (*E. curvula* and *E. planiculmis*) levels were observed (Figure 3.6D, G). In all but *E. barbinodes*, various univalents were observed which resulted in laggards and later formed micronuclei. According to Church (1929), the presence of unpaired or univalent chromosomes is one of the most prominent suggestions that a plant is of hybrid origin and therefore, in this complex with its large-scale hybridization, these phenomena will be very prevalent.

This is the third report for the species *E. heteromera* Stapf with  $2n = 4x = 40$  (De Wet 1958, 1960). As far as is known only two reports for *E. tef* (Zucc.) Trotter exist (Avdulov 1931; Moffet and Hurcombe 1949). This study corroborates

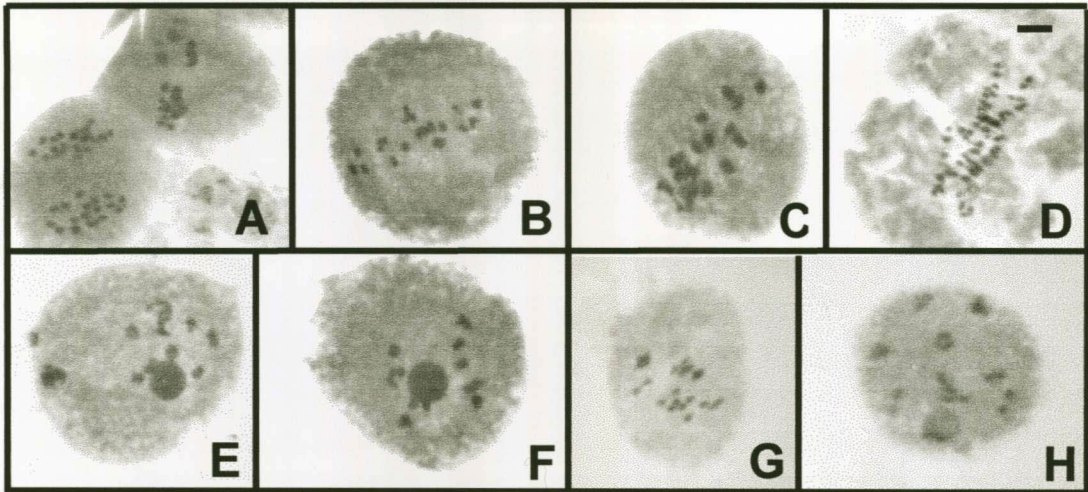


tetraploidy present in this species. Previously only tetraploid levels have been observed in the species *E. tenuifolia* (A.Rich.) Steud. (Ornduff 1967; Moore 1973, 1977; Goldblatt 1983). Here we report on a diploid specimen from Siteki in Swaziland. Univalents were observed in some cells in this specimen. The tetraploid chromosome count for *E. trichophora* Coss. & Dur. reported here is second for the species (Davidse *et al.* 1986) and a hexaploid specimen was reported by Spies and Jonker (1987). One previous report of  $2n = 4x = 40$  for *E. racemosa* (Thunb.) Steud. (Ornduff 1967) is confirmed in this study where three tetraploid specimens were found.

Two *Fingerhuthia africana* Lehm. specimens were investigated and were tetraploid ( $n = 2x = 20$ ) (Figure 3.7A, B). Previous reports by Spies and Du Plessis (1987) and De Wet (1958, 1960) also found tetraploidy in the genus, but diploidy has been reported (De Wet 1958, 1960) as well.

*Odyssea* Stapf is a xerophytic grass genus with two species, one indigenous to southern Africa (Clayton and Renvoize 1986). It has a very distinct, much-branched, spiny habit, which is an adaptation to its specialized sandy and saline habitats (Phillips 1982). This is a first report for the genus. The specimen, *O. paucinervis* (Nees) Stapf, was tetraploid ( $n = 2x = 18$ ) and, therefore the basic chromosome number for this genus is 9.

The genus *Sporobolus* R.Br. is cytogenetically complex and different basic chromosome numbers,  $x = 6, 9$  and  $10$  may be present (Davidse *et al.* 1986). Four *Sporobolus* species were investigated. Three diploid [*S. africanus* (Poir.) Robyns & Tournay, Spies 4508, *S. ioclados* (Trin.) Nees, *S. virginicus* (L.) Kunth], one tetraploid (*S. africanus*, Spies 2369) and one hexaploid (*S. albicans* Nees) specimen were found (Figure 3.7C–F). They all displayed multiples of nine and this confirms  $x = 9$  as the basic chromosome number for the genus (Darlington and Wylie 1955; Pienaar 1955; Ornduff 1967, 1968, 1969; Federov 1969; Moore 1970, 1971, 1972, 1973, 1974, 1977; Goldblatt 1981, 1983, 1985, 1988; Goldblatt and Johnson 1990, 1991, 1994, 1996, 1998). A chromosome number for *S. albicans* has not been published yet and this constitutes a first report for this species (Figure 3.7D). Large-scale studies in this genus are still necessary to investigate the different basic chromosome numbers present and their possible phylogenetic relationships.



**Figure 3.7.** Photomicrographs of meiotic chromosomes in the genera *Fingerhuthia*, *Sporobolus* and *Stiburus*. **A, B.** *Fingerhuthia africana*, Spies 2947,  $2n = 4x = 40$ , late anaphase I (**A**) and metaphase I (**B**). **C.** *S. africanus*, Spies 2369,  $2n = 4x = 36$ , metaphase I. **D.** *Sporobolus albicans*, Spies 3141,  $2n = 6x = 54$ , metaphase I. **E, F.** *S. virginicus*, Du Plessis 122,  $2n = 2x = 18$ , diakinesis with  $9_{II}$ . **G, H.** *Stiburus conrathii*, Du Plessis 19,  $2n = 2x = 20$ , metaphase I (**G**), diakinesis with  $10_{II}$  (**H**). Scale bar:  $8 \mu\text{m}$ .

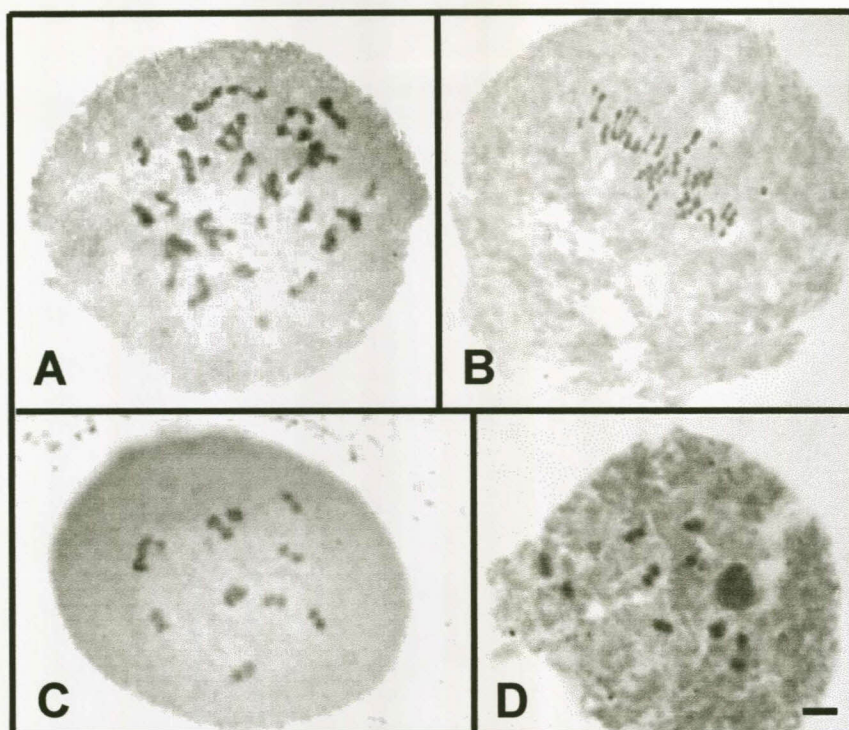
One specimen each of the two species *Stiburus alopecuroides* (Hack.) Stapf and *S. conrathii* Hack. was examined. Both were diploid ( $n = x = 10$ ) (Figure 3.7G, H) and this confirms chromosome counts based on multiples of 10 for this genus (Spies and Du Plessis 1986a).

A single *Trichoneura grandiglumes* (Nees) Ekman specimen was found with a chromosome count of  $n = x = 10$ . Moffet and Hurcombe (1949) and De Wet and Anderson (1956) also reported on diploid specimens.

### Tribe Pappophoreae

The genus *Enneapogon* Desv. ex P.Beauv. is a very uniform genus in which most species (28 in total) closely resemble one another. Four specimens representing two species and one unidentified specimen were examined in this study (Figure 3.8A–D). One specimen, *Enneapogon cenchroides* (Roem. & Schult.) C.E.Hubb. was tetraploid with  $n = 2x = 20$ , but another specimen was hexaploid





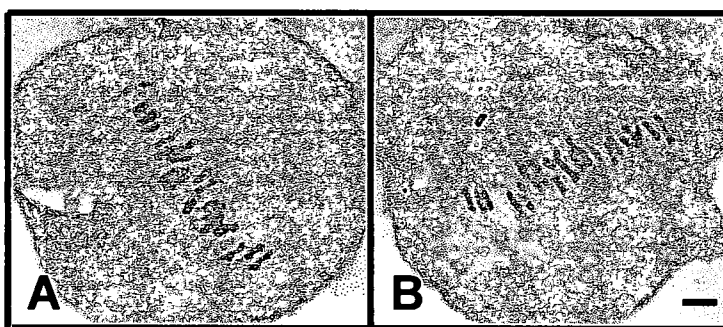
**Figure 3.8.** Photomicrographs of meiotic chromosomes in the genus *Enneapogon*. **A, B.** *E. cenchroides*, *Spies 2709*,  $2n = 6x = 60$ , diakinesis with  $30_{II}$  (**A**) and metaphase I (**B**). **C, D.** *E. pretoriensis*, *Spies 3716*,  $2n = 2x = 20$ , diakinesis with  $10_{II}$ . Scale bar: A, C, D = 5  $\mu\text{m}$ ; B = 8  $\mu\text{m}$ .

(Figure 3.8A, B). *Spies 5532*, the unidentified specimen, was also found to be tetraploid. A single *E. pretoriensis* Stent specimen had  $n = x = 10$  (Figure 3.8C, D). Two basic chromosome numbers are reported for this genus,  $x = 9, 10$  [Darlington and Wylie ( $x = 9, 10$ ); Ornduff 1968 ( $x = 10$ ), 1969 ( $x = 10$ ); Moore 1970 ( $x = 10$ ), 1977 ( $x = 10$ ); Goldblatt 1981 ( $x = 10$ ), 1985 ( $x = 10$ ); Goldblatt and Johnson 1991 ( $x = 10$ ), 1998 ( $x = 10$ )]. The majority of the studies support a basic chromosome number of ten. Only three studies, De Wet (1954a), Thomas unpublished (listed in Darlington and Wylie 1955) and De Wet and Anderson (1956) ever reported  $x = 9$ . They found one diploid (De Wet and Anderson 1956) and five tetraploid [De Wet 1954a; Thomas unpublished (listed in Darlington and Wylie 1955); De Wet and Anderson 1956] specimens based on  $x = 9$ . Davidse *et al.* (1986) also reported on aneuploidy in the genus.

The genus *Schmidtia* Steud. ex J.A.Schmidt comprises only two species, both widespread in southern Africa. Two specimens were examined representing *S. pappophoroides* Steud. and an unidentified *Schmidtia* species. Both were tetraploid but *Spies* 5536 had 0-4 B chromosomes present in some cells. This confirms the basic chromosome number of 9 for this genus (De Wet and Anderson 1956; De Wet 1958), although Reeder and Singh (1968) reported on a basic chromosome number of ten.

### Unplaced

*Centropodia* Rchb. was formerly recognized as an arundinoid genus but has recently (GPWG 2001) been included in the subfamily Chloridoideae. A single specimen of this genus was investigated and found to be octaploid ( $n = 4x = 24$ ) (Figure 3.9A, B). This confirms the basic chromosome number of the genus as six (Du Plessis and Spies 1988; Hoshino and Davidse 1988).



**Figure 3.9.** Photomicrographs of meiotic chromosomes in the genus *Centropodia*. **A, B.** *C. glauca*, *Spies* 5706,  $2n = 8x = 48$ , metaphase I. Scale bar: 8  $\mu\text{m}$ .

## 3.3 Conclusions

Chromosome numbers are reported for three of the five tribes of the subfamily Chloridoideae. Basic chromosome numbers of  $x = 9$  and  $10$  occur in all the tribes. A basic chromosome number of six is also corroborated for the genus *Centropodia*. The high incidence of polyploidy (65% in this study) in Poaceae and especially the southern African grasses are once again confirmed by this study.

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## CHAPTER 4

# Basic chromosome numbers

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IN PRESS (TAXON):

R. ROODT AND J.J. SPIES. CHROMOSOME STUDIES IN THE GRASS  
SUBFAMILY CHLORIDOIDEAE. I. BASIC CHROMOSOME NUMBERS. IN PRESS  
(TAXON)

Two main basic chromosome numbers occur in the majority of southern African Chloridoideae, i.e. 9 and 10, the majority being  $x = 10$ . Two main evolutionary pathways, one following  $x = 10$  and the other following an aneuploid/dysploid reduction to  $x = 9$  are suggested for the subfamily. Evidence indicates that  $x = 9, 10$  are paleopolyploid basic chromosome numbers, with  $x = 10$  derived from  $x = 5$ . This corresponds to the closely related panicoid tribe Andropogoneae where five is accepted as the basic chromosome number.

## 4.1 Introduction

Avdulov (1931) was the first to recognize the taxonomic significance of the grass chromosomes. He studied 232 species and recognized three types of grass karyotypes, based on basic chromosome number ( $x$ ) and chromosome size: the first type has  $x = 9, 10$  with small chromosomes; the second has  $x = 12$  with small chromosomes; and the third  $x = 7$  with large chromosomes.

Stebbins (1956) considered four main lines of evolution in the grasses on the bases of morphological and cytogenetic characters. The panicoid and the eragrostoid-chloridoid lines ( $x = 9, 10$ ) developed in the tropics and show a reduction in the number of florets and specialization in leaf anatomy. The third line is the Pooideae Benth. (= Festucoideae Link) ( $x = 7$ ), which are the principle grasses of temperate climates with the largest chromosomes in the family. A fourth line was Bambuseae Dumort., which became dominant in the moist tropical forests ( $x = 12$ ). Three additional smaller lines, which were also seen as successful, were the arundinoid ( $x = 6$ ), oryzoid ( $x = 12$ ) and stipoid lines ( $x = 6$ ). Smaller groups containing one or a few genera, such as *Anomochloa* Brongn. ( $x = 9$ ), *Nardus* L. ( $x = 13$ ) and *Pariana* Aubl. ( $x = 11$ ), were also recognized.

In 1961, Carnahan and Hill proposed that grasses fall into two groups:

- Tropical and subtropical species with small chromosomes and  $x = 10$ , or derivatives thereof, such as 9 and 12.
- Species with a more temperate distribution that have larger chromosomes and usually a basic chromosome number of 7.

According to McWilliam (1964), the different basic chromosome numbers in Poaceae (R.Br.) Barnhart reflect taxonomic groupings in the family. This is confirmed by some of the most recent classifications, for example Clayton and Renvoize (1986), who recognized six subfamilies: Bambusoideae Asch. & Graebn. ( $x = 12$ ), Centothecoideae Soderstr. ( $x = 12$ ), Pooideae Macfarlane & Watson ( $x = 7$ ), Arundinoideae Tateoka ( $x = 6, 7, 13$ ), Chloridoideae Rouy ( $x = 9, 10$ ) and Panicoideae A.Braun ( $x = 9, 10$ ). Though similar basic chromosome numbers occur in different subfamilies, these numbers are not necessarily monophyletic (Panicoideae and Chloridoideae have predominantly  $x = 9$  and  $10$  and both Arundinoideae and Pooideae have  $x = 7$ ), but similar chromosome numbers with similar morphologies may be monophyletic (chloridoid chromosomes are smaller than the panicoid chromosomes and the arundinoid chromosomes are medium sized in comparison to the very large chromosomes of the Pooideae). Currently 12 subfamilies are recognized [Grass Phylogeny Working Group (GPWG) 2001] based on morphology and molecular data, but basic chromosome numbers still indicate the major evolutionary diversification in the Poaceae.

The two subfamilies with both 9 and 10 as basic chromosome numbers in the majority of the genera present in the groups are the Chloridoideae and Panicoideae. Evidence indicating a close phylogenetic relationship between these two subfamilies, except for basic chromosome number, includes immunological affinities (Esen and Hilu 1989), similar prolamin profiles and prolamin sizes (Hilu and Esen 1988), length of the embryo, lodicule structure and nature of the first leaf of the seedling (Hilu and Johnson 1991). Hubbard (1948) and Stebbins (1956) proposed that the Chloridoideae and Panicoideae came from closely related ancestors and Tateoka (1957) and Clayton (1981), suggested that the ancestor to these two groups might be arundinoid. From the Arundinoideae with its basic chromosome number of 6, a possible aneuploid reduction could have lead to five in the Chloridoideae and Panicoideae. This was increased by polyploidization to the paleopolyploid basic chromosome number of ten and subsequent aneuploid reduction lead to nine. This has been corroborated by various molecular studies (Barker *et al.* 1995, 1999; Hilu *et al.* 1999; Hilu and Alice 2001).

Goldblatt (1980) was of the opinion that high basic chromosome numbers, for example  $x \geq 9$ , are not of recent origin and most probably had polyploidy in their

evolutionary history. Taxa with these secondary basic chromosome numbers, therefore the Chloridoideae and Panicoideae, would be paleopolyploids, or ancient taxa that are rediploidized polyploids with high basic chromosome numbers (Grant 1963; Goldblatt 1980). According to Feldman *et al.* (1997), this diploidization process occurs by means of chromosome diploidization in which homoeologous chromosome pairing is suppressed and also by means of genetic diploidization. The genetic diploidization process includes various changes such as for example, structural rearrangements at various levels (Song *et al.* 1995; Wendel *et al.* 1995a; Leitch and Bennett 1997), the regulation of gene expression (Galitski *et al.* 1999; Matzke *et al.* 1999) and the amplification, reassortment or elimination of various types of sequences (Feldman *et al.* 1997; Hanson *et al.* 1998).

Rhoades (1951) first proposed that maize (Panicoideae) ( $2n = 2x = 20$ ), contains duplicated genes, and may be the result of ancient polyploidy from a  $2n = 2x = 10$  ancestor. This is strongly supported by isozyme evidence (Wendel *et al.* 1985, 1986, 1989) and DNA marker studies (Helentjaris *et al.* 1988). Structural changes in the maize genome include rearrangement and gene silencing (Gaut and Doebley 1997; White and Doebley 1998). The proposal of ancient polyploidy was later extended to other genera in the panicoid tribe Andropogoneae Dumort. (Garber 1950; Celarier 1956; Mehra and Sharma 1975; Clayton and Renvoize 1986). Molecular support for this includes RFLP studies on *Sorghum* Moench (Chittenden *et al.* 1994; Pereira *et al.* 1994) and molecular cytogenetic and linkage analysis studies in sugarcane (D'Hont *et al.* 1996; Ming *et al.* 1998). Comparative mapping in maize and sorghum renders evidence in support of an ancient polyploid origin (Whitkus *et al.* 1992). Further comparative genetic studies have shown that maize, sugar cane, sorghum, pearl millet, and foxtail millet share a common genome arrangement (Gale and Devos 1998; Devos and Gale 1997, 2000; Kellogg 1998b).

Could a similar phenomenon of ancient polyploidy and a basic chromosome number of five be present in the Chloridoideae, the subfamily with which the Panicoideae shares large affinities? Unlike the Panicoideae, which consists of a number of economically important genera and has been thoroughly investigated, studies in the Chloridoideae relating to ancient polyploidy is limited. These two subfamilies are seen as the most recent radiation in Poaceae (Hsiao *et al.* 1998; GPWG 2001), both of which have diversified into the tropical and subtropical



regions, especially the southern Hemisphere (Renvoize and Clayton 1992) and compose the largest majority of the southern African grass flora (Gibbs Russell 1986).

It is the aim of this study to investigate chromosome numbers of various southern African genera and species of the subfamily Chloridoideae, to determine and corroborate basic chromosome numbers, and by giving a review of chromosome numbers found in southern Africa and worldwide, to indicate that ancestral polyploidization has also lead to paleopolyploid basic chromosome numbers in the subfamily as is seen in the closely related Panicoideae.

## 4.2 Results

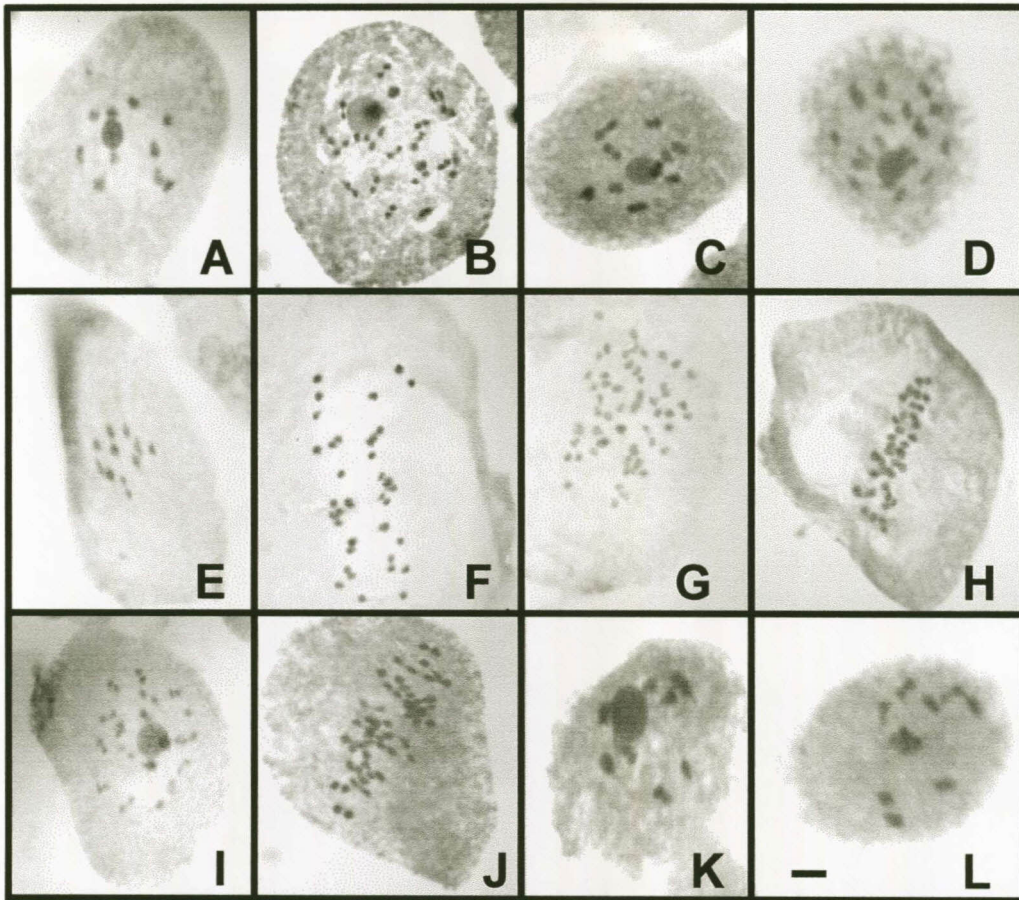
The major studies dealing with the chromosome numbers of South African chloridoid grasses (Figure 4.1) are those of Avdulov (1931); Hunter (1934); Nielson (1939); Moffet (1944); Hurcombe (1946, 1947); Moffett and Hurcombe (1949); Pienaar (1953); De Wet (1954a, b, 1958, 1960); De Wet and Anderson (1956); Reeder and Singh (1968); Nordenstam (1969); Vorster and Liebenberg (1977); Jones *et al.* (1978), Nordenstam (1982); Spies (1982); Davidse *et al.* (1986); Spies and Du Plessis (1986a, b, 1987); Spies and Jonker (1987); Du Plessis and Spies (1988); Hoshino and Davidse (1988); Spies and Gibbs Russell (1988); Spies and Voges (1988); Spies *et al.* (1991); Strydom and Spies (1994b) and the present study (Appendix A, B). For a list of worldwide studies see Appendix C. In these tables and in the text no reference is made to aneuploid deviations in somatic chromosome numbers and these were not considered further. This is to make interpretations easier, but these deviations do occur in various genera such as *Bouteloua* Lag., *Chloris* Sw., *Cynodon* Rich., *Dactyloctenium* Willd., *Hilaria* Kunth, *Muhlenbergia* Schreb. and *Sporobolus* R.Br.

In the seventy-nine specimens investigated in this study (Appendix A), 67% of the species have the paleopolyploid basic chromosome number of 10 (Figure 4.2A–E, L), with only 32% of the species investigated with  $x = 9$  (Figure 4.2F–K). Three other basic chromosome numbers which also occur in the Chloridoideae are  $x = 6, 7, 8$ . These chromosome numbers, however, occur only in a small percentage of genera and are, therefore not considered main basic chromosome numbers.



**Figure 4.1.** Herbarium specimen of *Chloris virgata* Sw., a typical member of Chloridoideae. Scale bar: 3cm.

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**Figure 4.2.** Photomicrographs of meiotic chromosomes in some genera of the subfamily Chloridoideae. **A.** *Cladoraphis cyperoides*, Spies 4889,  $2n = 2x = 20$ , diakinesis with  $10_{II}$ . **B.** *C. spinosa*, Spies 4885,  $2n = 4x = 40$ , diakinesis with  $20_{II}$ . **C.** *Enneapogon pretoriensis*, Spies 3716,  $2n = 2x = 20$ , diakinesis with  $10_{II}$ . **D.** *Enteropogon macrostachyus*, Venter 9339,  $2n = 4x = 40$ , diakinesis with  $20_{II}$ . **E.** *Eragrostis tenuifolia*, Spies 2595,  $2n = 2x = 20$ , diakinesis with  $10_{II}$ . **F, G.** *Harporchloa falx*, Spies 4729,  $2n = 4x = 36$ , anaphase I segregation (**F**), Spies 5128,  $2n = 6x = 54$ , anaphase I segregation (**G**). **H.** *Odyssea paucinervis*, Spies 3384,  $2n = 4x = 36$ , metaphase I. **I.** *Schmidtia pappophoroides*, Du Plessis 186,  $2n = 4x = 36$ , diakinesis with  $18_{II}$ . **J.** *Sporobolus albicans*, Spies 3141,  $2n = 6x = 54$ , metaphase I. **K.** *Sporobolus virginicus*, Du Plessis 122,  $2n = 2x = 18$ , diakinesis with  $9_{II}$ . **L.** *Stiburus conrathii*, Du Plessis 19,  $2n = 2x = 20$ , diakinesis with  $10_{II}$ . Scale bar: A, B, D = 5  $\mu$ m; C, L = 3  $\mu$ m; E–H = 6.5  $\mu$ m; I, J = 10  $\mu$ m; K = 8  $\mu$ m.



The basic chromosome number of six was only observed in one species, namely *Centropodia glauca* (Nees) Cope.

New chromosome numbers, reported for the first time or where new numbers deviate from previous reports, for southern African species found in this study is the following: *Cladoraphis cyperoides* (Thunb.) S.M.Phillips ( $2n = 2x = 20$ ) (Figure 4.2A); *C. spinosa* (L.f.) S.M.Phillips ( $2n = 4x = 40$ ) (Figure 4.2B); *Enneapogon pretoriensis* Stent ( $2n = 2x = 20$ ) (Figure 4.2C); *Enteropogon macrostachyus* (A.Rich.) Benth. ( $2n = 4x = 40$ ) (Figure 4.2D); *Eragrostis tenuifolia* (A.Rich.) Steud. ( $2n = 2x = 20$ ) (Figure 4.2E); *Harpochloa falx* (L.f.) Kuntze ( $2n = 4x = 36$ ;  $2n = 6x = 54$ ) (Figure 4.2F, G); *Odyssea paucinervis* (Nees) Stapf ( $2n = 4x = 36$ ) (Figure 4.2H); *Schmidtia pappophoroides* Steud. ( $2n = 4x = 36$ ) (Figure 4.2I); *Sporobolus albicans* Nees ( $2n = 6x = 54$ ) (Figure 4.2J); *Sporobolus virginicus* (L.) Kunth ( $2n = 2x = 18$ ) (Figure 4.2K) and *Stiburus conrathii* Hack. ( $2n = 2x = 20$ ) (Figure 4.2L).

From these somatic chromosome numbers, a paleopolyploid basic chromosome number of ten for the species *Cladoraphis cyperoides*, *C. spinosa*, *Enteropogon macrostachyus*, *Eragrostis tenuifolia* and *Stiburus conrathii* is accepted. A paleopolyploid basic chromosome number of 9 is indicated for the species *Odyssea paucinervis* and *Sporobolus albicans*. New evidence for the occurrence of both basic chromosome numbers is found for the southern African representatives of the genera *Enneapogon* and *Harpochloa* and the species *Schmidtia pappophoroides* and *Sporobolus virginicus*.

### 4.3 Discussion

In the Chloridoideae and the Panicoideae the most common paleopolyploid basic chromosome numbers are  $x = 9$  and  $x = 10$ . Stebbins (1985) was of the opinion that  $x = 5$ , 6 or 7 had equal probability of being the original basic chromosome number of the grasses. He proposed that from these widespread original basic chromosome numbers, the paleopolyploid numbers of  $x = 10$  and  $x = 12$  in different groups (such as the Chloridoideae and Panicoideae) arose (Stebbins 1985). This is in contrast to Raven (1975) who proposed that  $x = 10$ , 11 and 12 are the more primitive basic chromosome numbers in the grasses. Stebbins (1985) gave the following reasons in support of  $x = 10$ , 11 and 12 being derived by

polyploidy:

- The basic chromosome numbers of  $x = 5, 6$  appear in a few distantly related taxa that are among the more primitive genera in their respective subfamilies, such as *Danthonia* DC. ( $x = 6$ ) and *Sorghum* ( $x = 5$ ).
- Furthermore,  $x = 7, 6$ , and  $5$  cannot be derived by aneuploid reduction from  $x = 12$ , as proposed by Raven (1975), due to the extreme rarity of  $x = 8$  in the grasses, and the rarity of  $x = 9$ , except in the highly specialized members of the Chloridoideae and Panicoideae.

In 1987, De Wet reported on diploids,  $2n = 2x = 10$ , in the panicoid genera *Sorghum* and *Thelepogon* Roth ex Roem. & Schult. As early as 1950 and 1956, Garber (1950) and Celarier (1956), had already proposed that 5 was the basic chromosome number of the tribe Andropogoneae (Panicoideae). By 2000, Watson & Dallwitz (1992 onwards), reported a basic number of 5 for the panicoid genera *Andropogon* L., *Coix* L., *Cymbopogon* Spreng., *Elionurus* Kunth ex Willd., *Sorghum*, *Thelepogon*, and *Zea* L. (based on studies by for example Harlan *et al.* 1952; Sisodia 1970; Dujardin 1978a; Wu 1978; Sapre and Barve 1983; Molina and Naranjo 1986, 1987; Lavania 1987; Sapre and Deshpande 1987; Morakinyo and Olorode 1988; Spies *et al.* 1991; Rao and Nirmala 1994).

According to De Wet (1987), diploids ( $2n = 2x = 10$ ) seem to be absent from the Chloridoideae. In 1963, however, Swami found *Oropetium thomaeum* Trin. in India and Pohl (1969) found a specimen of *Muhlenbergia andina* Hitchc. in North America, both with  $2n = 2x = 10$ . In 1986, Davidse *et al.* found *Dactyloctenium giganteum* Fisch. & Schweick ( $2n = 2x = 10$ ) in Zimbabwe. This is evidence of a basic chromosome number of five. This was first proposed in 1965 by Roy when she observed secondary association of bivalents into five groups during meiosis in the Indian species *Eragrostis diarrhena* Steud. ( $2n = 4x = 20$ ).

Both the Chloridoideae and Panicoideae have been proposed to have an African origin (Hartley 1958b; Hartley and Slater 1960), more specifically in the East Africa-Madagascar region (Hartley 1958b). This is based on biogeographical evidence suggesting that the Panicoideae originated in the East-Africa-Madagascar region. The occurrence of the genus *Miscanthus* Anderss. in southern Africa, a

relatively primitive genus in the Panicoideae, supports the possible origin of the Panicoideae in this region (Hartley 1958b). Paleontological evidence that support this are Miocene fossil grasses belonging to the subfamilies Chloridoideae and Panicoideae in East Africa (Retallack *et al.* 1990). Five distinctly different plant morphologies, arrangements of stomata, phytoliths and hair bases were found among these fossil grasses. Three of these were identified as three different species of the subfamily Panicoideae and the remaining two as two separate species of the subfamily Chloridoideae. This renders evidence as to the presence of grasses and grasslands in East Africa for at least 14 million years (Retallack *et al.* 1990). The geographic distribution of the chloridoid genera *Triraphis* R.Br. and *Centropodia*, the most basal members of the subfamily in an analysis of the subfamily based on the *matK* gene (Hilu and Alice 2001), further supports the African origin hypothesis of Hartley (1964) for the Chloridoideae.

According to Hartley and Slater (1960) the differentiation of the Chloridoideae probably preceded that of the Panicoideae because the Panicoideae have not reached a balanced distribution in the different hemispheres (Hartley 1958a, b). Therefore, if the basic chromosome number of the Panicoideae is considered to be five, the same can be proposed for the Chloridoideae, even though less primary diploids have been found which is probably only due to sampling bias.

Therefore, five is the basic chromosome number for the subfamily Chloridoideae, and many current diploid species ( $2n = 2x = 20$ ) are paleotetrapolyploids ( $2n = 4x = 20$ ), etc. The scarcity of specimens with  $n = x = 5$  in the subfamily Chloridoideae indicates that a large number of diploids have become extinct (Hunziker and Stebbins 1987). According to Stebbins (1985), polyploidy and trends towards aneuploidy/dysploidy and a high degree of specialization with regard to some morphological characteristics originated quickly and shortly after the grasses first differentiated. Consequently, these early stages of grass evolution produced many genera and species that are now completely extinct. De Wet (1987) suggested that  $x = 9$  was probably derived via aneuploidy from  $x = 10$ , which in turn was derived from  $x = 5$ . Whether the reduction is due to aneuploidy or dysploidy (fusion or Robertsonian translocation) is not known.

The genus *Sporobolus* is the only chloridoid genus in southern Africa in which

both main basic chromosome numbers,  $x = 9, 10$  and the less common basic chromosome number  $x = 6$ , occur (Moffet and Hurcombe 1949; De Wet 1954a; De Wet and Anderson 1956; De Wet 1958, 1960; Davidse *et al.* 1986; Spies and Du Plessis 1986b; Spies and Jonker 1987; Spies and Voges 1988; Spies *et al.* 1991).

Hubbard (1934) stated that *Sporobolus* shows a close relationship to *Eragrostis* Wolf (which has a paleopolyploid basic chromosome number of  $x = 10$ ) and is probably derived from it. Brown (1950) also proposed that the basic chromosome number of  $x = 9$  in *Sporobolus* was probably derived from *Eragrostis* with  $x = 10$  by a reduction in basic chromosome number. This was before Roy (1965) reported on the basic chromosome number of *Eragrostis* as 5. Reports for diploids based on  $x = 6$  (none, however, based on  $x = 5$  for this genus), have been reported for *Sporobolus maderaspatanus* Bor ( $2n = 2x = 12$ ) from India (Christopher and Samraj 1985). *Sporobolus molleri* Hack. with  $2n = 2x = 12$  has been found in Zaire and Uganda (Tateoka 1965a; Auquier and Renard 1975; Dujardin 1978b, 1979) and diploids for another *Sporobolus* species, *S. tenuissimus* Kuntze ( $2n = 2x = 12$ ) are reported from Madagascar, Trinidad and Zaire (Tateoka 1965b; Davidse and Pohl 1972; Dujardin 1978b). The presence of *Sporobolus* with  $x = 6$  in the region of origin of the subfamily and in the region where diploids based on  $x = 5$  have also been reported, supports the likelihood that initially a *Sporobolus* lineage was derived from *Eragrostis* by a aneuploid increase in basic chromosome number from 5 to 6. Other chromosome numbers in *Sporobolus* probably arose by polyploidy ( $n = x = 5$  to  $n = x = 10$ ) (Christopher and Abraham 1974) and subsequent aneuploidy/dysploidy ( $n = x = 9$ ). This is supported by the fact that no *Sporobolus* species based on  $x = 5$  have been found. It seems likely that *Sporobolus* species with  $x = 6$  represents the older species in *Sporobolus* (some probably extinct), whereas the species with  $x = 9$  are derived from the paleopolyploids based on  $x = 10$ , and therefore younger members of the genus derived after initial polyploidization events.

The close relationship between *Eragrostis* and *Sporobolus* is also noticeable at morphological level, where the major difference is the number of florets per spikelet (Clayton and Renvoize 1986). It may be that *Sporobolus* represents a specialized lineage of the genus *Eragrostis* with the differentiation of 3 basic chromosome numbers as opposed to the single basic chromosome number of *Eragrostis*.

The other southern African taxa having  $x = 6$  is the genus *Centropodia* [ $2n = 8x = 48$  for *C. glauca* and  $2n = 4x = 24$  for *C. forskalii* (Vahl) Cope] and the species *Merxmuellera rangei* (Pilg.) Conert ( $2n = 6x = 36$ ) (De Wet 1954b; Du Plessis and Spies 1988; Hoshino and Davidse 1988; present study).

*Centropodia* has a well-developed Kranz anatomy (Ellis 1984b), which separates it from the Arundinoideae (Danthonioideae Barker & H.P.Linder in the sense of GPWG 2001) and groups it with the Chloridoideae. Verboom *et al.* (1994) also reported that this genus lacks haustorial synergids, a characteristic of the Danthonioideae, and is therefore excluded from this subfamily. The multinerved glumes and lemmas and several-flowered spikelets of this genus correspond to those in the basal lineages of the Chloridoideae, namely the Pappophoreae ( $x = 9, 10$ ) (Hilu *et al.* 1999; Hilu and Alice 2001).

*Merxmuellera rangei* is the only species of the genus to occur in Namibia, the other occurring mainly in mountainous areas of most of southern Africa, the most species being endemic (Fish 2000). According to Ellis (1982) the leaf anatomy of this species differs from all the other representatives of the genus and that this species occupies an isolated position in the genus and warrants generic status.

These formerly danthonioid species are now placed in the Chloridoideae, but have not been included into any tribe. They represent the most basal members of the Chloridoideae as now delimited (GPWG 2001).

The only genera in southern Africa that have exclusively  $x = 9$  are *Craspedorachis* Benth., *Eleusine* Gaertn. and *Odyssea* Stapf. However, the genera *Craspedorachis* and *Odyssea* are only represented by one chromosome report each (Moffet and Hurcombe 1949; present study). *Craspedorachis* is extremely rare in southern Africa and has a more widespread distribution in tropical Africa and North and South America. *Odyssea* does not have such a restricted distribution in southern Africa and also occurs in tropical Africa. No conclusions regarding the importance of  $x = 9$  can, therefore be made for these genera without a more thorough investigation.

Chromosome numbers based on  $x = 9$  and 10 have been reported for *Eleusine*, though only reports based on  $x = 9$  have been found in southern Africa. Reports of  $x = 10$  have been published by Hiremath and Chennaveeraiah (1982)



(India) [ $2n = 4x = 40$  for *E. compressa* (Forssk.) Asch. & Schweinf. ex. C. Chr. and  $2n = 2x = 20$  for *E. jaegeri* Pilg.]; Hiremath and Salimath (1991) (Tanzania) ( $2n = 2x = 20$  for *E. jaegeri*) and Mysore and Baird (1997) (Kenya and Tanzania) ( $2n = 2x = 20$  for *E. jaegeri*).

Due to the limited nature of reports for  $x = 9$ , it seems that all the Chloridoideae, which are present in southern Africa, have chromosome numbers based on either the paleopolyploid basic chromosome number of  $x = 10$ , or both  $x = 9, 10$  and that no genus is characterized only by  $x = 9$ .

In contrast to Africa, New World genera in the subfamily are characterized by single basic chromosome numbers derived by aneuploidy from either  $x = 5$  or the paleopolyploid number  $x = 10$  (Appendix C). This includes the genus *Hilaria* Kunth (Southern USA to Guatemala) (Clayton and Renvoize 1986), with only chromosome reports based on  $x = 9$  (Ornduff 1967, 1968, 1969; Federov 1969; Moore 1970, 1973; Goldblatt 1981, 1985, 1988; Goldblatt and Johnson 1991). Many New World genera such as *Blepharoneuron* Nash (Moore 1973; Goldblatt 1981; Goldblatt and Johnson 1991, 1994); *Chaboissaea* E. Fourn. (Ornduff 1969; Moore 1970; Goldblatt and Johnson 1996); *Erioneuron* Nash (Ornduff 1968; Moore 1970, 1973; Goldblatt 1981, 1988) and *Munroa* Torr. (Federov 1969; Moore 1973; Goldblatt 1981) and the Australian genus *Triodia* R. Br. (Federov 1969; Moore 1973) have basic chromosome numbers of 8 which might be a further aneuploid reduction from  $x = 9$  and *Blepharidachne* Hack. (Federov 1969; Goldblatt 1981, 1988) has multiples based on 7. The fact that most of these genera, which do not occur in southern Africa or even Africa, consist of derived new basic chromosome numbers are indications of the young age of these genera and implies that they are of more recent radiation in the Chloridoideae.

## 4.4 Conclusions

Chromosome numbers reported in this study confirm the presence of the two main basic chromosome numbers in the southern African Chloridoideae and indicates the high incidence of  $x = 10$ , which is then a paleopolyploid chromosome number. Although the reports of diploids ( $2n = 2x = 10, 12$ ) are few for the Chloridoideae, increased sampling and cytogenetic and molecular studies will render

more evidence that the same phenomenon, as is seen in the closely related Panicoideae, is present in the Chloridoideae. Although the mechanisms might differ, the large affinities between the Chloridoideae and Panicoideae and the occurrence of specimens based on  $x = 5$  indicates that the same phenomenon of polyploidization and subsequent rediploidization has occurred in these subfamilies. The African origin of the Chloridoideae is supported by the presence of taxa with either the paleopolyploid  $x = 10$  or both  $x = 10$  and  $x = 9$ , and by the restriction of deviations from these numbers to other continents where diversification took place later on in the history of the group.

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## CHAPTER 5

# Polyploidy

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**IN PRESS (TAXON):**

R. ROODT AND J.J. SPIES. CHROMOSOME STUDIES IN THE GRASS SUBFAMILY CHLORIDOIDEAE. II. AN ANALYSIS OF POLYPLOIDY IN SOUTHERN AFRICA AND THE WORLD. IN PRESS (TAXON)

Polyploidy is frequent among the grasses. This study indicates that it has a high frequency in the subfamily Chloridoideae, where more than 90% of the studied specimens are polyploids. These levels range from diploid ( $2n = 20$  for  $x = 10$  and  $2n = 18$  for  $x = 9$ ) to 16-ploid ( $2n = 160$ ) for  $x = 10$  in *Ctenium concinnum* Nees and 20-ploid ( $2n = 180$ ) for  $x = 9$  in *Hilaria mutica* Benth. This polyploid trend is seen in southern Africa, as well as worldwide. Analysis indicates that many polyploids are allopolyploids, suggesting extensive hybridization within this group. This is supported by the presence of apomixis in many members of the subfamily. Both polyploidy and apomixis are thought to be important evolutionary mechanisms in the Chloridoideae as well as the closely related Panicoideae, and have a high frequency in southern Africa. Due to the stable ecological, geographical and climatological history of Africa, the continent is ideally suited for the stabilization of hybrid complexes. This stabilization of newly formed hybrids occurs by means of apomixis and polyploidization.

## 5.1 Introduction

Polyploidy is the existence, in genetically related taxa, of chromosome numbers that are multiples of each other, with the polyploid specimens containing more than two genomes per nucleus (Stebbins 1950). This phenomenon occurs in 80% of all angiosperms and is one of the most important cytogenetic mechanisms in plant evolution (Masterson 1994), as it is a prime facilitator of rapid speciation (Hiremath and Salimath 1991). In Poaceae (R.Br.) Barnh., polyploidy has undeniably played an essential role in the evolution of this family, for as early as 1947, Stebbins reported that approximately 70% of the known wild species of grasses were of polyploid origin and by 1985, Stebbins stated that between 80% and 90% of the grass taxa have undergone polyploidy during their evolutionary history. In the South African grass flora this is also reflected, where polyploidy has been reported in more than 80% of the specimens investigated (Spies *et al.* 1996).

The occurrence of polyploidy in the grass subfamily Chloridoideae Kunth ex Beilschm. is especially high. According to Goldblatt (1980), chromosome numbers of  $x \geq 9$  are probably secondary derived, and in this subfamily, where  $x = 9$  and 10 dominate,  $x = 9$  (derived by aneuploidy from 10) and 10 (probably derived by

allopolyploidization from 5) are, therefore polyploid. For the rest of this paper these will be referred to as basic chromosome numbers, even though they essentially are not.

Apomixis includes various developmental pathways by which plants can reproduce asexually via seed (Nogler 1984; Asker and Jerling 1992). It is known from more than 400 flowering plant species and 35 plant families (Nogler 1984; Carman 1997). Despite this taxonomic diversity most apomicts are also polyploids (Asker 1980; Grant 1981; Mogie 1986; Asker and Jerling 1992; Carman 1997; Grossniklaus *et al.* 2001). This was observed as early as 1878 by Strasburger who indicated that apomictic plants often have high chromosome numbers. Rosenberg (1917) and Winge (1917) independently remarked on the connection between high chromosome numbers, hybrid origin and apomixis.

Apomixis has a marked localized distribution with 75% of apomicts belonging to only three families, namely Asteraceae Dumort, Poaceae and Rosaceae L. (Brown and Emery 1958; Richards 1990; Asker and Jerling 1992; Czapik 1996, 2000; Vielle Calzada *et al.* 1996). Apomixis is unusually common among grasses of South Africa (Brown and Emery 1957; Goldblatt 1978) compared to other herbaceous families. According to Brown and Emery (1958), apomixis is especially common in the subfamily Panicoideae Link (tribes Paniceae R.Br., Andropogoneae Dumort.) and Chloridoideae (tribes Chlorideae Kunth, Eragrostae Benth., Pappophoreae Kunth), compared to other subfamilies in the grasses. These two subfamilies together comprise 52% of the known reports of apomixis in the grass family (Czapik 2000). Apomixis is also frequent in the Pooideae Benth. (35% of known reports) (Czapik 2000) but this subfamily has a very low frequency in southern Africa (15%) where the Chloridoideae (24%), Arundinoideae (24%) [which are now three separate subfamilies, namely Aristidoideae Caro, Arundinoideae and Danthonioideae Barker & H.P.Linder (GPWG 2001)] and Panicoideae (35%) comprise the largest percentage of grasses (Gibbs Russell 1986). Brown and Emery (1958) proposed that the same apomictic genes are widespread amongst these two subfamilies (tribes according to the classification they used), and that many of the sexual species contain one or a few genes necessary for an effective balanced apomictic system. These sexual species would be carriers of one or more apomictic genes and subsequent hybridizations between these carrier taxa would

facilitate apomixis (Brown 1958). Carman (1997) felt that apomixis is a multigene phenomenon and that the parthenogenetic development of the embryo is the result of duplicated female development genes that are asynchronously expressed. The required increase in the number of genes can be achieved by polyploidy. Even if the polyploidization event is ancient (such as in the Chloridoideae and Panicoideae) and paleopolyploids exist that are diploidized, the chromatin of the multiple genomes are still present (Carman 1997). Various authors support this multigene apomictic regulation model (for example Blakey *et al.* 1997; Sokolov *et al.* 1998; Blakey *et al.* 2001). This is in line with initial studies of apomixis by Gustafsson (1947a, b), who was of the opinion that apomixis results from the genetic interactions following polyploidy in taxa that had a tendency towards apomixis (or then the "genes necessary for an effective balanced apomictic system" proposed by Brown and Emery 1958). This tendency will be the intergenomic heterozygosity for rates and timing of female development that exists within cosmopolitan families, such as the grass family (Carman 1997).

It is the aim of this study to investigate the incidence and type of polyploidy in the Chloridoideae in southern Africa and to indicate that hybridization, polyploidization and apomixis are correlated in the subfamily and are especially high in the region where the group is proposed to have originated.

## 5.2 Results

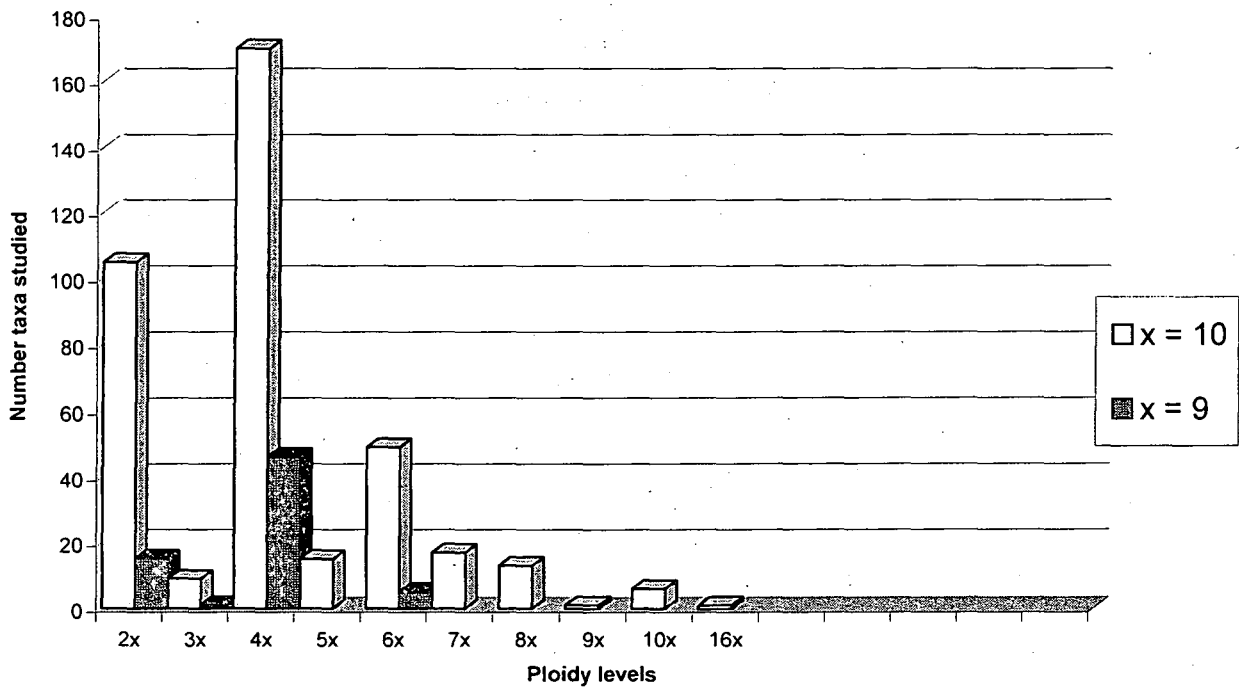
The past century has seen an increase in the number of studies dealing with chromosome numbers in the grasses. The following studies have increased our knowledge of the southern African chloridoid grasses: Avdulov (1931); Hunter (1934); Nielson (1939); Moffet (1944); Hurcombe (1946, 1947); Moffett and Hurcombe (1949); Pienaar (1953); De Wet (1954a, b, 1958, 1960); De Wet and Anderson (1956); Reeder and Singh (1968); Nordenstam (1969); Vorster and Liebenberg (1977); Jones *et al.* (1978); Nordenstam (1982); Spies (1982); Davidse *et al.* (1986); Spies and Du Plessis (1986a, b, 1987); Spies and Jonker (1987); Du Plessis and Spies (1988); Hoshino and Davidse (1988); Spies and Gibbs Russell (1988); Spies and Voges (1988); Spies *et al.* (1991); Strydom and Spies (1994a); present study (see appendix A, B).

These studies report on chromosome numbers for approximately two thirds (34) of the genera present in the subfamily (52) in southern Africa. The genera that have not been studied are mostly monotypic, some of which are exceedingly rare, for example *Brachyachne* (Benth.) Stapf, *Brachychnoa* S.M.Phillips and *Schoenefeldia* Kunth (Gibbs Russell *et al.* 1990).

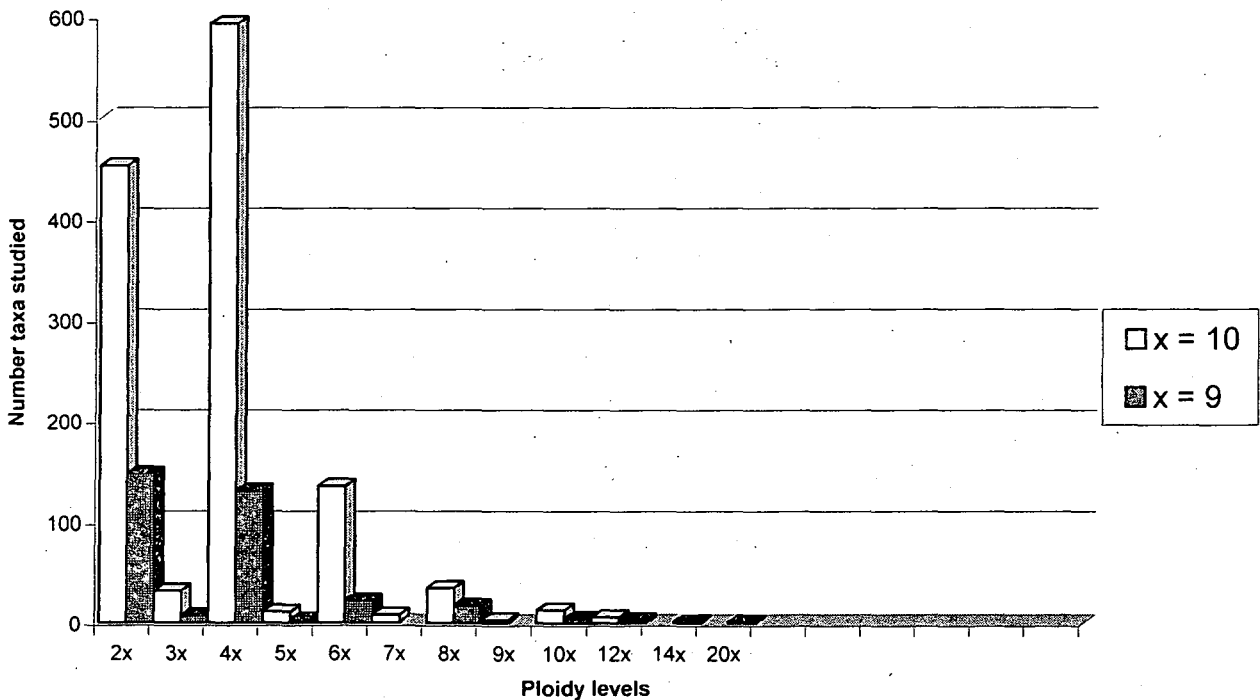
The results obtained in this study, together with the chromosome reports for the southern African Chloridoideae previously published, were used to compile a table of chromosome numbers in the southern African Chloridoideae (Appendix B). A table was also compiled of the known chromosome numbers of Chloridoideae from the rest of the world (Appendix C). These tables were used to calculate frequencies of polyploidy in the region (Figure 5.1), as well as in the rest of the world (Figure 5.2). To make interpretation easier, only  $x = 9$  and  $10$  are indicated on these graphs.

In southern Africa, 78% of the species with  $x = 9$  are polyploids and 71% of the species based on  $x = 10$  are polyploids. Only a single species from Zimbabwe, *Dactyloctenium giganteum* Fisch. & Schweick., has ever been reported as truly diploid (based on  $x = 5$ ) ( $2n = 2x = 10$ ) with five bivalents observed during metaphase I (Davidse *et al.* 1986). In the rest of the world only two diploid species, with  $x = 5$  ( $2n = 2x = 10$ ), *Muhlenbergia andina* A.Hitch. (Pohl 1969) and *Oropetium thomaeum* Trin. (Swami 1963) have been reported. Other basic chromosome numbers such as 6 (Figure 5.3A), 7 and 8 (aneuploid derivatives of the original basic chromosome number of  $x = 5$  or the secondarily derived basic number of  $x = 9$ ) contribute only 11% of the total number of chromosome reports for the group worldwide, and could indicate that these chromosome numbers either developed recently in the history of the group and have not become established in large numbers or, are specialization in groups in which  $x = 10$  or  $9$  was already present.

The vast majority of species possess multiples of 10 (Figure 5.1). When the two main basic chromosome numbers,  $x = 10$  (Figure 5.3B–F) and  $9$  (Figure 5.3G–J) are compared, it can be seen that the basic number of 10 forms a much larger polyploid complex, with the most frequent polyploid level being tetraploid. The same general trend is seen worldwide (Figure 5.2). Both in southern Africa and worldwide

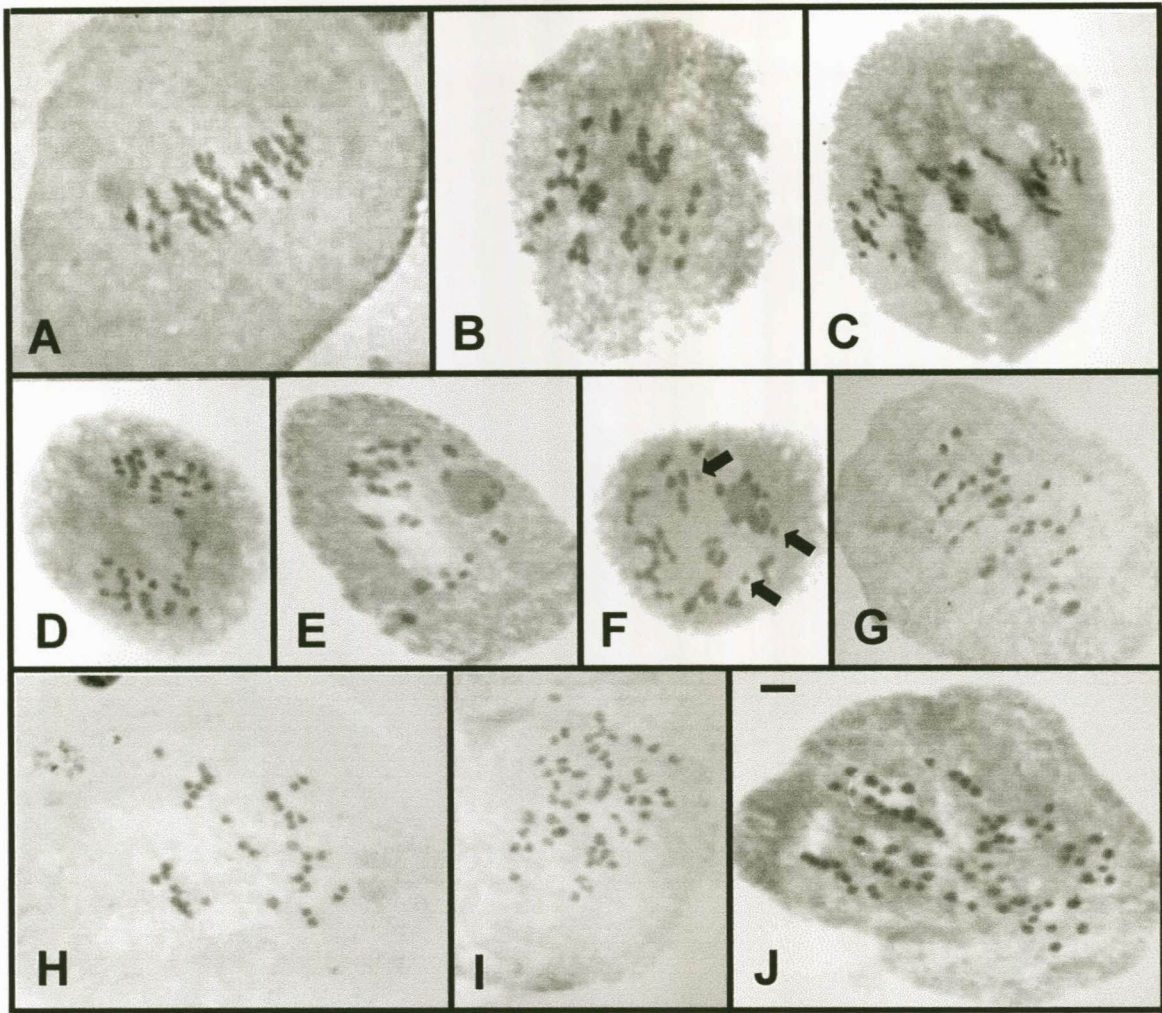


**Figure 5.1.** Ploidy levels present in southern Africa for basic chromosome numbers  $x = 9, 10$ .



**Figure 5.2.** Ploidy levels present worldwide for basic chromosome numbers  $x = 9, 10$ .





**Figure 5.3.** Photomicrographs of meiotic chromosomes in some genera of the subfamily Chloridoideae. **A.** *Centropodia glauca*, Spies 5706,  $2n = 48$ , early anaphase I. **B.** *Dactyloctenium aegyptium*, Spies 2403,  $2n = 40$ , diakinesis with  $20_{II}$ . **C.** *Enneapogon cenchroides*, Spies 2709,  $2n = 60$ , early metaphase I. **D.** *Eragrostis capensis*, Spies 3498,  $2n = 40$ , anaphase I with one lagging chromosome pair. **E.** *E. inamoena*, Spies 2392,  $2n = 40$ , diakinesis with  $20_{II}$ . **F.** *E. planiculmis*, Spies 1116,  $2n = 60$ , diakinesis with  $30_{II}$ . B chromosomes are indicated. **G.** *Harpochloa falx*, Spies 4695,  $2n = 36$ , early anaphase I. **H.** *H. falx*, Spies 5078,  $2n = 36$ , late anaphase I. **I.** *H. falx*, Spies 5128,  $2n = 54$ , late anaphase I. **J.** *H. falx*, Spies 6955,  $2n = 36$ , anaphase I, two cells, each with an 18-18 segregation. Scale bar: A, D, E = 5  $\mu\text{m}$ ; F, J = 6  $\mu\text{m}$ ; B, C, G-I = 7  $\mu\text{m}$ .

$x = 10$  has the same type of distribution with a large number of polyploids levels present and tetraploid levels occurring in the highest frequency.

## 5.3 Discussion

De Wet (1954a) and Stebbins (1956) were both of the opinion that the subfamily Arundinoideae represents the nearest living representative of the ancestral grass line that moved into the savannas from the forests. According to Clayton (1974), Arundinoideae apparently evolved into two directions namely the Chloridoideae and the Panicoideae. The origin of the Chloridoideae from arundinoid ancestors has been advocated by several researchers (for example Stebbins 1956; Tateoka 1957; Clayton 1981; Renvoize 1981; Phillips 1982; Campbell 1985; Watson *et al.* 1985; Clayton and Renvoize 1986; Conert 1987; Jacobs 1987; Kellogg and Campbell 1987; Davis and Soreng 1993; Hilu and Esen 1993; Liang and Hilu 1996; Soreng and Davis 1998; Hilu *et al.* 1999; Hsiao *et al.* 1999; Hilu and Alice 2001).

From the Arundinoideae with its basic chromosome number of 6, a possible aneuploid reduction could have lead to five in the Chloridoideae and Panicoideae, which was probably largely unstable and therefore polyploidization lead to the secondary basic chromosome number of ten and subsequent aneuploid reduction to nine which are now the most frequent basic chromosome numbers in these subfamilies (but see study by Spangler *et al.* 1999 based on *ndhF* sequences proposing that  $x = 5$  represents a reduction in the Andropogoneae and not the ancestral basic chromosome number). This is corroborated by the studies of Barker *et al.* (1995, 1999) and Hilu *et al.* (1999) in which *Danthonia* DC. and *Centropodia* Rchb. (previously arundinoid genera but now Danthonioideae and Chloridoideae respectively) ( $x = 6$ ) are most basal to the Chloridoideae.

Mehra *et al.* (1968) felt that  $x = 10$  became stabilized early in the history of the Panicoideae, explaining the prevalence of multiples of 10, whereas  $x = 9$  occurs in the more advanced tribes such as the Paniceae. This is probably also the case with the Chloridoideae where only as small number of primary diploids ( $2n = 2x = 10$ ) have been reported (Swami 1963; Pohl 1969; Davidse *et al.* 1986) and  $x = 10$  is more prevalent than  $x = 9$ . Hilu and Alice (2001) felt that  $x = 10$  is a plesiomorphic character in the Chloridoideae and that  $x = 9$  along with characters such as the evolution of the spicate inflorescence, a reduction in fertile florets and fewer-nerved

lemmas became established soon after the appearance of early chloridooids because these characters are present in the more basal clades in their study based on the *matK* gene. These basal clades are represented by the tribe Pappophoreae Kunth, subtribe Uniolinae Clayton and the genus *Eragrostis* Wolf. Clayton and Renvoize (1986) placed the Uniolinae as basal in their circumscription of the Chloridoideae and consider them the link between the Chloridoideae and Arundinoideae. In the Uniolinae,  $x = 10$  is frequent, but in the genus *Uniola* L.,  $x = 6$  also occurs. Van den Borre and Watson (1997) place the Pappophoreae as near basal in their phylogeny based on morphology. Both  $x = 9$  and  $10$  occur in this tribe and indicate that the polyploidization event from  $x = 5$  to  $10$  and subsequent aneuploid reduction to  $x = 9$  occurred early in the history of the group.

Polyploid complexes, as those found in this study within and among species, can be classified into five types according to the age of the complex

- An initial complex, which has the most recent origin and diploid specimens are most common.
- The young complex in which the diploids are far less common than polyploids and tend to be endemics.
- The mature complex where nearly all diploid specimens have become extinct and new cycles of polyploidy have arisen based on hybridization and/or chromosome doubling of specimens which behave like diploids, but have basic chromosome numbers of ancient polyploid origin. This diploid-like behavior is possibly due to chromosomal reorganization and gene silencing that can be so extensive that the genome is not structured as a polyploid any longer (Soltis and Soltis 1993, 1999).
- The declining complex where all the diploid ancestors have died out, but the basic chromosome number for the genus can still be determined by comparison with closely related taxa.
- The relictual complex comprising mainly of monotypic genera of very high chromosome number, with no close existing relatives. It is impossible to determine the basic chromosome number of the genus

because no remaining closely related taxa exist to make comparisons to (Stebbins 1956, 1971).

The Chloridoideae with  $x = 10$  represents a mature polyploid complex in which most original diploids (based on  $x = 5$ ) have become extinct, and as a result of hybridization and polyploidization, basic chromosome number of ancient polyploid origin ( $x = 10$ ) are now found in most genera, whereas  $x = 9$  represents a much younger complex with a relatively large frequency of diploids still apparent, compared to the other polyploid levels present.

To determine what type of polyploidy is present in the genera studied and whether hybridization occurs the meiotic configurations in tetraploid southern African specimens were analyzed with the model of Kimber and Alonso (1981). Kimber and Alonso (1981) formulated different tetraploid models, 4:0, 3:1, 2:1:1 and 2:2, based on the degree of homology between the genomes present in the tetraploid. Chromosome configurations are used to determine the relative affinity between the genomes and this is expressed by an  $x$ -value. Expected and observed chromosome configurations are compared and the sums of squares between these values for each specimen are calculated (Kimber and Alonso 1981). The model with the lowest sum of squares is considered to be the model best suited for that particular specimen. In the case of a 3:1 model, an  $x$ -value of 0.5 indicates that the last genome corresponds to the first three and that the specimen has a genomic constitution of for example AAAA (autopolyploidy). With this same model, an  $x$ -value of 1 indicates no homology between the last genome and the first three and the specimen will have a genomic constitution of for example AAAB (allopolyploidy). The 2:2 model with an  $x$ -value of 1 would indicate a genomic composition of AABB. Values ranging from 0.5 to 1 indicate varying degrees of homoeology and would indicate segmental allopolyploidy (Kimber and Alonso 1981).

The results of these analyses are presented in Table 5.1. The 2:2 model fit all the specimens best. These results indicate that the studied taxa are either segmental allopolyploids, which have diversified so much as to almost be a true allopolyploid ( $x$  value of 0.99), or allopolyploids ( $x$  value of 1). Even though the chromosomes studied are small, meiotic configurations could still be discerned and chiasma frequencies calculated. Sybenga (1994) stated that pairing in polyploids

**Table 5.1.** Genomic relationships in the tetraploid specimens analyzed according to the models of Kimber and Alonso (1981). Values indicated represent the sums of squares calculated for the four possible tetraploid models. The x-values are indicated in parentheses. The model best suited for each specimen is indicated in bold.

Specimen	2n	Voucher number	Chiasma frequency	Models			
				4:0	3:1	2:2	2:1:1
<b>Cynodonteae</b>							
<i>Cynodon dactylon</i>	36	Spies 2549	0.5	45.34	57.52 (0.95)	<b>30.39</b> (1)	45.42 (0.5)
<i>Cynodon dactylon</i>	36	Spies 2966	0.89	18.34	18.88 (0)	<b>0.074</b> (0.99)	0.48 (0.994)
<i>Harpochloa falx</i>	36	Spies 5065	0.50	45.34	45.71 (0.5)	<b>30.38</b> (0.99)	45.42 (0.5)
<i>Harpochloa falx</i>	36	Spies 5113	0.53	39.04	53.60 (0)	<b>25.72</b> (0.99)	39.13 (0.5)
<i>Harpochloa falx</i>	36	Spies 6955	0.50	44.54	44.91 (0.5)	<b>29.77</b> (1)	44.62 (0.5)
<i>Harpochloa falx</i>	36	Spies 4695	0.50	45.34	45.71 (0.5)	<b>30.38</b> (0.99)	45.42 (0.5)
<i>Harpochloa falx</i>	36	Spies 4729	0.50	45.34	50.44 (0.8)	<b>30.38</b> (0.99)	45.42 (0.5)
<i>Harpochloa falx</i>	36	Spies 4691	0.53	38.65	52.61 (0.99)	<b>24.21</b> (0.99)	38.74 (0.5)
<b>Eragrostideae</b>							
<i>Bewsia biflora</i>	40	Spies 1531	0.88	22.02	22.76 (0)	<b>0.2X10<sup>-5</sup></b> (0.99)	0.77 (0.995)
<i>Cladoraphis spinosa</i>	40	Spies 4885	0.51	54.40	73.46 (0)	<b>36.03</b> (0.99)	54.50 (0.50)
<i>Eleusine indica</i> subsp. <i>africana</i>	36	Spies 2783	0.71	15.90	18.47 (0.85)	<b>3.65</b> (0.99)	11.45 (0.95)
<i>Eragrostis capensis</i>	40	Spies 3498	0.59	32.80	46.99 (0)	<b>20.12</b> (0.99)	32.91 (0.58)

<i>Eragrostis capensis</i>	40	HdP 110	0.58	36.38	37.22 (0.58)	<b>19.58</b> <b>(0.99)</b>	36.35 (0.75)
<i>Eragrostis capensis</i>	40	Spies 3483	0.71	13.32	13.86 (0.5)	<b>5.17</b> <b>(1)</b>	10.97 (0.9)
<i>Eragrostis capensis</i>	40	Spies 5069	0.51	54.40	54.86 (0.5)	<b>36.04</b> <b>(1)</b>	54.50 (0.5)
<i>Eragrostis capensis</i>	40	Spies 1595	0.55	41.98	58.35 (0)	<b>24.62</b> <b>(1)</b>	42.08 (0.61)
<i>Eragrostis echinochloidea</i>	40	Spies 2799	0.80	17.89	20.36 (0.999)	<b>1.010</b> <b>(0.99)</b>	5.20 (0.999)
<i>Eragrostis inamoena</i>	40	Spies 2392	0.58	35.47	49.93 (0.999)	<b>19.04</b> <b>(1)</b>	35.44 (0.75)
<i>Eragrostis racemosa</i>	40	Spies 3279	0.50	55.98	60.62 (0.76)	<b>37.52</b> <b>(1)</b>	56.08 (0.5)
<i>Eragrostis racemosa</i>	40	Spies 4743	0.50	45.34	45.71 (0.5)	<b>30.38</b> <b>(0.99)</b>	45.42 (0.5)
<i>Eragrostis racemosa</i>	40	Spies 5066	0.50	55.98	75.3 (0)	<b>37.52</b> <b>(1)</b>	56.08 (0.5)
<i>Eragrostis superba</i>	40	Spies 3326	0.52	50.86	51.33 (0.5)	<b>33.21</b> <b>(0.99)</b>	0.96 (0.5)
<i>Eragrostis superba</i>	40	HdP 136	0.54	44.18	61.16 (0)	<b>27.25</b> <b>(0.89)</b>	44.29 (0.5)
<i>Fingerhuthia africana</i>	40	Spies 2947	0.52	50.27	50.75 (0.5)	<b>32.19</b> <b>(0.99)</b>	50.37 (0.5)
<i>Sporobolus africanus</i>	36	Spies 2369	0.58	27.69	28.17 (0.5)	<b>14.57</b> <b>(0.99)</b>	27.60 (0.76)
<b>Pappophoreae</b>							
<i>Enneapogon cenchroides</i>	40	Spies 3288	0.64	18.36	18.89 (0.5)	<b>8.41</b> <b>(1)</b>	17.60 (0.85)
<i>Enneapogon sp.</i>	40	Spies 5532	0.62	23.14	23.65 (0.5)	<b>10.89</b> <b>(1)</b>	22.60 (0.83)
<i>Schmidtia pappophoroides</i>	36	HdP 186	0.50	45.34	48.30 (0.73)	<b>30.38</b> <b>(0.99)</b>	45.42 (0.5)
<i>Schmidtia sp.</i>	36	Spies 5536	0.57	29.93	42.04 (0)	<b>16.29</b> <b>(0.99)</b>	29.93 (0.73)



could be affected by factors that may increase or decrease the amount of multivalent formation, and that these are independent of the chromosome affinities. These factors could include the position and number of pairing initiation points and their interference with chiasmata formation. Low multivalent frequencies (as observed in this study) are, therefore, not necessarily an indication of limited pairing because even in natural autopolyploids quadrivalent formation may be rare (Sybenga 1992, 1994, 1996). This has been observed in many species in which autopolyploids are known to exist such as *Triticum* L. (Yen and Kimber 1990) and rye (Wang and Berdahl 1990; Santos *et al.* 1995). The occurrence of a few multivalents in some specimens in this study furthermore indicates that *Ph* like genes are probably not present. In wheat it was found that the *Ph1* gene suppresses homoeologous chromosome pairing and bivalent formation will take place (Riley and Chapman 1958; Sears 1976). The mechanism of this phenomenon is controlled by a series of genes on the homoeologous group 5 chromosomes. These genes suppress or, in some cases, promote homoeologous chromosome pairing. A mutant (*Ph*) of the gene allows homoeologous pairing and is located on the long arm of chromosome 5 of the B genome (Wall *et al.* 1971a, b). The dominant allele suppresses homoeologous pairing and multivalent formation and leads to bivalent formation (Riley *et al.* 1961).

The cytogenetic results could indicate the existence of mostly segmental allopolyploid and allopolyploid tetraploid specimens (secondary derived). The cytogenetic results provide provisional evidence for the existence of large-scale hybridization in the southern African members of the subfamily, in a family in which hybridization is known to have a high occurrence. The abundance of naturally occurring interspecific hybrids in Chloridoideae (and Poaceae in general) seems to be related to the occurrence of different species growing together in dense stands, the production of vast amounts of wind-borne pollen and the existence of self-incompatibility systems which promote crosspollination (Hilu 1993).

According to Vorster (1974), the stable ecological, geographical and climatological history of the African continent makes it very suitable for the stabilization of hybrid complexes. Studies by Van Zinderen-Bakker (1964) on the fossil pollen in southern Africa, indicates fluctuations in the rainfall, dominated by various dry and wet rainfall cycles. This leads to increases or decreases in the

geographical distribution of plant populations. During dry seasons populations may break up into various smaller sub-populations, which might become isolated from one another. If the dry season persists these populations may become karyotypically distinct from one another, by evolutionary development (Van Zinderen-Bakker 1964). Wet seasons could later lead to an increase in the distribution of these populations and hybridization between the separate populations. Depending on the divergence between the species, the hybrids may or may not be sterile with varying degrees of abnormality during meiosis. This could lead to subsequent polyploidization (Van Zinderen-Bakker 1964). This scenario is in concordance with Stebbins' (1985) secondary contact hypothesis characterized by cycles of polyploidy.

The survival and successful establishment of many newly formed hybrids are associated with their perenniality and effective means of vegetative or asexual propagation. Perenniality implies that a plant does not need to be immediately fit, and provides a better chance for polyploids to recover from sterility problems which are associated with chromosome pairing and restore nucleocytoplasmic stability (Hilu 1993). The high frequency of polyploidy in the grasses has provided a means of fixing and spreading hybrid combinations (Stebbins 1956; McWilliam 1964; Knox 1967; De Wet and Stalker 1974; Asker 1980; Rieseberg 1997; Ellstrand and Schierenbeck 2000).

An effective means of asexual propagation to successfully establish hybrids is found in the form of apomixis. Apomixis, asexual reproduction via seed (Nogler 1984; Asker and Jerling 1992), is almost consistently associated with polyploidy in many plant families. There is no evidence that polyploidy promotes apomixis, but in association with hybridization it tends to strengthen the action of genes conditioning apomixis, which are not exhibited in a diploid background (Stebbins 1950; Bell 1982; Michod 1995). Kellogg (1990) found that in the grass family, apomictic groups are hybridizing groups. Apomixis allows the hybrids, which may only be partially fertile or sterile, to survive and increases the generation time for each genotype.

In the Chloridoideae apomixis has been found in *Bouteloua* Lag. (Harlan 1949; Freter and Brown 1955; Mohammed and Gould 1966; Bierzychudek 1985); *Chloris* Sw. (Brown and Emery 1958; Hutton 1961); *Eragrostis* Wolf (Streetman 1963a, b; Voight 1971; Voight and Bashaw 1972, 1976; Brix 1974; Vorster and



Liebenberg 1984; Voight *et al.* 1992); *Eustachys* Desv. (Strydom and Spies 1994a); *Fingerhuthia* Nees (Brown and Emery 1958); *Harpochloa* Kunth (Strydom and Spies 1994a); *Hilaria* Kunth (Brown and Emery 1958); *Rendlia* Chiov. (Strydom and Spies 1994a) and *Schmidtia* Steud. (Brown and Emery 1958). The higher incidence of apomixis in the Old World for the Chloridoideae (only *Bouteloua* and *Hilaria* are New World genera) (and the closely related Panicoideae) can be explained by the fact that these apomicts are more abundant towards the center of distributions of these groups (Brown 1958). As species spread out from a center of origin the more sexual species will be able to migrate the most rapidly. Apomicts will be more restricted and remain in the original habitats to which they are best adapted (Brown 1958).

## 5.4 Conclusions

Polyploidy occurs in the subfamily Chloridoideae. It is widely known that polyploidy has played an important role in the evolution of the grasses. The nature of the polyploidy present is probably related to hybridization and the large amount of apomicts present in southern Africa is also related to polyploidy. Various factors, such as the ecological, geographical and climatological history of Africa make it ideal for hybridization to occur. This can clearly be seen in the amount and type of polyploidy present in the subfamily Chloridoideae and in the closely related Panicoideae (also characterized by ancient polyploidy) in Africa. Furthermore, there is an unusually large concentration of apomictic genera for these two grass subfamilies in southern Africa, probably resulting from polyploidization. This renders evidence that the subfamily has been present long enough for these phenomena to become established and can be linked to the African origin of this subfamily.

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**CHAPTER 6**

**Nuclear DNA sequencing**

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**SUBMITTED (AMERICAN JOURNAL OF BOTANY):**

R. ROODT AND J.J. SPIES. PHYLOGENETIC RELATIONSHIPS IN THE SOUTHERN AFRICAN CHLORIDOIDEAE (POACEAE) BASED ON NUCLEAR ITS SEQUENCE DATA. SUBMITTED (*AMERICAN JOURNAL OF BOTANY*)

A phylogenetic study was conducted on the Chloridoideae, based on sequences of the internal transcribed spacer regions of the nuclear ribosomal DNA. Emphasis was placed on southern African representatives of the subfamily. Sequence data from 63 species in the subfamily Chloridoideae (Poaceae) and six outgroup taxa from the grass subfamilies Danthonioideae and Panicoideae were phylogenetically analyzed. Phylogenetic analyses of the sequences obtained during the study, together with published *ITS* sequences were used to determine relationships within southern African genera of the subfamily. The monophyly of the subfamily was confirmed, as was the polyphyletic nature of the Eragrostideae and Cynodonteae and the genus *Eragrostis*. Although hybridization and polyploidization are frequent in the subfamily, concerted evolution seems to have homogenized these *ITS* regions, as low levels of polymorphisms were observed. The *ITS* phylogeny renders evidence for the paleopolyploid nature of the basic chromosome numbers in the tribes and that from an initial chromosome number of 5,  $x = 10$  and 9 became quickly established as these occur in the more basal groups.

## 6.1 Introduction

The Chloridoideae is one of 12 subfamilies recognized in Poaceae (R.Br.) Barnhart [Grass Phylogeny Working Group (GPWG) 2001]. This subfamily is seen as anatomically distinct in the grass family and is delimited by spherical, inflated bicellular microhairs, the Kranz syndrome and associated anatomy, distinctive leaf-blade anatomy and the chloridoid embryo (Reeder 1957; Tateoka *et al.* 1959; Hattersley and Watson 1976; Ellis 1987). At the tribal and generic levels, however, the subfamily has been characterized by taxonomic problems. This is because many of the morphological characters used are very variable and lack consistency (Hilu and Alice 2000).

Chloridoideae Kunth ex Beilschm. and Panicoideae Link, both members of the PACCAD clade (GPWG 2001), are seen as the most recent radiation in Poaceae (Hsiao *et al.* 1998; GPWG 2001). Both of these subfamilies have diversified into the tropical and subtropical regions, especially the southern Hemisphere (Renvoize and Clayton 1992) and compose the largest majority of the southern African grass flora (Gibbs Russell 1986).

In this study the phylogeny of mainly southern African species of the subfamily Chloridoideae was investigated. This was done due to the large concentration of Chloridoideae in southern Africa, with 50 genera and 230 species of the 194 genera and 912 species of grasses present in southern Africa, belonging to the Chloridoideae (Fish 2000).

The nuclear ribosomal internal transcribed spacer regions were investigated in this study. These internal transcribed spacer (*ITS*) regions of the 18-26S nuclear ribosomal DNA (nrDNA) have been used extensively to study lower level phylogenetic questions. The region includes three components: the 5.8S subunit, an evolutionarily conserved sequence, and two spacer regions flanking the gene, i.e. *ITS1* and *ITS2*.

Within the family Poaceae, *ITS* sequences have been successfully used to resolve phylogenetic relationships at various levels. This includes studies of various genera such as *Bouteloua* Lag. (Columbus *et al.* 1998, 2000); *Brachypodium* P.Beauv. (Catalan and Olmstead 2000); *Bromus* L. (Ainouche and Bayer 1997; Ainouche *et al.* 1999); *Chloris* Sw. (Alice *et al.* 2000); *Festuca* L. (Torrecilla and Catalan 2002); *Festuca* and *Lolium* L. (Charmet *et al.* 1997; Gaut *et al.* 2000); *Miscanthus* Anderss. (Hodkinson *et al.* 2002a); *Phyllostachys* Sieb. & Zucc. (Hodkinson *et al.* 2000); *Sorghum* Moench. (Sun *et al.* 1994; Dillon *et al.* 2001); *Spartina* Schreb. (Baumel *et al.* 2002); *Sporobolus* (L.) R.Br. (Ortiz-Diaz and Culham 2000) and *Zea* L. (Buckler and Holtsford *et al.* 1996a, b). Studies have also been conducted on a related group of grasses known as the stipoid grasses (Jacobs *et al.* 2000) and on the chloridoid tribe Triodieae Benth. (Mant *et al.* 2000), the panicoid tribe Andropogoneae Dumort. (Hodkinson *et al.* 2002b) and the pooid tribes Aveneae Dumort. (Grebenstein *et al.* 1998) and Triticeae Dumort. (Hsiao *et al.* 1995a). The region has also been used to investigate subfamily relationships in the Arundinoideae Burmeist. (Hsiao *et al.* 1998), Danthonioideae Barker & H.P.Linder (published as tribe Danthonieae Zotov by Barker *et al.* 2000) and the Pooideae Benth. (Hsiao *et al.* 1994, 1995b). The phylogeny of the entire grass family (Hsiao *et al.* 1999) was also investigated using this region.

Ribosomal DNA regions are usually situated in the nucleolar organizing regions (NORs) and may be present on several different chromosomes (Thompson

and Flavell 1988). Ribosomal DNA, however, evolves consistently within a single species and displays limited sequence divergence between copies in a single individual (Arnheim *et al.* 1980). Between species normal levels of sequence divergence are found. These two phenomena are collectively called concerted evolution (Dover 1982). Unequal crossing over (Smith 1976), gene conversion (Arnheim 1983) and biased gene conversion (Hillis *et al.* 1991) are the driving forces behind concerted evolution. Concerted evolution leads to the homogenization of the members of a multigene family. This process overcomes the problems with orthologous and paralogous genes (Zimmer *et al.* 1980; Doyle *et al.* 1992b; Moritz and Hillis 1996). The duplication of genes, followed by divergence, usually leads to greater similarity between some members of a multigene family across species, than within the multigene family of the same species (Doyle *et al.* 1992b). Therefore, the need arises to identify orthologous genes, which are similar due to speciation. Concerted evolution can produce situations in which the genes in a single species are more closely related to one another than any genes from another species. The 18S-25S ribosomal DNA is an example of such a large, but homogeneous, multigene family in most plants (Doyle *et al.* 1992b).

There is, however, a growing consciousness of *ITS* polymorphism in plants. Buckler *et al.* (1997) reviewed evidence for recombination among divergent paralogues as well as the presence of pseudogenes and functional *ITS* regions in several studies. *ITS* region polymorphisms in angiosperms have been attributed to processes such as hybridization, polyploidy, slow concerted evolutionary rates across nrDNA loci on non-homologous chromosomes and agamospermy (Suh *et al.* 1993; Sang *et al.* 1995; Buckler *et al.* 1997; Campbell *et al.* 1997). It might be that concerted evolution among paralogues is not homogenizing the sequences rapidly enough for them to behave as a single orthologue (Sanderson and Doyle 1992).

In taxa that are of a hybrid origin multiple copies (from different parents) may be preserved (Buckler and Holtsford 1996a; Cronn *et al.* 1999; Baumel *et al.* 2001), but this is not always the case. Other processes such as bidirectional concerted evolution (Wendel *et al.* 1995a) and intergenomic introgression (Wendel *et al.* 1995b) has also been found to occur and then hybrids do not show polymorphism or parental additivity.

In the Chloridoideae phenomena such as hybridization, polyploidy (more than 70%) and agamospermy have a high occurrence (Chapter 4, 5). According to Hsiao *et al.* (1999), complete concerted evolution of the *ITS* regions occurs in the grasses, although this was not tested by individual cloning. Divergent paralogues have, however, been found in *Festuca* and *Lolium* (Gaut *et al.* 2000).

In this study *ITS* sequence data was used to determine phylogenetic relationships within South African chloridoid genera. Usually sequencing studies, especially due to the high cost of sequencing, lead to the investigation of only one specimen per species and more often one specimen per genus. It was specifically the aim of this study to investigate more specimens of a southern African origin and more specimens per genus and in some instances per species. We focused on southern African genera and species in the subfamily due to the large concentration of genera and species in southern Africa.

## 6.2 Results

During this study the entire *ITS1*, 5.8S and *ITS2* regions were amplified with the four primers used. The assembled sequences of the entire region of the 69 specimens consist of 668 characters (Appendix D), 810 with indel information included (Appendix E, F). *ITS1* had a consensus length of 249 bp, 5.8S was 163bp in length and *ITS2*, 256 bp in length. For the entire region 430 characters (53%) are informative when gaps are considered as character states. These sites are distributed mostly in *ITS1* (36.3%) and *ITS2* (38.6%). The 5.8S region had 5.8% informative sites and indels contributed 19.3% of the total number of informative sites. The percentage guanine (G) and cytosine (C) in the 69 specimens ranged from 49.3% [*Sporobolus fimbriatus* (Trin.) Nees, Venter 9328) to 70% (*Centropodia glauca* (Nees) Cope, AF019861]. The average GC content for the entire region was 57.3%. The average GC contents for *ITS1*, 5.8S and *ITS2* are 56.9%, 55.2% and 59.2% respectively.

No clear evidence of multiple rDNA repeat types was found. Each PCR product was resolved as a single band on 3% agarose gels and low levels of nucleotide polymorphisms, that could indicate multiple repeat types, were observed. Direct sequencing did not result in many ambiguities, suggesting that within an

individual there is little variation. Although intragenomic polymorphisms were observed they were rare overall. For this reason cloning was not done.

Phylogenetic analysis using heuristic search options yielded 1936 equally parsimonious cladograms with a length of 2085, CI of 0.39 excluding uninformative characters and RI of 0.72 excluding uninformative characters. A strict consensus cladogram was used to summarize the data (Figure 6.1). Bootstrap (BS), jackknife (JK) and decay values (DV) are indicated on the cladogram. The monophyly of the Chloridoideae is strongly supported. Two large clades are found within the Chloridoideae although these are rather weakly supported with support values below or in the region of 50%. Most of the genera are monophyletic with only *Eragrostis* Wolf being polyphyletic.

## 6.3 Discussion

The length of the *ITS* region is in concordance with those found by Baldwin *et al.* (1995). These authors found *ITS1* to vary from 187-298 bp in angiosperms and *ITS2* from 187-252 bp. These two regions evolve more rapidly than the coding regions 18S, 5.8S and 26S and this can be seen from the homogeneity across the 5.8S region in the different species. The uniform size of the 5.8S subunit is one of two sizes reported for this region that is highly conserved in angiosperms (163-164 bp) (Baldwin 1993).

The high percentage of informative sites found in this study is in concordance with similar values found in the grasses: in the entire subfamily (53%, Hsiao *et al.* 1999); subfamily Arundinoideae (42%, Hsiao *et al.* 1998); and *Bouteloua* and related genera in the Chloridoideae (46%, Columbus *et al.* 1998). This trend has also been seen in other taxonomic groupings such as the families Acanthaceae Juss. (44%, McDade *et al.* 2000) and Vitaceae Juss. (64%, Rosetto *et al.* 2002); Apiaceae Lindl. subfamily Apioideae Seem. (55%, Downie and Katz-Downie 1996; 60%, Downie *et al.* 1998) and Orchidaceae Juss. subtribe Pleurothallidinae Lindl. ex G. Don (52%, Pridgeon *et al.* 2001).

The GC% in the *ITS* areas of angiosperm genomes is often in the region of 50% or more (Baldwin *et al.* 1995). The GC contents of the *ITS* region of most

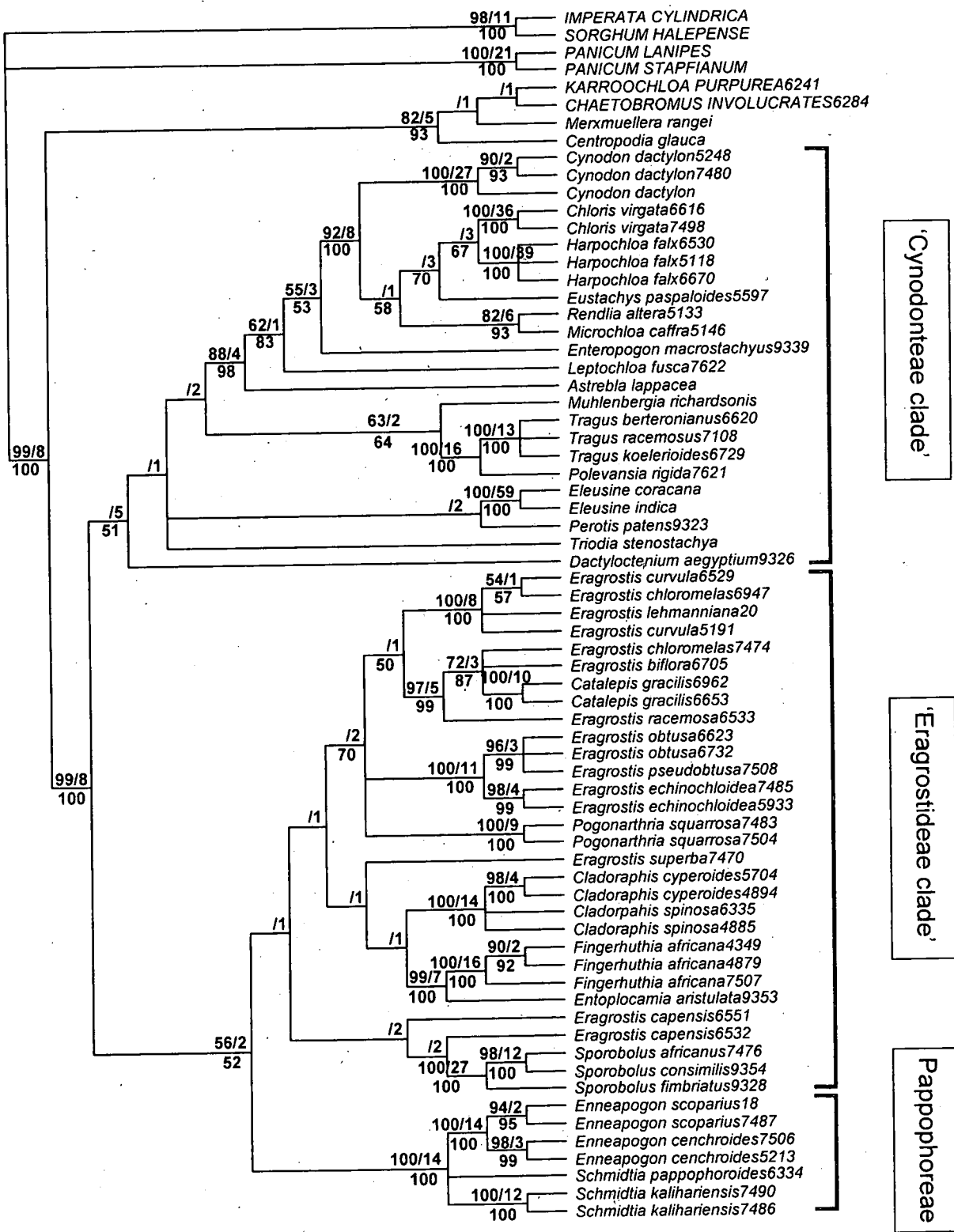


Figure 6.1. Strict consensus cladogram based on ITS sequencing data constructed from 1936 equally parsimonious cladograms with a length of 2085, CI of 0.39 and RI of 0.72 (both excluding uninformative characters). Bootstrap and decay values (BS/DV) are indicated on branches. Jackknife values are indicated below the branches.



grasses is higher than 50%, up to as high as 77% (Takaiwa *et al.* 1985). The average of 57.1% found in this study is similar to that of other grass subfamilies (Hsiao *et al.* 1994, 1995a, b, 1998, 1999). Salinas *et al.* (1988) showed by reassociation kinetics of single-stranded DNA that grasses growing in arid regions have on average a higher GC content than plants from temperate areas. This suggests an adaptive significance of the higher GC content. Furthermore, the high GC content observed in most of the specimens studied could account for the difficulties experienced in sequencing the 5.8S and *ITS2* region. These high GC levels cause stronger template secondary structures, which may confound the sequencing reactions (Steane *et al.* 1999). For this reason the addition of DMSO is essential.

In the Chloridoideae, where polyploidization and hybridization are frequent, additivity of parental types (Soltis, P.S. *et al.* 1990) and polymorphisms is expected. Interlocus concerted evolution may, however, homogenize repeats from different arrays and then effectively remove one parent from the hybrid (Wendel *et al.* 1995a; Ainouche and Bayer 1997). Interlocus concerted evolution usually needs many sexual generations to reach completion (Wendel *et al.* 1995a; Brochman *et al.* 1996) and the lack of additivity might be accounted for by ancient polyploidy and also ancient hybridization, with complete concerted evolution in high polyploids (McDade 1995; Vargas *et al.* 1999). Where concerted evolution is present, direct evidence of hybrid speciation is not present any more but nrDNA may still be used when compared with other data (Hughes *et al.* 2002). The loss of an rDNA locus is another reason for a lack of non-additive profiles especially in hybrids of recent origin. This may be due to the terminal location of nucleolar organizer regions (NORs) where they are surrounded by heterochromatin and could be subject to gene loss (Soltis, P.S. *et al.* 1990). Concerted evolution among rDNA loci may be more common than previously believed, if taken into consideration that diploid species suspected of being paleopolyploids have *ITS* sequence homogeneity (Baldwin 1992; Baum *et al.* 1994; Wendel *et al.* 1995a). Furthermore, according to Soltis and Soltis (1993) and Campbell *et al.* (1997), hybridization may not have a large impact on polymorphisms and the amount of polymorphism depends on the sequence divergence of parents. In the Chloridoideae, any of the above could have

played a role, but ancient polyploidy (Chapter 4, 5), which has been suggested for the group, could be one of the main contributing factors.

The Chloridoideae was found to be a well-supported monophyletic lineage (BS = 99%, JK = 100%, DV = 8) (Figure 6.1). This has long been accepted and illustrated in various previous molecular studies such as Nadot *et al.* (1994); Barker *et al.* (1995); Clark *et al.* (1995); Duvall and Morton (1996); Liang and Hilu (1996); Van den Borre and Watson (1997); Soreng and Davis (1998); Hilu *et al.* (1999); Hsiao *et al.* (1999); Hilu and Alice (2000, 2001); Matthews *et al.* (2000); Van den Borre and Watson (2000). Clayton and Renvoize (1986) believe the Chloridoideae to be monophyletic based on the adoption of the C<sub>4</sub> photosynthetic pathway. Hilu and Esen (1993) and Hilu (2000) also supported the monophyly of the subfamily based on prolamin and immunological studies.

*Centropodia glauca* and *Merxmuellera rangei* (Pilg.) Conert are not yet placed in any of the tribes of the Chloridoideae. These two taxa were previously placed in Danthonioideae but consistently group with the Chloridoideae as the most basal lineages (GPWG 2001). In this analysis they group with the Danthonioideae with strong support (BS = 82%, JK = 93%, DV = 5). This grouping with a group from which they were removed, might be due to the fact that their immediate sister group, Aristidoideae Caro (GPWG 2001), was not included in the study.

The current study distinguishes two main groupings in the Chloridoideae corresponding largely to the two main tribes, Eragrostideae Stapf and Cynodonteae Dumort. These are, however, only weakly supported (BS = < 50%, 56%; JK = 51%, 52%; DV = 5, 2) and are clearly not monophyletic. In the studies by Van den Borre and Watson (1997), Hilu *et al.* (1999) and Hilu and Alice (2000, 2001) based on molecular and morphological and anatomical data, the main tribes, Cynodonteae and Eragrostideae, were also not monophyletic. The difference between these tribes are based on inflorescences, with the Cynodonteae defined by a one-sided spike-like inflorescence that carries spikelets with one fertile floret subtending one or more sterile ones. The Eragrostideae are characterized by several flowered spikelets that are arranged in a panicle (Clayton and Renvoize 1986). Genera like *Acrachne* Chiov., *Dactyloctenium* Willd., *Eleusine* Gaertn. and *Leptochloa* P.Beauv. are difficult to place because they combine the spiciform inflorescence as well as the several flowered spikelets (Hilu and Wright 1982). This can be seen from the

cladogram with *Dactyloctenium*, *Eleusine* and *Leptochloa* being classified as Eragrostideae, but grouping within the 'Cynodonteae clade'. Other studies that could not distinguish between the two main tribes include the immunological and prolamin studies of Hilu and Esen (1993), leaf blade anatomical studies of Ellis (1987) and studies by Phillips (1982) and Campbell (1985). According to Hilu and Alice (2000), too much emphasis is placed on separating Eragrostideae from Cynodonteae on number of florets, due to the variability in morphological characters and the lack in consistency at tribal and even generic levels. Campbell (1985) and Watson and Dallwitz (1992 onwards) proposed the merging of the chloridoideae tribes.

The monophyly of Pappophoreae Kunth is strongly supported (BS = 100%, JK = 100%, DV = 12) and it forms the most basal lineage grouping with the Eragrostideae. The Pappophoreae is an isolated tribe (Clayton and Renvoize 1986) and is characterized by unusual elongated bulbous-tipped microhairs (Jacques Felix 1962; Reeder 1965; Renvoize 1985). Van den Borre and Watson (1997) also found Pappophoreae near basal in their study. In their analysis the Pappophoreae group with the outgroup genera indicating their isolated position (see also Watson *et al.* 1985).

The largest genus in the Chloridoideae, *Eragrostis*, was polyphyletic in this study. Based on the *matK* chloroplast study of Hilu and Alice (2000, 2001) and the morphological and anatomical study of Van den Borre and Watson (1997, 2000), *Eragrostis* was also polyphyletic. Clayton and Renvoize (1986) and Kellogg and Campbell (1987) have suggested that *Eragrostis* is paraphyletic. *Sporobolus* forms a monophyletic clade in this study (BS = 100%, JK = 100%, DV = 27) and groups with *Eragrostis capensis* (Thunb.) Trin., although this grouping has very weak support (BS < 50%, JK < 50%, DV = 2). *Sporobolus* is closely related to *Eragrostis*. This was proposed by Phillips (1982) and studies by Hilu and Wright (1982), Duvall *et al.* (1994) and Hilu and Alice (2000, 2001) support this. Jacobs (1987) was of the opinion that *Sporobolus* and *Eragrostis* should not be separate genera and should be grouped together as *Eragrostis sensu lato* and reclassified.

The genus *Muhlenbergia* Schreb. falls in the 'Cynodonteae clade' even though this is a member of the Eragrostideae tribe. Clayton and Renvoize (1986) proposed that *Muhlenbergia* is an offshoot of the *Eragrostis-Sporobolus* complex.

Duvall *et al.* (1994), however, found the placement of *Muhlenbergia* problematic. They found *Muhlenbergia* to group with other exclusive New World genera such as *Bealia* Scribn., *Chaboissaea* Fourn. and *Pereilema* Presl (Duvall *et al.* 1994). Prolamin size and immunological studies also do not support the grouping of *Calamovilfa* (A.Gray) Scribn., *Sporobolus* and *Muhlenbergia* in one tribe (Hilu and Esen 1993). This is supported by the fact that their delimiting feature is one-flowered spikelets, but two-flowered spikelets are also found in species of all three genera (Clayton and Renvoize 1986).

Chromosome numbers are related to the major phylogenetic groupings in the grass phylogeny. Avdulov (1931) first proposed this and the increase in the number of chromosome studies in the family is evidence of the recognition of this fact.

Hilu and Alice (2001) in their study based on *matK* chloroplast data proposed that a basic chromosome number of  $x = 10$  and characters such as paniculate inflorescences and pedicelled many-flowered spikelets, as found in the Eragrostideae, were polymorphic characters in the Chloridoideae. The evolution of  $x = 9$ , along with spicate inflorescences and a reduction in the number of fertile florets, as found in the Cynodonteae, happened soon after emergence of earliest chloridoids as these apomorphic traits were found in the most basal clades. These basal clades comprise of mainly the Pappophoreae, *Sporobolus* and species of *Eragrostis*. This same trend is seen in this *ITS* study. The Eragrostideae are seen as the more primitive of the groupings and the Cynodonteae more derived, although in this study this distinction is not obvious. In the 'Eragrostideae clade',  $x = 10$  is the predominant basic chromosome number with 9 and 10 occurring only in the Pappophoreae and in *Sporobolus*. The 'Cynodonteae clade' has  $x = 9$  and 10, 10 being more frequent but with  $x = 9$  more prevalent than in the 'Eragrostideae clade'. Mehra *et al.* (1968) also felt that  $x = 10$  became stabilized early in the history of the group, explaining the prevalence of multiples of 10.

These basic chromosome numbers, which are found in the Chloridoideae, are paleopolyploid according to Goldblatt (1980), or ancient taxa that are rediploidized polyploids with high basic chromosome numbers (Grant 1963; Goldblatt 1980). The establishment of these most likely occurred by an aneuploid reduction from  $x = 6$ , which lead to five. This was increased by polyploidization to the paleopolyploid

basic chromosome number of ten and subsequent aneuploid reduction lead to nine. This has been corroborated by various molecular studies (Barker *et al.* 1995, 1999; Hilu *et al.* 1999; Hilu and Alice 2001). The basic chromosome number of 6 is characteristic of the more basal grass subfamilies (Hunziker and Stebbins 1987) and also of the Arundinoideae, which is proposed to be the ancestor of the Chloridoideae. *Centropodia glauca*, *Merxmuellera rangei* and the two danthonioid species in this study are characterized by a basic chromosome number of 6.

## 6.4 Conclusions

*ITS* analyses have proved useful in the elucidation of the relationships between some representatives of the southern African Chloridoideae. The monophyly of the Chloridoideae and the problems with tribal delimitation have been observed in this study. Support for the ancient polyploidy origin of the two basic chromosome numbers of 9 and 10 and the aneuploid origin of  $x = 5$  from  $x = 6$  can be deduced from this study.

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## CHAPTER 7

# Chloroplast DNA sequencing and combined analyses in the Chloridoideae

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SUBMITTED (SYSTEMATIC BOTANY):

R. ROODT AND J.J. SPIES. *TRNL-F* AND COMBINED ANALYSIS IN REPRESENTATIVES OF THE CHLORIDOIDEAE (POACEAE) FROM SOUTHERN AFRICA. SUBMITTED (SYSTEMATIC BOTANY)

Phylogenetic relationships in 53 species of the subfamily Chloridoideae were investigated based on DNA sequences from the chloroplast *trnL* (UAA) 5' exon-*trnF* (GAA) exon region, using eight species from the grass clade PACCAD as outgroups. This data was used in combination with *ITS* data for this group and a combined matrix of 50 taxa was compiled, using 6 taxa from the Panicoideae and Danthoioideae as outgroups. The individual data sets were largely congruent and combined into a single data set, with the Chloridoideae found to be monophyletic and two groups, corresponding to the tribes Eragrostideae and Cynodonteae, polyphyletic in all three analyses. The basal position of the unioloid-pappophoroid group, seen as a link to the subfamily Danthoioideae, renders evidence of the origin of the Chloridoideae from arundinoid (in the broad sense) and more specifically danthonioid ancestors and renders molecular evidence for paleopolyploidy in the Chloridoideae. Putative hybrids could not be detected in the analyses, which indicates once again that hybrids do not behave predictably in phylogenetic analysis, as large-scale hybridization is a trademark of the grasses and subfamily Chloridoideae.

## 7.1 Introduction

The vast majority of extant grasses fall in two large groupings, namely the PACCAD clade and the BEP clade [Grass Phylogeny Working Group (GPWG 2001)]. More than half of the grass species are included in the PACCAD (Panicoideae-Arundinoideae-Chloridoideae-Centothecoideae-Aristidoideae-Danthoioideae) clade and from as early as the 1930s based on amongst others, chromosome studies (Avdulov 1931), taxa in this clade have been grouped together (GPWG 2001). One of the strongly supported lineages in this clade is the subfamily Chloridoideae. This is a tropical to subtropical subfamily with centers of distribution in southern Africa and Australia (Jacobs 1987). Together with the Panicoideae Link, which is also a member of the PACCAD clade, Chloridoideae Kunth ex Beilschm. comprise the largest majority of the southern African grass flora (Gibbs Russell 1986).

Two major tribes, the Eragrostideae Stapf and the Cynodonteae Dumort., are recognized in the subfamily although many authors feel this is not justified and proposed the merger of these two tribes (Campbell 1985; Watson and Dallwitz 1992 onwards). Various studies [for example Phillips 1982 (numerical morphological

analysis); Ellis 1987 (bundle sheath anatomy); Van den Borre and Watson 1997 (morphology and leaf anatomy); Hilu *et al.* 1999 (*matK* sequence data)] could also not distinguish between the two tribes. The smaller tribes recognized in the subfamily are the Leptureae Dumort., the Orcuttieae Reeder and the Pappophoreae Kunth (GPWG 2001).

Chloroplast DNA is being increasingly used in plant phylogenetic studies. This is due to the small size of the chloroplast genome, low rate of structural and sequence evolution, rarity of recombination and the uniparental inheritance of the genome (Palmer *et al.* 1988). The non-coding regions of the chloroplast genome evolve more rapidly than coding regions by accumulating indels at a rate almost equal to that of nucleotide substitutions (Curtis and Clegg 1984; Wolfe *et al.* 1987; Zurawski and Clegg 1987; Clegg and Zurawski 1992). This increases these regions' functionality at lower taxonomic levels, for example below family level.

The non-coding region investigated in this study comprises the *trnL* (UAA) intron, *trnL* (UAA) 3' exon, and the *trnL* (UAA) 3'-*trnF* (GAA) intergenic spacer (hereafter referred as the *trnL*-F region).

This region has been used at the intergeneric and interspecific level in various groups (for example Kita *et al.* 1995; Freeman and Ybarra 1996; Gielly and Taberlet 1996; Hauk *et al.* 1996; Gomez Martinez and Culham 1997; Compton *et al.* 1998).

In the grass family the region has been used at various taxonomic levels: the species *Poa jemtlandica* (Almq.) K.Richter (Brysting *et al.* 2000); *Spartina anglica* C.E.Hubb. (Baumel *et al.* 2001); the genera *Axonopus* P.Beauv. (Gomez-Martinez and Culham 1997); *Bouteloua* Lag. (Columbus *et al.* 2000); *Miscanthus* Anderss. (Hodkinson *et al.* 2002a) and *Spartina* Shreb. (Ferris *et al.* 1997); the panicoid tribes Andropogoneae Dumort. (Hodkinson *et al.* 2002b) and Paniceae R.Br. (Gomez-Martinez and Culham 2000; Doust and Kellogg 2002) and the pooid tribe Triticeae Dumort. (Mason-Gamer *et al.* 2002).

The occurrence of interspecific hybridization in plants has long been recognized (Stebbins 1950; Grant 1981). More than 50% of angiosperms have been estimated to have hybridization to some extent in their ancestry and Kellogg and Watson (1993) proposed it to be a major force in angiosperm evolution. This is based on the high frequency of polyploidy (80%, Masterson 1994) and the belief that



most polyploidy is associated with reticulation (De Wet 1987). Especially in the grass family, hybridization is thought to have played a major role in the evolution of the group with more than 80% of grasses believed to be polyploid (Stebbins 1985) and the frequency of polyploidy in the Chloridoideae, specifically more than 70% (Chapter 5).

Stebbins (1950) suggested that most plant genera that create problems in classification are of hybrid origin. Potential hybridization is an ongoing problem for phylogenetic analyses and uncertainties about its frequency obscure the interpretation of cladograms (Kellogg and Watson 1993). Due to the fact that hybridization introduces a reticulate pattern into an evolutionary cladogram, it cannot easily be contained in an analytical method that assumes strict divergence (Funk 1981; Cronquist 1987; Kellogg 1989).

Three approaches have been proposed to reconstruct hybrid speciation (Sang and Zhong 2000):

- Identification of hybrids beforehand.
- Detecting hybrids according to their behavior on the cladogram.
- Comparison of discordant positions of hybrids between phylogenetic cladograms obtained from independent data sets.

The identification of hybrids beforehand is not always that easy as the morphology of hybrids may be intermediate (McDade 1990), or completely unlike either parent (Rieseberg *et al.* 1990), or nearly indistinguishable from one parent (Bennett 1984).

Identifying hybrids on bases of the behavior on cladograms may also be problematic. Hybrids can jump between parents (McDade 1992), or place basally to lineage with the most derived parent (McDade 1990), or group with other hybrid taxa (McDade 1992). McDade (1992) also states that hybrids do not disrupt phylogenies as much as has been proposed and that hybrids cannot readily be identifiable solely on their behavior in cladograms. Furthermore, hybrids between distantly related parents cause more topological problems than do those between more closely related parents.

The last option of comparing independent data sets (for example nuclear versus chloroplast phylogenies) has been used increasingly in hybrid identification

as well as a means of increasing phylogenetic signal (Morrell and Rieseberg 1998; Widmer and Baltisberger 1999; Richardson *et al.* 2001; Zomlefer *et al.* 2001; Ferguson and Jansen 2002). Studies using nuclear and chloroplast regions enhance the potential of discovering for example hybridization because these genomes represent genetically unlinked regions (Andreasen *et al.* 1999). Furthermore, many studies in recent years have indicated that combined molecular data sets that utilize different levels of variation provide resolution at different levels of the cladogram and, therefore, phylogenetic resolution and bootstrap values are improved by combining separate data sets (Chase and Cox 1998; Soltis and Soltis 1998; Whitten *et al.* 2000).

When investigating hybridization, comparing incongruent gene trees renders opportunities to investigate ancient hybridization, where molecular additivity and morphological intermediacy have been obscured in the hybrid (Sang and Zhong 2000). In the grasses polyploidization is believed to be ancient. In the *ITS* study (Chapter 6), little polymorphism and molecular additivity was detected in the *ITS* regions and the basic chromosome numbers now present in the group ( $x = 9$  and  $10$ ) are believed to be paleopolyploid in origin, rendering further support for ancient hybridization in the grass family and subfamily Chloridoideae.

Our aim with this study is to extend the work on phylogenetic relationships among the Chloridoideae based on *ITS* data (Chapter 6) by using data from both nuclear and chloroplast genomes. In this study additional sequencing of the *trnL-F* region was done and this data used in conjunction with *ITS* data to explore whether putative hybrids could be identified and to render a phylogeny for southern African Chloridoideae based on different genomic regions.

## 7.2 Results

The assembled sequences of the entire *trnL-F* region of the 61 specimens consist of 1191 characters (Appendix G), 1339 with indel information included (Appendix H, I). The *trnL* intron had a consensus length of 617 bp and the *trnL-F* intergenic spacer consensus was 524 bp in length. For the entire region 240 characters (17.9%) are informative when gaps are considered as character states. The informative sites are distributed mostly in the intron (34.2%), the intergenic

spacer (32.1%) and the indels contributed 31.7% of the total number of informative sites. The percentage of adenine (A) and thymine (T) in the 61 specimens ranged from 66.2% [*Cynodon dactylon* (L.) Pers., AJ489466] to 69.5% (*Eustachys paspaloides* Lanza & Mattei, Spies 5597). The average AT content for the entire region was 67.6%.

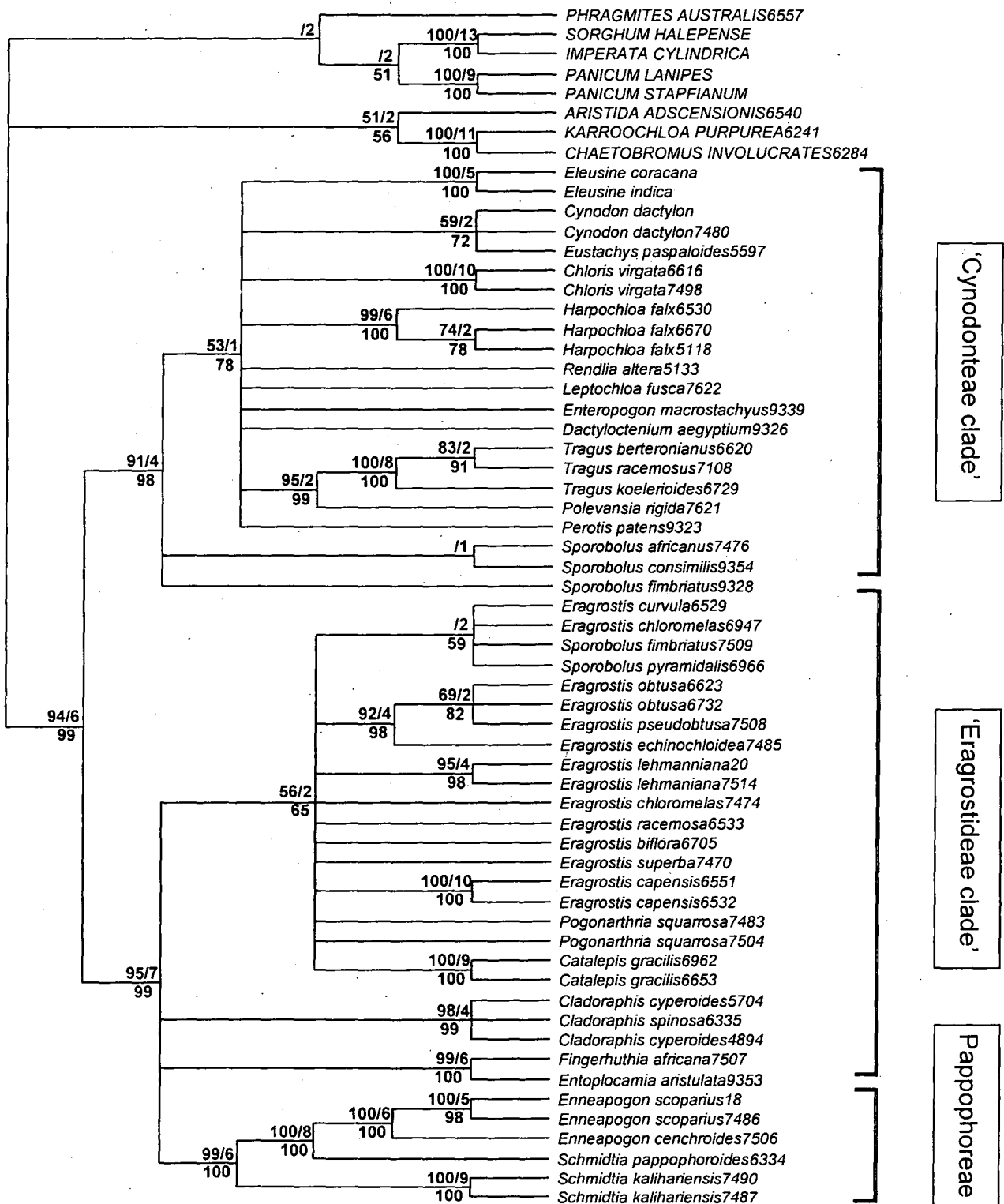
Indels were frequent in the *trnL-F* region. Many of these were phylogenetically informative for various species or genera (Table 7.1).

Phylogenetic analysis using heuristic search options yielded 20 000 equally most parsimonious cladograms with a length of 483, CI of 0.60 excluding uninformative characters and RI of 0.80 excluding uninformative characters. A strict consensus cladogram was used to summarize the data (Figure 7.1). Bootstrap (BS), jackknife (JK) and decay values (DV) are indicated on the cladogram. The monophyly of the Chloridoideae is strongly supported. Two large clades, corresponding approximately to the groups Eragrostideae and Cynodonteae, are found within the Chloridoideae as was the case with the *ITS* data (Figures 6.1) although with the *trnL-F* data these two groupings are very strongly supported (BS = 91%, 95%; JK = 98%, 99%; DV = 4, 7). The cladogram is largely congruent with the *ITS* cladogram except for the lack of resolution within these two groupings. Another difference is the grouping of three *Sporobolus* R.Br. specimens, which are grouped with the 'Cynodonteae clade'. In the *ITS* analysis these three formed part of the 'Eragrostideae clade'. This, however, forms a soft incongruence with very little significant BS and JK support for this grouping in both analyses.

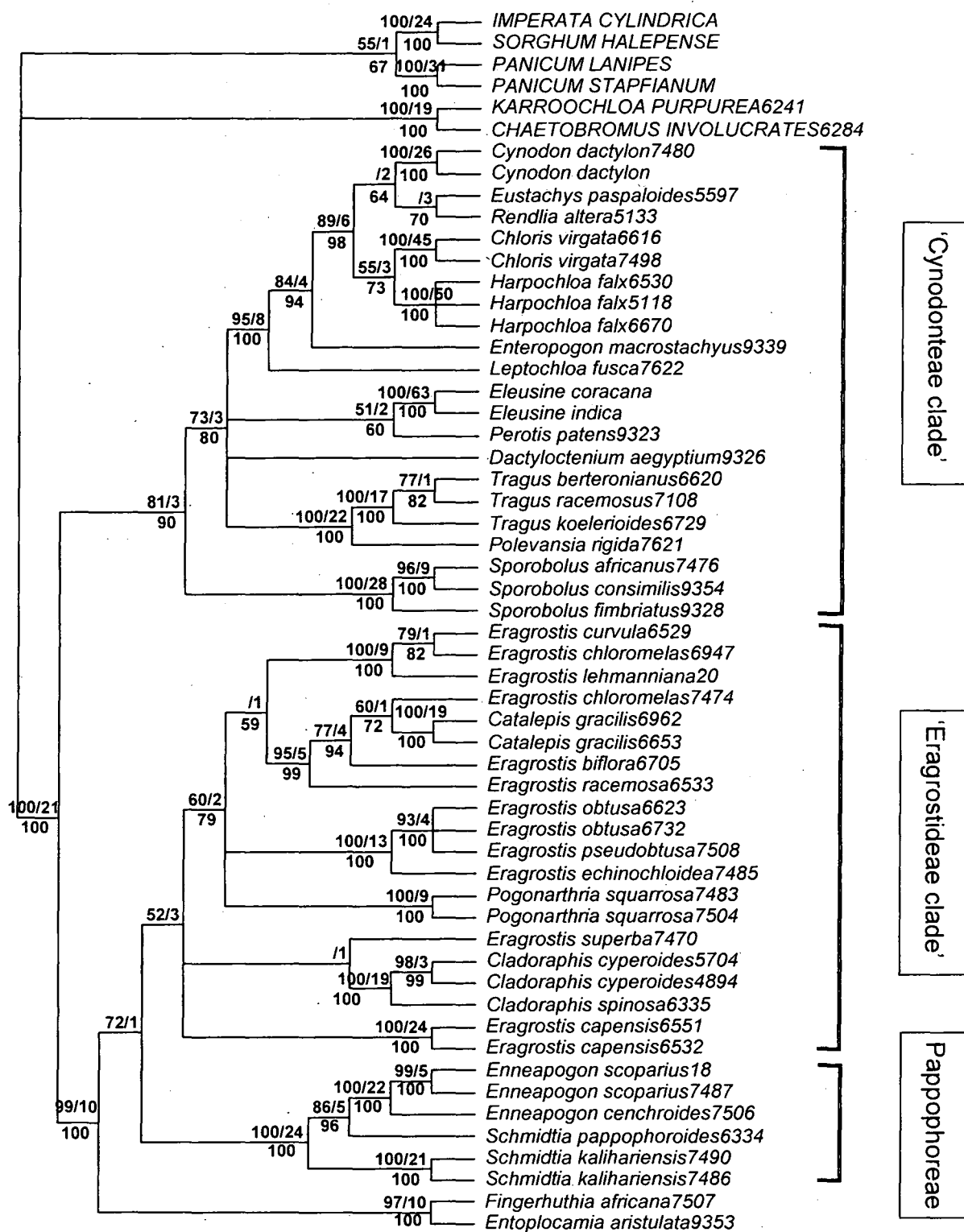
Because the two regions investigated produced nearly identical results, the regions were directly combined. The combined data matrix consisted of 1831 characters (Appendix J), 2098 including indels (Appendix K, L) of which 647 (30.8%) were parsimony informative. Analysis of this combined matrix resulted in 72 equally most parsimonious cladograms with a length of 2264, CI of 0.46 and RI of 0.72 (both excluding uninformative characters) (Figure 7.2). Although some polytomies are present (six), the combined cladogram provides resolution within the more basal and derived lineages of the Chloridoideae, whereas the *trnL-F* showed poor resolution within genera in the subfamily.

**Table 7.1.** List of phylogenetically informative indels.

Indel position	Taxa indel is phylogenetically informative for
106–109	<i>Eleusine coracana</i> <i>Eleusine indica</i>
155–160	<i>Eragrostis capensis</i> Spies 6532 <i>E. capensis</i> Spies 6551
450–455	<i>Chloris virgata</i> Spies 6616 <i>C. virgata</i> Spies 7498
774–778	<i>Eragrostis obtusa</i> Spies 6623 <i>E. obtusa</i> Spies 6732 <i>E. echinochloidea</i> Spies 7485 <i>E. pseudobtusa</i> Spies 7508
1053–1060	<i>Schmidtia kalihariensis</i> Spies 7486 <i>S. kalihariensis</i> Spies 7490
1102–1116	<i>Harpochloa falx</i> Spies 5118 <i>H. falx</i> Spies 6530 <i>H. falx</i> Spies 6670
1102–1108	<i>Cladoraphis cyperoides</i> Spies 4894 <i>C. cyperoides</i> Spies 5704 <i>C. spinosa</i> Spies 6335
1185–1188	<i>Harpochloa falx</i> Spies 5118 <i>H. falx</i> Spies 6530 <i>H. falx</i> Spies 6670



**Figure 7.1.** Strict consensus cladogram for *trnL-F* data sets produced from 20 000 equally most parsimonious cladograms with a length of 483, CI of 0.60 and RI of 0.80 (both excluding uninformative characters). Bootstrap and decay values (BS/DV) are indicated on branches. Jackknife values are indicated below branches.



**Figure 7.2.** Strict consensus cladogram obtained from combined *ITS* and *trnL-F* data sets. Eighteen equally most parsimonious cladograms with a length of 2186, CI of 0.46 and RI of 0.73 (both excluding uninformative characters) were found. Bootstrap and decay values (BS/DV) are indicated on branches. Jackknife values are indicated below branches.

## 7.3 Discussion

In contrast to the nuclear genome, which has a high GC content (almost 60% in the *ITS* study, Chapter 6), chloroplast DNA is more adenosine (A) and thymine (T) rich (Palmer 1991). This can be seen in this study with the *trnL-F* region in the Chloridoideae having an average AT content of 67%. Similar values have been found in other non-coding regions in Poaceae (R.Br.) Barnhart (Morton and Clegg 1995; Morton *et al.* 1997). This has implications for the type of base substitution to frequent the region as AT rich favors transversions and GC rich favors transitions (Morton 1997). Due to difference in length between the *trnL-F* regions of the different studied specimens, numerous indels had to be introduced. According to Palmer (1985) these small indels (1-1000bp) known as minor rearrangements, are most common in non-coding regions of chloroplast DNA and can be informative within closely related taxa (Doebley *et al.* 1987; Soltis *et al.* 1989, 1990b). These indels probably result from molecular mechanisms such as slipped strand mispairing (Takaiwa and Sugaira 1982); intramolecular recombination (Ogihara *et al.* 1988; Palmer 1991) and stem-loop formation (Sears *et al.* 1996).

As proposed by Hilu and Alice (2001), the classification of Clayton and Renvoize (1986) is used as a foundation for the discussion of the results. This is due to the fact that a large number of classification schemes have been proposed over the years and the number of tribes and subtribes might be confusing.

Recent studies have indicated that the use of non-coding chloroplast regions may only be useful at intergeneric but not infrageneric level due to a lack of informative sequence variation (Sang *et al.* 1997; Cros *et al.* 1998; Small *et al.* 1998; McDade and Moody 1999; Mummenhoff *et al.* 2001). This was seen in this study with the *trnL-F* region investigated providing resolution between, but not within, genera in the Chloridoideae.

As with the *ITS* data, the monophyly of the Chloridoideae is once again confirmed with the *trnL-F* (BS = 94%, JK = 99%, DV = 6) as well as combined (BS = 100%, JK = 100%, DV = 21) analyses. The sister group relationships between the Danthonioideae Barker & H.P.Linder and Arundinoideae Burmeister as proposed by the GPWG (2001) can be seen from the *trnL-F* cladogram with *Aristida* L.

(Aristidoideae Caro), *Karoochloa* Conert & Türpe and *Chaetobromus* Nees (Danthonioideae) grouping together, although the support for this is rather weak (BS = 51%, JK = 56%, DV = 2).

Two main clades, roughly comparable with the main tribes Eragrostideae and Cynodonteae, are once again found as was also evident from the *ITS* analysis. The support for the separation is, however, far stronger based on chloroplast data (BS = 91%, 95%; JK = 98%, 99%; DV = 4, 7) and in the combined analysis (BS = 81%, 99%; JK = 90%, 100%; DV = 3, 10). The consistent nature of division into two groupings in the Chloridoideae indicates that there exists a subdivision in the subfamily, but the current nature of the division seems to be superficial as the two tribes are clearly not monophyletic. For example, the genus *Catalepis* Stapf & Stent consistently groups in the 'Eragrostideae clade' with nuclear, *trnL-F* and combined analyses. In their study based on morphological and leaf anatomical data, Van den Borre and Watson (1997) did not place *Catalepis* into any of the five informal groupings they proposed. This genus has been classified in the subtribe Zoysiinae Benth. (Clayton and Renvoize 1986) of the Cynodonteae, but did not group with this subtribe in their analysis and they state that this genus 'probably belongs elsewhere'. The Zoysiinae share characters such as a bottlebrush inflorescence that is unusual in the Chloridoideae and *Catalepis* forms the link to the subtribe Chloridinae J.Presl (Clayton and Renvoize 1986). Clearly this genus is problematic and should be studied more thoroughly.

The Pappophoreae is one of the strongest supported groupings in all analyses (*ITS*: BS = 100%, JK = 100%, DV = 12; *trnL-F*: BS = 100%, JK = 99%, DV = 6; combined: BS = 100%, JK = 100%, DV = 24). This tribe is only seen as distantly related to the Eragrostideae but consistently groups with the 'Eragrostideae clade', although always in a basal position. Only two genera in the tribe, namely *Enneapogon* Desv. ex P.Beauv. and *Schmidtia* Steud. ex J.A.Schmidt, were included in this study. According to Clayton and Renvoize (1986), many awns and unusual microhairs distinguish this small tribe. Studies by Van den Borre and Watson (1997) and Hilu and Alice (2000, 2001) also found this tribe in a more basal position confirming its position as early diverging chloridoid lineage.



The two members of the subtribe Uniolae (*Entoplocamia* Stapf and *Fingerhuthia* Nees) form the most basal lineage of the 'Eragrostideae' grouping although with weak to moderate support (BS = 72%, JK = 82%, DV = 1). This only became apparent in the combined analysis. In all analyses these two genera group together (*ITS*: BS = 100%, JK = 99%, DV = 7; *trnL-F*: BS = 100%, JK = 99%, DV = 6; combined: BS = 97%, JK = 100%, DV = 10), but the basal position is only evident in the combined analysis. The basal position of this group was also found in the study by Hilu and Alice (2000, 2001) based on *matK* sequence data, and they called this an 'intriguing finding' due to the fact that no previous studies had indicated phylogenetic affinities between these two subtribes. This study corroborates their findings and provides further proof of the basal position of this group of grasses referred to as the 'pappophoroid-unioloid' group in the Chloridoideae and indicates the link between the Arundinoideae and the Chloridoideae as the Uniolae is seen as the link to the subfamily Arundinoideae through the genus *Tribolium* Desv. (now known as the subfamily Danthonioideae; GPWG 2001) (Clayton and Renvoize 1986). According to Clayton and Renvoize (1986) the similarities between the chloridoid subtribes Uniolae and Triodiinae and the arundinoid-danthonioid genera *Spartochloa* C.E.Hubb., *Styppeiochloa* DeWinter and *Tribolium* suggest that these are the primitive genera near the point of divergence of these subfamilies.

Except for *Eragrostis* Wolf and *Sporobolus* R.Br., most of the generic groupings are well supported. The two genera *Cladoraphis* Franch. and *Pogonarthria* Stapf form well supported lineages in the 'Eragrostideae clade' in all analyses conducted. Their generic status is clearly justified and their close relationship to *Eragrostis* is also evident. The pubescent rachilla tips in some *Eragrostis* species indicate affinities with *Pogonarthria* and this genus is seen as a link between the genera *Harpachne* Hochst. and *Eragrostis* (Clayton and Renvoize 1986). *Cladoraphis* was previously placed in *Eragrostis* by de Winter (1955), but later authors (Phillips 1982; Clayton and Renvoize 1986; Gibbs Russell *et al.* 1990; Watson and Dallwitz 1992 onwards) retained its separate generic status. It has a very distinctive morphology and very specific habitats, occurring in sandy desert [*C. spinosa* (L.f.) S.M.Phillips] and coastal dunes [*C. cyperoides* (Thunb.) S.M.Phillips] in southern Africa (Clayton and Renvoize 1986). Genera such as *Chloris* Sw.,

*Cynodon* Rich. and *Harpochloa* Kunth in the 'Cynodonteae clade' are also well supported in all analyses.

The polyphyletic nature of *Eragrostis* is once again confirmed by the ITS, *trnL-F* and combined analyses. This is the largest genus in the subfamily, and as Phillips (1982) stated, large genera are the exception to the rule, because the Chloridoideae seems to be undergoing adaptive radiation, resulting in many small but distinctive genera. This genus has always been problematic with 350 species included in the subfamily by Clayton and Renvoize (1986) and Watson and Dallwitz (1992 onwards) and the 'full range of morphological and anatomical variation found in Chloridoideae' exhibited in the genus (Van den Borre and Watson 1994). Species boundaries may further be confounded by interspecific and intergeneric hybridization in the genus (Vorster and Liebenberg 1977; Davidse *et al.* 1986; Burson and Voight 1996).

A grouping in the genus *Eragrostis*, which is apparent in the separate and combined data sets, is the connection between the species *E. echinochloidea* Stapf, *E. obtusa* Munro ex Ficalho & Hiern. and *E. pseudobtusa* DeWinter (ITS: BS = 100%, JK = 100%, DV = 11; *trnL-F*: BS = 92%, JK = 98%, DV = 4; combined: BS = 100%, JK = 100%, DV = 13). *Eragrostis pseudobtusa* is believed to be intermediate between *E. obtusa* and *E. echinochloidea* based on palea structure (Gibbs Russell *et al.* 1990).

The other polyphyletic genus in this study is *Sporobolus*. Like *Eragrostis*, this is also one of the larger genera in the subfamily (160 species) (the only other large genus is *Muhlenbergia* Schreb. with 160 species) (Clayton and Renvoize 1986; Watson and Dallwitz 1992 onwards). *Eragrostis* and *Sporobolus* are seen as closely related being separated based on 1-flowered spikelets and 1-nerved lemmas versus multi-flowered spikelets and 3-nerved lemmas in *Eragrostis* (Clayton and Renvoize 1986), but these authors felt that the distinction between the two genera 'is not as great as has been proposed'. Within the genus itself the boundaries are not sharply defined and species intergrade with one another (Clayton and Renvoize 1986; Watson and Dallwitz 1992 onwards; Duvall *et al.* 1994). Crawford (1983) offered three hypotheses explaining groups that are taxonomically difficult to delimit and in which the species boundaries are blurred morphologically:

- The taxa originated recently and divergence is not yet noticeable.
- There exists a high level of phenotypic plasticity that allows divergent genomes to converge phenotypically under similar conditions.
- Interspecific hybridization takes place between some populations of each species.

All three these phenomena are probably at play in the problematic taxa *Eragrostis* and *Sporobolus*.

The genera *Dactyloctenium* Willd., *Enteropogon* Nees and *Leptochloa* P.Beauv. occupy rather isolated position in all analyses. Hilu and Wright (1982) said these genera are difficult to place because they combine features delimiting both main tribes Eragrostideae and Cynodonteae, namely spiciform inflorescences and multi-flowered spikelets.

One of the main aims was to try and identify putative hybrids based on incongruence between the different data sets. In the grasses in general and the Chloridoideae specifically, hybridization and polyploidization are known to occur with high frequency (Chapters 4, 5). Therefore, not detecting any hard incongruence between the nuclear and chloroplast data sets was disappointing. Various explanations for this might be sought:

- The hybridizing groups may be closely related, for example *Eragrostis* and *Sporobolus*. According to McDade (1992) hybrids do probably not affect cladistic analyses as much, unless the hybrids are between distantly related taxa.
- Interlocus concerted evolution homogenized repeats from different arrays and effectively removed one parent from the hybrid (Wendel *et al.* 1995a; Ainouche and Bayer 1997) and this might have been the paternal parent, leaving no incongruence between the chloroplast (maternal) (Mogensen 1996) and nuclear (biparental) phylogenies. According to Hughes *et al.* (2002), where concerted evolution is present, direct evidence of hybrid speciation is not present any more.
- The hybridization might be too ancient to detect. Interlocus concerted evolution usually needs many sexual generations to reach completion (Wendel *et al.* 1995a; Brochman *et al.* 1996).

- No hybrids may have been included in the analyses. Especially in variable genera such as *Eragrostis* and *Sporobolus* not all of the variation might have been sampled.
- None of the putative parents of the hybrids present may be included in the study. McDade (1992) showed that many hybrids are placed between the two parents and as the basal member of the clade that includes the most derived parent. If no parent is included in the analysis this might be problematic for the hybrid and result in grouping with more distantly related taxa.

We believe basic chromosome numbers to be important indicators of evolution in the grasses and specifically the Chloridoideae. Many disagreements on which chromosome number developed from which, have led to many hypothesis regarding basic chromosome numbers and grass evolution (Mehra *et al.* 1968; Sharma 1979; Stebbins 1985). This study renders evidence of the relationship of the Chloridoideae to arundinoid (more specifically danthonioid) ancestors. Many previous authors have proposed this (e.g. Stebbins 1956; Clayton 1981; Phillips 1982; Campbell 1985; Clayton and Renvoize 1986; Conert 1987; Kellogg and Campbell 1987, Hilu and Esen 1993; Soreng and Davis 1998; Hsiao *et al.* 1999; Hilu and Alice 2001) but the basal position of *Entoplocamia*, *Fingerhuthia* and the Pappophoreae, seen as links to the Danthonioideae, once again provides cytogenetic proof. The basic chromosome number in the Danthonioideae (sister group in this study and found by GPWG 2001) is  $x = 6$ . Two other basic chromosome numbers also occur in this subfamily but only in certain genera [ $x = 7$  in *Pentameris* P.Beauv., *Prionanthium* Desv. and *Pseudopentameris* Conert and  $x = 13$  in *Pentaschistis* (Nees) Spach (for example De Wet 1960; Davidse *et al.* 1986; Spies and Du Plessis 1988; Morton 1993; Klopper *et al.* 1998; Spies and Roodt 2001)]. From  $x = 6$  an aneuploid or dysploid reduction leads to five which has been found in some chloridoid genera (Swami 1963; Pohl 1969; Davidse *et al.* 1986). Relatively early in their history the paleopolyploid basic chromosome numbers of  $x = 10$  and  $x = 9$ , which are the most prevalent in the Chloridoideae, became stabilized in the group. These two basic chromosome numbers are not restricted to any tribes, but  $x = 9$  is less frequent in the Eragrostideae than in the Cynodonteae.

## 7.3 Conclusions

To conclude, the high level of congruence among the data sets, representing nuclear and chloroplast regions and the high bootstrap values in the combined analysis indicate that the resultant cladogram is a good estimate of phylogenetic relationships in the group. We feel, as McDade *et al.* (2000), that the combined analyses of the *ITS* and *trnL-F* data sets reinforced more strongly supported components of the separate analyses.

No putative hybrids were identified. This should not be taken as an indication of no hybridization in this group. As there may be many reasons for incongruence between data sets (Wendel and Doyle 1998; Sang and Zhong 2000), not finding this incongruence does not imply that these processes are not at work, only that they were not satisfactorily detected.

This study renders molecular support for paleopolyploidy in the Chloridoideae and supports the notion that polyploidy along with its close counterpart, hybridization, is an essential part of the evolution of this subfamily.

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**CHAPTER 8**

**General conclusions**

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Poaceae is one of the largest families of plants in the world (Watson 1990; Liang and Hilu 1996). Its success can be related to morphological, geographical, cytogenetical, ecological and physiological diversity, which renders this one of the most successful plant families (Kahler and Price 1987).

Cytogenetically the most prevalent phenomenon in the grass family is polyploidy. It is important in the grasses as this phenomenon is rare in some groups and common in others (De Wet 1971) and Stebbins (1987) named polyploidization and hybridization the major evolutionary forces in Poaceae. In this study, polyploidy was found to be frequent in the grass subfamily Chloridoideae in general and specifically in southern Africa, which was the focus point of the study. More than 70% of the studied specimens were polyploid.

Studies on the nature of the polyploidy revealed that most of the tetraploid specimens were allopolyploids or segmental allopolyploids, clear evidence of a hybrid origin. The close relationship between hybridization and polyploidy is nothing new as the major role of polyploidy in evolution has been the fixing and spreading of hybrid combinations. It is also one of the quickest methods of producing completely different, but still vigorous and well-adapted genotypes (McWilliam 1964).

The survival and successful establishment of many newly formed hybrids are associated with an effective means of vegetative or asexual propagation (Stebbins 1956). In Poaceae this is found in the form of apomixis. Gametophytic apomixis is inseparably connected to both polyploidy and hybridization in restoring fertility to organisms that become sexually sterile as a result of polyploidy and hybridization (De Wet and Stalker 1974). According to Asker and Jerling (1992), most plants, characterized by gametophytic apomixis, are polyploid. More often they are allopolyploids formed after hybridization between sexual plants (Bierzychudek 1985). Grant (1981) and Mogie (1986) report that more than 99% of apomicts are polyploids and according to Goldblatt (1980) and Stebbins (1985) in the grass family, polyploids are mostly apomicts. Especially in the Chloridoideae and Panicoideae, this phenomenon is frequent with these two subfamilies comprising 52% of the known reports of apomixis in the grass family (Czapik 2000). Furthermore, apomixis is especially frequent in the grasses of South Africa (Brown and Emery 1957; Goldblatt 1978). In this study apomixis was not investigated, but based on previous

results, apomixis is seen to play an essential role in the high frequency of polyploidy and hybridization in the southern African Chloridoideae by providing an effective means of asexual reproduction.

According to Wendel (2000), diploidization after polyploidization is a common evolutionary event. This implies gene expression and chromosomal behavior of a diploid, although the plant has the amount of genetic material and the number of gene copies of a polyploid plant (e.g. Ohno 1970). This may happen by means of various mechanisms such as functional diversification of duplicated genes, gene silencing, long-term maintenance of similar but not identical function, or interactions between duplicated genes or their products (reviewed in Wendel 2000). This makes it difficult to distinguish more ancient polyploidization events. The processes that characterize diploidization, fragment the genomes and produce meso and paleopolyploids (Ehrendorfer 1980; Goldblatt 1980).

Paleopolyploids are defined as rediploidized polyploids with large basic chromosome numbers (Grant 1963; Goldblatt 1980). The basic chromosome numbers in the Chloridoideae are proposed to be paleopolyploid basic chromosome numbers. In this study the occurrence of two main basic chromosome numbers in the Chloridoideae, namely  $x = 9$  and  $10$  have been confirmed. The notion that these are derived from a lower basic chromosome number is based on the presence of primary diploids ( $x = 5$ ) reported for the subfamily. Furthermore, the basic chromosome number in the closely related Arundinoideae and Danthonioideae is  $x = 6$ , and  $x = 5$  could have developed from this by aneuploid or dysploid reduction. The origin of the chloridoids from arundinoid ancestors has been corroborated by various studies (for example Stebbins 1956; Renvoize 1981; Phillips 1982; Campbell 1985; Watson *et al.* 1985; Conert 1987; Kellogg and Campbell 1987; Davis and Soreng 1993; Liang and Hilu 1996; Soreng and Davis 1998; Hilu *et al.* 1999; Hsiao *et al.* 1999; Hilu and Alice 2001). This is corroborated by the fact that Stebbins (1971, 1980) concluded lower basic chromosome numbers is the more primitive condition and that evolution leads to an increase in basic chromosome number. Lastly, paleopolyploidy has been proven for the closely related Panicoideae (tribe Andropogoneae) based on various cytogenetic (for example Garber 1950; Celarier 1956) and molecular studies (for example Chittenden *et al.* 1994; Pereira *et al.* 1994; D'Hont *et al.* 1996; Ming *et al.* 1998).



In groups with high frequencies of hybridization, phylogenetic reconstruction may become problematic. Hybridization (reticulate evolution) is inconsistent with the bifurcating way to depicting hierarchies (Funk 1981, 1985; Cronquist 1987; Kellogg 1989). Stebbins (1956) proposed that the evolution of the grasses can be considered to be reticulate. One of the most widely used methods in identifying hybridization is by comparing independent data sets (Morrell and Rieseberg 1998; Widmer and Baltisberger 1999; Richardson *et al.* 2001; Zomlefer *et al.* 2001; Ferguson and Jansen 2002). Palmer (1985) stressed the importance of comparing the uniparental chloroplast genome with the biparental phylogeny derived from the nuclear genome in groups where hybridization appears to be frequent.

In this study nuclear and chloroplast DNA have been used to investigate the phylogeny of southern African Chloridoideae and to investigate whether putative hybrids could be identified based on incongruent results. These two regions investigated were the nuclear *ITS* and chloroplast *trnL-F* regions. The main difference between these two regions is in their mode of inheritance, with chloroplast DNA being typically maternally inherited (Mogensen 1996) and nuclear DNA being biparentally inherited.

In the analyses of the *ITS* and *trnL-F* data, the results were largely congruent. The Chloridoideae were found to be monophyletic, thus corroborating previous studies (for example Barker *et al.* 1995; Duvall and Morton 1996; Van den Borre and Watson 1997; Soreng and Davis 1998; Hsiao *et al.* 1999; Matthews *et al.* 2000; Hilu and Alice 2001). Although two groupings were distinguished in the analysis corresponding to the main tribes, Cynodonteae and Eragrostideae, both were polyphyletic. These tribes are separated mainly on inflorescence structure (Clayton and Renvoize 1986) and it may be that this issue needs to be readdressed, as Stebbins (1950) and Schlichting (1986) were of the opinion that morphological characters, especially in the grasses, are greatly influenced by the environment and exhibit a high degree of phenotypic plasticity.

With the *ITS* sequencing data very little polymorphism was found. *ITS* region polymorphisms in angiosperms are proposed to result from processes such as hybridization, polyploidy and agamospermy (Suh *et al.* 1993; Sang *et al.* 1995; Buckler *et al.* 1997; Campbell *et al.* 1997) known, and shown in this study, to have a

high frequency in the Chloridoideae. Various explanations for this may be sought but it is believed that ancient polyploidy, which has been proposed for the group in this study, has led to the lack of additivity with complete concerted evolution in the Chloridoideae (McDade 1995; Vargas *et al.* 1999).

Due to the congruence of the separate data sets the data could be combined and a combined analysis conducted. The combined results were also largely congruent with the individual data sets. The *trnL-F* phylogeny was the least resolved of the three analyses. This is not surprising, as the region has been shown not to resolve relationships at infrageneric level (Sang *et al.* 1997; Cros *et al.* 1998; Small *et al.* 1998; McDade and Moody 1999; Mummenhoff *et al.* 2001). The lack of incongruence resulted in the failure of successfully identifying hybrids in this study. McDade (1992) stated that this might happen because hybrids do not disrupt phylogenies as much as has been proposed and hybrids cannot readily be identifiable solely on their behavior in cladograms. Furthermore, the relatedness of the parents of the hybrids may also play a role with hybrids between distantly related parents causing more topological problems than do those between more closely related parents. Therefore, even in a group characterized by polyploidy and hybridization, well-resolved phylogenetic topologies can be obtained and hybrids not necessarily identified based on their cladistic behavior.

The molecular results in this study confirm the paleopolyploidy present in the Chloridoideae, which was evident from the cytogenetic results as well. Of the two main tribes in the Chloridoideae, the Eragrostideae is accepted to be the most primitive. The basic chromosome number in this group is mostly  $x = 10$ , with  $x = 9$  only occurring in the more specialized Pappophoreae and *Sporobolus*. In the less advanced Cynodonteae,  $x = 9$  is more prevalent but less so than  $x = 10$ . It is believed that the polyploidization from an ancestral  $x = 5$  occurred rather swiftly (Mehra *et al.* 1968), with subsequent aneuploid reduction to  $x = 9$ . This is probably one of the single most important events that set the Chloridoideae on an evolutionary path of their own.

Grass phylogeny forms the basis for various functional, genomic and evolutionary studies (Gaut 2002) and, therefore investigating and understanding the evolutionary relationships in this group are essential when studying for example

cytogenetic relationships. But the opposite is also true. In this study the importance of incorporating cytogenetic data into phylogenetic studies was investigated. Without cytogenetic knowledge the magnitude of an occurrence such as polyploidy and especially paleopolyploidy might be greatly underestimated.

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**CHAPTER 9**  
**Literature cited**

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# Appendices

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**APPENDIX A.** Gametic chromosome numbers (n) of representatives of the subfamily Chloridoideae (Poaceae) in southern Africa with their voucher specimen numbers and provincial localities. Species are listed alphabetically. Separate entries for the same province for a species indicate different localities.

Taxon	n	Voucher	Locality
<b>Cynodonteae</b>			
<i>Chloris virgata</i>	10	Spies 6616	FREE STATE
<i>Cynodon dactylon</i>	18	Spies 2549	SWAZILAND
		Spies 2966	NORTHERN CAPE
<i>Enteropogon macrostachyus</i>	20	Venter 9339	NAMIBIA
<i>Harpochloa falx</i>	18	Spies 5113	MPUMALANGA
		Spies 5065	FREE STATE
		Spies 5078	FREE STATE
		Spies 6629	FREE STATE
		Spies 3986	EASTERN CAPE
		Spies 4729	EASTERN CAPE
		Spies 4691	EASTERN CAPE
		Spies 4695	EASTERN CAPE
		Spies 6955	EASTERN CAPE
			27
		Spies 5128	MPUMALANGA
		Spies 4712	EASTERN CAPE
<i>Tragus</i> sp.	10	Spies 4803	FREE STATE
<b>Eragrostideae</b>			
<i>Bewisia biflora</i>	20	Spies 1531	MPUMALANGA
<i>Cladoraphis cyperoides</i>	10	Spies 5704	NORTHERN CAPE
		Spies 4894	NORTHERN CAPE
		Spies 4889	NORTHERN CAPE
		Spies 5356	WESTERN CAPE
<i>C. spinosa</i>	20	Spies 4885	NORTHERN CAPE
<i>Dactyloctenium aegyptium</i>	20	Spies 2403	KWAZULU-NATAL
<i>Eleusine coracana</i> subsp. <i>africana</i>	9	Spies 2365, 2366	KWAZULU-NATAL
	18	Spies 2783	WESTERN CAPE
<i>Eragrostis barbinodes</i>	20	Spies 3317	LIMPOPO
<i>E. capensis</i>	10	Spies 4696	EASTERN CAPE
	20	Spies 1595	MPUMALANGA
		Spies 5069	FREE STATE
		Du Plessis 110	KWAZULU-NATAL

<i>E. capensis</i>	20	<i>Spies 3483</i>	EASTERN CAPE
		<i>Spies 3498</i>	EASTERN CAPE
		<i>Davidse 33552</i>	LOCALITY UNKNOWN
<i>E. chloromelas</i>	20	<i>Spies 6947</i>	EASTERN CAPE
<i>E. ciliaris</i>	10	<i>Du Plessis 111</i>	KWAZULU-NATAL
<i>E. curvula</i>	30	<i>Spies 1137</i>	EASTERN CAPE
<i>E. echinochloidea</i>	20	<i>Spies 2799, 2800</i>	NORTHERN CAPE
<i>E. heteromera</i>	20	<i>Spies 2634</i>	SWAZILAND
<i>E. inamoena</i>	20	<i>Spies 2392</i>	KWAZULU-NATAL
<i>E. obtusa</i>	10	<i>Spies 2886</i>	NORTHERN CAPE
<i>E. planiculmis</i>	30+0-4B	<i>Spies 1116</i>	EASTERN CAPE
<i>E. racemosa</i>	20	<i>Spies 5066</i>	FREE STATE
		<i>Spies 3279</i>	KWAZULU-NATAL
		<i>Spies 4743</i>	EASTERN CAPE
<i>E. sclerantha</i> subsp. <i>sclerantha</i>	20	<i>Spies 4844</i>	FREE STATE
<i>E. superba</i>	20	<i>Spies 3326</i>	LIMPOPO
		<i>Du Plessis 136</i>	KWAZULU-NATAL
<i>E. tef</i>	20	<i>Spies 2405</i>	KWAZULU-NATAL
<i>E. tenuifolia</i>	10	<i>Spies 2595</i>	SWAZILAND
<i>E. trichophora</i>	20	<i>Spies 2774</i>	WESTERN CAPE
<i>Fingerhuthia africana</i>	20	<i>Spies 2947</i>	NAMIBIA
		<i>Spies 4349</i>	WESTERN CAPE
<i>Leptochloa fusca</i>	10	<i>Spies 4875</i>	NORTHERN CAPE
		<i>Spies 2991A</i>	NORTHERN CAPE
		<i>Spies 3037</i>	NORTHERN CAPE
		<i>Spies 4316</i>	NORTHERN CAPE
		<i>Spies 3794</i>	WESTERN CAPE
		<i>Spies 5200</i>	WESTERN CAPE
		<i>Spies 3932</i>	WESTERN CAPE
		<i>Davidse 33407</i>	LOCALITY UNKNOWN
<i>Odyssea paucinervis</i>	18	<i>Spies 3384</i>	NORTHERN CAPE
<i>Sporobolus africanus</i>	9	<i>Spies 4508</i>	WESTERN CAPE
	18	<i>Spies 2369</i>	KWAZULU-NATAL
<i>S. albicans</i>	27	<i>Spies 3141</i>	WESTERN CAPE
<i>S. ioclados</i>	9	<i>Spies 2717</i>	NORTHERN CAPE
<i>S. virginicus</i>	9	<i>Du Plessis 122</i>	KWAZULU-NATAL
<i>Stiburus alopecuroides</i>	10	<i>Spies 1470</i>	MPUMALANGA
<i>S. conrathii</i>	10	<i>Du Plessis 19</i>	MPUMALANGA
<i>Trichoneura grandiglumes</i>	10	<i>Spies 4833</i>	FREE STATE



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<b>Pappophoreae</b>			
<i>Enneapogon cenchroides</i>	20	<i>Spies 3288</i>	LIMPOPO
	30	<i>Spies 2709</i>	FREE STATE
<i>E. pretoriensis</i>	10	<i>Spies 3716</i>	NORTH-WEST
<i>Enneapogon sp.</i>	20	<i>Spies 5532</i>	NORTH-WEST
<i>Schmidtia pappophoroides</i>	18	<i>Du Plessis 186</i>	LIMPOPO
<i>Schmidtia sp.</i>	18+0-4B	<i>Spies 5536</i>	NORTH-WEST
<b>Unplaced</b>			
<i>Centropodia glauca</i>	24	<i>Spies 5706</i>	NORTHERN CAPE

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**APPENDIX B.** Chromosome numbers for studies done on southern African Chloridoideae. Chromosome numbers found during present study are indicated by \*.

Species	x	n	# of specimens examined	Reference
<b>Cynodonteae</b>				
<i>Chloris gayana</i> Kunth	10	10	8	Hunter 1934; Moffet and Hurcombe 1949; Davidse <i>et al.</i> 1986; Spies and Jonker 1987; Strydom and Spies 1994b
		20	8	Moffet 1944; Moffet and Hurcombe 1949
<i>C. pycnothrix</i> Trin.	10	15	1	De Wet 1954a
		20	1	Moffet and Hurcombe 1949
<i>C. virgata</i> Sw.	10	10	12	Moffet and Hurcombe 1949; Spies and Jonker 1987; Strydom and Spies 1994b; *
		20	1	Spies and Du Plessis 1987
<i>Craspedorachis</i> <i>rhodesiana</i> Rendle	9	27/2	1	Moffet and Hurcombe 1949
<i>Ctenium concinnum</i> Nees	10	c.50	1	Spies and Du Plessis 1986a
		80	1	De Wet 1960
<i>Cynodon bradleyi</i> Stent	9	9	1	Hurcombe 1946
<i>C. dactylon</i> (L.) Pers.	10	20	3	Hurcombe 1947; Moffet and Hurcombe 1949
		18	5	Strydom and Spies 1994b; *
<i>C. magennisii</i> Hurcombe	10	15	1	Hurcombe 1947
<i>C. plectostachyum</i> Pilg.	9	9	1	Moffet and Hurcombe 1949
		27	1	Moffet and Hurcombe 1949
<i>C. transvaalensis</i> Burt Davy	10	10	1	Hurcombe 1947
<i>Enteropogon</i> <i>macrostachyus</i> (A.Rich.) Benth.	10	20	1	*
<i>Eustachys paspaloides</i>	10	20	1	Strydom and Spies 1994b
Lanza & Mattei	9	18	1	De Wet 1960

<i>Harpochloa falx</i> (L.f.) Kuntze	10	20	12	De Wet 1958; Spies and Du Plessis 1986a; Spies <i>et al.</i> 1991; Strydom and Spies 1994b
		25	2	Strydom and Spies 1994b
		30	3	Strydom and Spies 1994b
	9	27	3	*
		18	9	*
<i>Lintonia nutans</i> Stapf	10	15	1	De Wet 1960
<i>Microchloa caffra</i> Nees	10	10	1	De Wet and Anderson 1956
		40	1	Spies and Du Plessis 1986b
		c.50	5	Spies and Du Plessis 1986a; Strydom and Spies 1994b
<i>M. kunthii</i> Desv.	10	20	1	Moffet and Hurcombe 1949
<i>Perotis patens</i> Gand.	10	20	3	Moffet and Hurcombe 1949; De Wet 1960; Davidse <i>et al.</i> 1986
	9	18	1	De Wet and Anderson 1956
<i>Rendlia altera</i> (Rendle) Chiov.	10	20	9	Strydom and Spies 1994b
<i>Tragus berteronianus</i> Schult.	10	10	2	Strydom and Spies 1994b
<i>T. racemosus</i> (L.) All.	10	10	1	Strydom and Spies 1994b
		20	1	De Wet 1954a
<i>Tragus</i> sp.	10	10	1	*
<b>Eragrostideae</b>				
<i>Bewisia biflora</i> (Hack.) Gooss.	10	15	1	De Wet and Anderson 1956
		20	1	*
		45/2	1	Davidse <i>et al.</i> 1986
<i>Cladoraphis cyperoides</i> (Thunb.) S.M.Phillips	10	10	4	*
<i>C. spinosa</i> (L.f.) S.M.Phillips	10	20	1	*
<i>Dactyloctenium aegyptium</i> (L.) Willd.	10	20	1	*
	9	18	1	Moffet and Hurcombe 1949
<i>D. giganteum</i> Fisch. & Schweick.	10	5	1	Davidse <i>et al.</i> 1986
<i>Diandrochloa namaquensis</i> (Nees) DeWinter (as <i>Eragrostis namaquensis</i> Nees ex Schrad.)	10	10	1	Moffet and Hurcombe 1949

<i>Dinebra retroflexa</i> Panz.	10	10	1	De Wet and Anderson 1956
var. <i>condensata</i>				
S.M.Phillips				
<i>Eleusine coracana</i> (L.) Gaertn.	9	18	1	Davidse <i>et al.</i> 1986
<i>Eleusine coracana</i> (L.) Gaertn. subsp. <i>africana</i> (Kenn.-O'Byrne) Hilu & DeWet	9	9	4	Spies and Voges 1988; *
		18	2	Spies and Du Plessis 1986a; *
<i>E. indica</i> (L.) Gaertn.	9	18	2	Moffet and Hurcombe 1949; De Wet 1954a
<i>Eragrostis acraea</i> DeWinter	10	10	1	Davidse <i>et al.</i> 1986
<i>E. aethiopica</i> Chiov.	10	10	1	Jones <i>et al.</i> 1978
<i>E. annulata</i> Rendle ex Scott-Elliott	10	10	1	Nordenstam 1969
<i>E. aspera</i> (Jacq.) Nees	10	10	3	Avdulov 1931; Moffet and Hurcombe 1949
<i>E. barbinodes</i> Hack.	10	10	1	Spies and Du Plessis 1986a
		20	4	Pienaar 1953; De Wet 1958; *
		25	1	Spies and Jonker 1987
<i>E. barrelieri</i> Daveau	10	30	1	Moffet and Hurcombe 1949
<i>E. biflora</i> Hack. ex Schinz	10	10	2	Moffet and Hurcombe 1949
<i>E. caesia</i> Stapf	10	20	1	Davidse <i>et al.</i> 1986
<i>E. capensis</i> (Thunb.) Trin.	10	10	2	De Wet 1958; *
		20	12	Avdulov 1931; Pienaar 1953; Davidse <i>et al.</i> 1986; Spies and Du Plessis 1986a; Spies <i>et al.</i> 1991; *
		30	2	Moffet and Hurcombe 1949; Spies and Voges 1988
<i>E. chapelieri</i> (Kunth) Nees	10	10	1	Moffet and Hurcombe 1949
<i>E. chloromelas</i> Steud.	10	20	3	Pienaar 1953; Vorster and Liebenberg 1977; *
		41/2	1	Pienaar 1953
		30	2	Pienaar 1953
		61/2	1	Pienaar 1953
		31	2	Pienaar 1953
		63/2	1	Pienaar 1953

<i>E. cilianensis</i> (All.) Vignolo ex Janch.	10	10	2	Moffet and Hurcombe 1949; Spies <i>et al.</i> 1991
<i>E. ciliaris</i> (L.) R.Br.	10	10	2	De Wet 1960; *
		20	1	Moffet and Hurcombe 1949
<i>E. crassinervis</i> Hack.	10	20	1	Spies and Du Plessis 1986b
<i>E. curvula</i> (Schrad.) Nees	10	10	8	Pienaar 1953; Spies 1982
		20	31	Nielson 1939; Pienaar 1953; De Wet 1958; Vorster and Liebenberg 1977; Spies 1982; Spies and Du Plessis 1986a, 1986b; Spies and Jonker 1987
		25	10	De Wet 1954a; Vorster and Liebenberg 1977; Spies 1982; Davidse <i>et al.</i> 1986; Spies and Du Plessis 1986b
		30	28	Pienaar 1953; Vorster and Liebenberg 1977; Spies 1982; Spies and Jonker 1987; *
		35	11	Spies 1982
		40	8	Vorster and Liebenberg 1977; Spies 1982; Spies and Jonker 1987
<i>E. curvula</i> (Schrad.) Nees (as <i>E. robusta</i> Stent)	10	35	4	Pienaar 1953
<i>E. curvula</i> (Schrad.) Nees var. 1 (as <i>E. robusta</i> Stent)	10	35	1	Pienaar 1953
<i>E. curvula</i> (Schrad.) Nees var. 2 (as <i>E. robusta</i> Stent)	10	35	1	Pienaar 1953
<i>E. curvula</i> (Schrad.) Nees var. 3 (as <i>E. robusta</i> Stent)	10	40	1	Pienaar 1953
<i>E. echinochloidea</i> Stapf	10	15	1	De Wet 1960
		20	6	Moffet and Hurcombe 1949; Pienaar 1953; Spies and Du Plessis 1987; *
		30	2	Spies and Du Plessis 1987; Spies and Voges 1988
<i>E. gummiflua</i> Nees	10	20	2	Pienaar 1953; De Wet 1954a
		30	1	Spies and Du Plessis 1986a

<i>E. habrantha</i> Rendle	10	30	1	Moffet and Hurcombe 1949
		45	1	Moffet and Hurcombe 1949
<i>E. heteromera</i> Stapf	10	20	3	De Wet 1958, 1960; *
<i>E. hierniana</i> Rendle (as <i>E. uniglumes</i> Hack.)	10	20	2	Pienaar 1953; De Wet 1958
<i>E. inamoena</i> K.Schum.	10	20	1	*
<i>E. inamoena</i> K.Schum. (as <i>E. atrovirens</i> (Desf.) Trin.)	10	20	1	De Wet 1960
<i>E. inamoena</i> K.Schum. (as <i>E. galpinii</i> Stent)	10	20	1	Pienaar 1953
		41/2	1	Pienaar 1953
<i>E. lappula</i> Nees	10	20	2	De Wet 1954a, 1958
<i>E. lappula</i> var. <i>divaricata</i> Stapf	10	20	1	De Wet and Anderson 1956
<i>E. lehmanniana</i> Nees	10	20	6	Pienaar 1953; Spies and Du Plessis 1987; Spies and Jonker 1987; Spies and Voges 1988
		30	2	Spies and Du Plessis 1986b; Spies and Jonker 1987
<i>E. micrantha</i> Hack. ex Schinz	10	20	2	De Wet 1958; Spies and Du Plessis 1986a
<i>E. nindensis</i> Ficalho & Hiern	10	20	1	Davidse <i>et al.</i> 1986
<i>E. nindensis</i> Ficalho & Hiern (as <i>E. denudata</i> Hack. ex Schinz)	10	10	2	De Wet and Anderson 1956; De Wet 1960
		20	1	Moffet and Hurcombe 1949
<i>E. obtusa</i> Munro ex Ficalho & Hiern	10	10	2	De Wet 1960; *
		20	4	Moffet and Hurcombe 1949; Pienaar 1953; Spies and Du Plessis 1986b
		30	1	Spies and Du Plessis 1986b
<i>E. pallens</i> Hack. ex Schinz	10	10	1	Moffet and Hurcombe 1949
<i>E. patens</i> Oliv.	10	10	1	Moffet and Hurcombe 1949
<i>E. plana</i> Nees	10	10	6	Pienaar 1953; Spies and Jonker 1987
<i>E. planiculmis</i> Nees	10	21	1	Spies and Jonker 1987
		30	1	Spies and Du Plessis 1986b
		30+0-4B	1	*
<i>E. pseudobtusa</i> DeWinter	10	20	2	Pienaar 1953; Spies and Du Plessis 1987
		40	1	Spies and Du Plessis 1987

<i>E. pseudosclerantha</i>	10	20	2	Spies and Du Plessis 1986a
Chiov.		25	1	De Wet 1960
<i>E. racemosa</i> (Thunb.)	10	20	3	*
Steud.		30	1	De Wet 1958
<i>E. racemosa</i> (Thunb.)	10	30	1	Pienaar 1953
Steud. (as <i>E. chalcantha</i>		31	1	Pienaar 1953
Trin.)				
<i>E. rigidior</i> Pilg.	10	20	1	De Wet 1960
<i>E. rotifer</i> Rendle	10	20	1	Nordenstam 1969
<i>E. rotifer</i> Rendle (as <i>E.</i>	10	20	1	Moffet and Hurcombe 1949
<i>margaritacea</i> Stapf)				
<i>E. sarmentosa</i> Trin.	10	20	1	Spies <i>et al.</i> 1991
<i>E. sclerantha</i> Nees	10	20	1	Davidse <i>et al.</i> 1986
<i>E. sclerantha</i> Nees (small	10	20	1	Moffet and Hurcombe 1949
type)				
<i>E. sclerantha</i> Nees (large	10	20	1	Moffet and Hurcombe 1949
type)				
<i>E. sclerantha</i> Nees subsp.	10	20	2	Spies and Du Plessis 1986a; *
<i>sclerantha</i>				
<i>E. stapfii</i> DeWinter	10	10	1	De Wet 1960
<i>E. stapfii</i> DeWinter (as <i>E.</i>	10	30	1	Pienaar 1953
<i>sporoboloides</i> Stapf)				
<i>E. superba</i> Peyr.	10	20	8	Moffet and Hurcombe 1949; Pienaar
				1953; De Wet 1954a; Spies and Du
				Plessis 1986a, 1986b, Spies and
				Jonker 1987; *
		25	1	Spies and Du Plessis 1986b
		40	1	Spies and Jonker 1987
<i>E. tef</i> (Zucc.) Trotter	10	20	2	Moffet and Hurcombe 1949; *
<i>E. tenuifolia</i> (A.Rich.)	10	10	1	*
Steud.				
<i>E. trichophora</i> Coss. & Dur.	10	20	2	Davidse <i>et al.</i> 1986; *
		30	1	Spies and Jonker 1987
<i>E. trichophora</i> Coss. & Dur.	10	10	3	Moffet and Hurcombe 1949; Pienaar
(as <i>E. atherstonei</i> Stapf)				1953
<i>E. truncata</i> Hack.	10	10	1	Moffet and Hurcombe 1949
<i>E. viscosa</i> Trin.	10	20	2	Moffet and Hurcombe 1949; De Wet
				1958

<i>E. walterii</i> Pilg.	10	20	2	Spies and Du Plessis 1986b
<i>E. wilmaniae</i> C.E.Hubb. & Schweick.	10	10	1	Moffet and Hurcombe 1949
<i>E. wilmsii</i> Stapf	10	20	1	De Wet 1954a
<i>Eragrostis</i> sp.	10	10	1	Pienaar 1953
<i>Fingerhuthia africana</i> Lehm.	10	20	5	De Wet 1958, 1960; Spies and Du Plessis 1987; *
<i>F. sesleriiformis</i> Nees	10	10	2	De Wet 1958, 1960
<i>Leptochloa fusca</i> (L.) Kunth	10	10	9	Spies <i>et al.</i> 1991; *
		19	1	Spies and Voges 1988
<i>Odyssea paucinervis</i> (Nees) Stapf	9	18	1	*
<i>Oropetium capense</i> Stapf	10	10	1	De Wet 1960
<i>Pogonarthria squarrosa</i> (Roem. & Schult.) Pilg.	10	20	4	Moffet and Hurcombe 1949; De Wet 1954a; De Wet and Anderson 1956; Davidse <i>et al.</i> 1986
		21	1	Moffet and Hurcombe 1949
		40	1	Spies and Jonker 1987
<i>Sporobolus africanus</i> (Poir.) Robyns & Tournay	10	30	2	Spies and Jonker 1987
	6	12	1	Spies <i>et al.</i> 1991
		24	1	Spies <i>et al.</i> 1991
	9	9	1	*
		18	3	Spies and Du Plessis 1986b; Spies and Voges 1988; *
		18+0-6B	1	Spies <i>et al.</i> 1991
<i>S. africanus</i> (Poir.) Robyns & Tournay (as <i>S. capensis</i> (Willd.) Kunth)	9	9	1	De Wet 1954a
		18	1	Moffet and Hurcombe 1949
<i>S. albicans</i> Nees	9	27	1	*
<i>S. festivus</i> A.Rich.	9	18	1	De Wet 1960
<i>S. fimbriatus</i> (Trin.) Nees	9	9	2	Moffet and Hurcombe 1949; De Wet 1958
		18	2	De Wet and Anderson 1956; De Wet 1958
<i>S. ioclados</i> (Trin.) Nees	9	9	1	*
<i>S. ioclados</i> (Trin.) Nees (as <i>S. smutsii</i> Stent)	9	18	2	De Wet and Anderson 1956; De Wet 1958
<i>S. nitens</i> Stent	9	9	1	De Wet and Anderson 1956



<i>S. panicoides</i> A.Rich.	10	10	2	Davidse <i>et al.</i> 1986
	6	12	1	Moffet and Hurcombe 1949
<i>S. pectinatus</i> Hack.	10	20	1	Spies and Jonker 1987
<i>S. piliferus</i> Kunth	9	18	1	Davidse <i>et al.</i> 1986
<i>S. pyramidalis</i> P.Beauv.	10	15	3	Moffet and Hurcombe 1949; De Wet 1960; Spies and Voges 1988
	6	12	2	Moffet and Hurcombe 1949; Spies and Voges 1988
	9	18	1	De Wet 1958
<i>S. stapfianus</i> Gand.	6	12	1	De Wet 1960
	9	18	1	De Wet and Anderson 1956
<i>S. virginicus</i> (L.) Kunth	10	15	1	De Wet 1960
	9	9	1	*
<i>Stiburus alopecuroides</i> (Hack.) Stapf	10	10	2	Spies and Du Plessis 1986a; *
<i>S. conrathii</i> Hack.	10	10	1	*
<i>Tetrachne dregei</i> Nees	10	10	3	Moffet and Hurcombe 1949; De Wet and Anderson 1956; De Wet 1960
<i>Trichoneura grandiglumes</i> (Nees) Ekman	10	10	3	Moffet and Hurcombe 1949; De Wet and Anderson 1956; *
<i>Triraphis andropogonoides</i> (Steud.) Phillips	10	10	1	De Wet 1954a
<i>T. purpurea</i> Hack. (as <i>T. fleckii</i> Hack.)	10	10	1	Hoshino and Davidse 1988
<i>T. ramosissima</i> Hack.	10	10	2	Hoshino and Davidse 1988
<b>Pappophoreae</b>				
<i>Enneapogon brachystachyus</i> Stapf	9	18	1	De Wet and Anderson 1956
<i>E. cenchroides</i> (Roem. & Schult.) C.E.Hubb.	10	20	3	Reeder and Singh 1968; Spies and Voges 1988; *
		24	1	Davidse <i>et al.</i> 1986
		30	1	*
	9	18	2	Thomas, unpubl (listed Darlington and Wylie 1955); De Wet and Anderson 1956
<i>E. pretoriensis</i> Stent	10	10	1	*
	9	9	2	De Wet and Anderson 1956; De Wet 1958

<i>E. scaber</i> Lehm.	10	20	1	Spies and Voges 1988
<i>E. scoparius</i> Stapf	10	20	1	Nordenstam 1982
	9	18	3	De Wet 1954a, 1958; De Wet and Anderson 1956
<i>Enneapogon</i> sp.	10	20	1	*
<i>Kaokochloa nigrirostris</i> DeWinter	10	11	1	Reeder and Singh 1968
<i>Schmidtia pappophoroides</i> Steud.	10	20	1	Reeder and Singh 1968
	9	18	1	*
<i>S. pappophoroides</i> Steud. (as <i>S. bulbosa</i> Stapf)	9	18	1	De Wet 1958
<i>S. pappophoroides</i> Steud. (as <i>S. glabra</i> Pilg.)	9	18	1	De Wet and Anderson 1956
<i>Schmidtia</i> sp.	9	18+0-4B	1	*
<b>Unplaced</b>				
<i>Centropodia glauca</i> (Nees) Cope	6	24	5	Du Plessis and Spies 1988; Hoshino and Davidse 1988; *
<i>C. forskalii</i> (Vahl) Cope (as <i>Danthonia forskalii</i> R.Br.)	6	12	1	De Wet 1954b
<i>Merxmuellera rangei</i> (Pilg.) Conert	6	18	1	Du Plessis and Spies 1988

**APPENDIX C.** Chromosome numbers for studies done in the Chloridoideae (excluding southern African specimens - see appendix B). The number of specimens examined, are indicated in brackets after each corresponding chromosome count. The summary listings of Ornduff (1967, 1968, 1969); Federov (1969); Moore (1970, 1971, 1972, 1973, 1974, 1977); Goldblatt (1980, 1981, 1983, 1985, 1988) and Goldblatt and Johnson (1990, 1991, 1994, 1996, 1998, 2000) were used to compile the table and original authors are not cited individually.

Species	x	n	Reference
<i>Acrachne racemosa</i> Ohwi	9	18	Goldblatt and Johnson 1994
<i>Aegopogon cenchroides</i> Humb. & Bonpl. ex Willd.	10	10, 20 (4), 30, 40	Ornduff 1968, 1969; Moore 1970, 1973, 1974
<i>A. tenellus</i> Trin.	10	10, 30	Ornduff 1967, 1981
<i>Aeluropus lagopoides</i> Trin. ex Thwaites	10	10 (2), 20, 25 (2)	Moore 1971, 1972; Goldblatt 1983; Goldblatt and Johnson 1998
<i>A. littoralis</i> Parl.	10	10 (5), 15	Federov 1969; Goldblatt and Johnson 1994, 1996
<i>A. littoralis</i> var. <i>intermedius</i> Coss. & Germ.	10	10?	Moore 1972
<i>Allolepis texana</i> (Vasey) Soderstr. & H.F. Decker	10	20 (2)	Ornduff 1968, 1969
<i>Astrebla lappacea</i> (Lindl.) Domin	10	20 (3)	Federov 1969
<i>A. pectinata</i> (Lindl.) Benth. (as <i>Astrebla pectinacea</i> F. Muell.)	10	20	Federov 1969
<i>Blepharidachne benthamiana</i> Hitc.	7	7 (2)	Federov 1969; Goldblatt 1988
<i>B. bigelovii</i> Hack.	7	7 (3)	Federov 1969; Goldblatt 1981, 1988
<i>B. kingii</i> Hack.	7	7 (2)	Goldblatt 1981, 1988
<i>Blepharoneuron tricholepis</i> (Torr.) Nash	8	8 (4)	Moore 1973; Goldblatt 1981; Goldblatt and Johnson 1991, 1994
<i>Bouteloua alamosana</i> Vasey ex Rose	10	30	Moore 1974
<i>B. americana</i> (L.) Scribn.	10	20	Moore 1973
<i>B. aristidoides</i> Griseb.	10	20 (2)	Federov 1969; Moore 1973
<i>B. barbata</i> Lag.	10	10 (5), 20 (2)	Ornduff 1967, 1968; Federov 1969; Moore 1973; Goldblatt 1981, 1988
<i>B. barbata</i> var. <i>major</i> (Vasey) R. Govaerts (as <i>Bouteloua</i> <i>rothrockii</i> Vasey)	10, 7	10, 11 (2), 20, 41/2?, 21, 45/2, 47/2, 49/2, 25	Federov 1969; Goldblatt 1981

<i>B. breviseta</i> Vasey	10, 7	10 (2), 21/2, 14, 20 (5)	Ornduff 1967, 1968; Federov 1969; Goldblatt 1981, 1983; Goldblatt and Johnson 1990
<i>B. chaseii</i> Swallen	10	20 (4), 30 (2)	Ornduff 1967, 1968, 1969; Moore 1970, 1973; Goldblatt and Johnson 1990
<i>B. chondrosioides</i> Benth. ex S. Watson	10, 7	7, 11, 10 (5), 20 (7)	Ornduff 1967, 1968, 1969; Federov 1969; Moore 1970, 1973; Goldblatt 1981
<i>B. curtispindula</i> Torr.	10, 7	10 (3), 14, 35/2, 20 (7), 41/2, 21 (2), 43/2, 45/2, 26, 28, 29, 59/2, 30, 61/2, 31, 63/2, 35, 37, 40, 41, 43, 48, 49, 62, 69/2-103/2, 85/2-101/2	Federov 1969; Goldblatt 1981, 1983
<i>B. curtispindula</i> var. <i>caespitosa</i> Gould & Kapadia	10	c.46	Moore 1973
<i>B. curtispindula</i> var. <i>tenuis</i> Gould & Kapadia	10	20, 41/2	Moore 1972
<i>B. dactyloides</i> (Nutt.) J.T. Columbus (as <i>Buchloë</i> <i>dactyloides</i> (Nutt.) Engelm.)	10, 7	10 (2), 20 (6), 28, 30 (7)	Ornduff 1967, 1968, 1969; Federov 1969; Moore 1970, 1973; Goldblatt 1983, 1988
<i>B. dimorpha</i> J.T. Columbus (as <i>Opizia stolonifera</i> J. Presl.)	10	20 (3), 43/2	Ornduff 1967, 1968, 1969; Federov 1969
<i>B. disticha</i> Benth.	10	10, 20	Moore 1973; Goldblatt 1988
<i>B. diversispicula</i> J.T. Columbus (as <i>Cathestecum brevifolium</i> Swallen)	10	30	Goldblatt 1988
<i>B. elata</i> C. Reeder	10	10, 20	Ornduff 1968, 1969
<i>B. eludens</i> Griffiths	10	20	Goldblatt and Johnson 1994
<i>B. eriopoda</i> Torr.	10, 7	10 (3), 21/2, 14 (2)	Federov 1969; Ornduff 1969; Moore 1970; Goldblatt 1981
<i>B. eriostachya</i> (Swallen) Reeder	10	30	Ornduff 1969

<i>B. gracilis</i> (Kunth) Lag. ex Griffiths	10, 7	10 (4), 21/2, 14, 35/2, 20 (3), 21 (3), 30 (3), 61/2, 77/2, 42	Ornduff 1967, 1968; Federov 1969; Moore 1970; Goldblatt 1981
<i>B. gracilis</i> (Kunth) Lag. ex Griffiths (as <i>Bouteloua oligostachya</i> Torr. ex A.Gray)	10	20	Federov 1969
<i>B. hirsuta</i> Lag.	10, 7	6, 10 (3), 21/2, 11, 12(2), 14 (3), 18, 37/2, 20 (5), 21 (3), 23 (3), 25, c.26, c.60	Ornduff 1967; Federov 1969; Moore 1970, 1972, 1974; Goldblatt 1988
<i>B. karwinskii</i> (E.Fourn.) Griffiths	10	10 (4)	Ornduff 1968, 1969; Moore 1970; Goldblatt and Johnson 1990
<i>B. longeseta</i> Gould	10	40	Moore 1971
<i>B. media</i> (E.Fourn.) Gould & Kapadia	10	10 (5), 27/2	Ornduff 1968; Federov 1969; Moore 1972, 1973
<i>B. parryi</i> (E.Fourn.) Griffiths var. <i>parryi</i> (as <i>Bouteloua parryi</i> (E.Fourn.) Griffiths)	10	10	Goldblatt 1988
<i>B. pectinata</i> Feath.	10	10 (5)	Federov 1969; Moore 1970, 1972, 1974
<i>B. purpurea</i> Gould & Kapadia	10	20 (3)	Ornduff 1968; Federov 1969
<i>B. radicata</i> Griffiths	10	10, 30	Ornduff 1967; Federov 1969
<i>B. ramosa</i> Scribn.	10	20 (2)	Goldblatt 1983, 1988
<i>B. reflexa</i> Swallen	10	10	Ornduff 1968
<i>B. repens</i> Scribn. & Merr.	10	10, 23/2, 20 (2), c.45/2, 23, 30 (3)	Moore 1971, 1972, 1974; Goldblatt 1988
<i>B. repens</i> Scribn. & Merr. (as <i>Bouteloua filiformis</i> Griffiths)	10, 7	7, 10 (2), 21/2, 11 (2), 20, c.45/2, 23, 30	Ornduff 1967; Federov 1969
<i>B. repens</i> Scribn. & Merr. (as <i>Bouteloua heterostega</i> Griffiths)	10	10, c.20	Federov 1969; Ornduff 1969

<i>B. rigidiseta</i> (Steud.) Hitchc.	10, 7	14, 35/2, 20 (2)	Federov 1969; Moore 1970
<i>B. scorpioides</i> Lag.	10	10 (2), 20 (2)	Ornduff 1968, 1969; Moore 1970; Goldblatt 1988
<i>B. simplex</i> Lag.	10	10, 20 (3)	Ornduff 1967; Federov 1969
<i>B. triaena</i> Scribn.	10	10 (4)	Federov 1969; Moore 1972
<i>B. trifida</i> Thurb.	10, 7	10 (5), 14	Ornduff 1967, 1968; Moore 1970, 1973
<i>B. uniflora</i> Vasey	10	10 (4)	Federov 1969
<i>B. warnockii</i> Gould & Kapadia	10, 7	21/2, 11 (2), 23/2, 12, 25/2, 14, 29/2-39/2, 20	Federov 1969
<i>B. williamsii</i> Swallen	10	10 (2), 20	Moore 1971; Moore 1973; Goldblatt 1981
<i>Buchlomimus nervatus</i> (Swallen) Reeder, C.Reeder & Rzed.	10	20 (2)	Ornduff 1967, 1969
<i>Calamovilfa gigantea</i> Scribn. & Merr.	10	20	Ornduff 1969
<i>C. longifolia</i> Hack.	10	20 (2)	Goldblatt 1981; Goldblatt 1983
<i>C. longifolia</i> var. <i>magna</i> Scribn. & Merr.	10	20	Ornduff 1969
<i>Cathestecum annuum</i> Swallen	10	10 (2)	Ornduff 1967, 1968
<i>C. erectum</i> Vasey & Hack.	10	10 (3), 15, 20, 30 (2)	Ornduff 1967, 1969; Moore 1970, 1973; Goldblatt 1981
<i>C. multifidum</i> Griffiths	10	10 (2)	Ornduff 1967, 1969
<i>Chaboissaea atacamensis</i> (Parodi) P.M.Peterson & Annable	8	8	Goldblatt and Johnson 1996
<i>C. ligulata</i> E.Fourn.	8	8 (2)	Ornduff 1969; Moore 1970
<i>Chloris acuminata</i> Trin.	10	20	Federov 1969
<i>C. andropogonoides</i> E.Fourn.	10	20 (3)	Federov 1969
<i>C. barbata</i> Sw.	10	10 (4), 20 (10), 25 (2)	Ornduff 1968, 1969; Federov 1969; Moore 1971, 1972, 1974, 1977; Goldblatt 1981, 1985; Goldblatt and Johnson 1991, 1998
<i>C. barbata</i> Sw. (as <i>Chloris inflata</i> Link)	10	20 (3), 28	Moore 1972, 1974, 1977; Goldblatt 1983
<i>C. berroi</i> Arechav.	10	20	Federov 1969
<i>C. bourneii</i> Rang. & Tadul.	10	20, 25, 30	Federov 1969; Goldblatt and Johnson 1991
<i>C. caudata</i> Trin. ex Bunge	10	20	Federov 1969

<i>C. chloridea</i> (C.Presl.) Hitchc.	10	20 (2)	Moore 1972; Goldblatt 1988
<i>C. chloroides</i> (J.Presl.) Hitchc.	10	40	Ornduff 1967
<i>C. ciliata</i> Sw.	10	20 (7)	Ornduff 1968, 1969; Federov 1969; Moore 1970
<i>C. cucullata</i> Bisch.	10	20 (5)	Ornduff 1968; Federov 1969; Moore 1970
<i>C. dandyana</i> C.D.Adams (as <i>Cynodon polydactyla</i> Sw.)	10	36	Federov 1969
<i>C. distichophylla</i> Lag.	10	10 (2), 20 (2)	Ornduff 1967; Federov 1969
<i>C. dolichostachya</i> Lag.	10	10, 20 (7), 30 (2)	Moore 1972, 1977; Goldblatt 1981, 1985; Goldblatt and Johnson 1990
<i>C. ekmanii</i> Hitchc.	10	20	Moore 1974
<i>C. formosana</i> (Honda) Keng	10	20	Moore 1973
<i>C. gayana</i> Kunth	10	10 (10), 15, 20 (8)	Ornduff 1967, 1968; Federov 1969; Moore 1970, 1977; Goldblatt 1983; Goldblatt and Johnson 1990
<i>C. halophila</i> Parodi	10	40	Federov 1969
<i>C. incompleta</i> Roth ex Roem. & Schult.	10	20	Moore 1977
<i>C. mollis</i> (Nees) Swallen	10	20	Moore 1977
<i>C. montana</i> Roxb.	10	20 (3)	Goldblatt 1988; Goldblatt and Johnson 1990, 1991
<i>C. petraea</i> Sw.	10	20 (4)	Federov 1969; Moore 1970, 1973
<i>C. pilosa</i> Schumach.	10	10 (2), 15	Federov 1969; Moore 1977; Goldblatt 1981
<i>C. prieurii</i> Kunth	10	20	Moore 1974
<i>C. pycnothrix</i> Trin.	10	18, 20 (3)	Ornduff 1967; Federov 1969; Goldblatt 1981, 1983
<i>C. radiata</i> (L.) Sw.	10	20 (4)	Ornduff 1968; Federov 1969; Moore 1973
<i>C. radiata</i> (L.) Sw. (as <i>Chloris</i> <i>gracilis</i> P.Durand)	10	15	Federov 1969
<i>C. roxburghiana</i> Schult.	10	10 (2)	Federov 1969; Goldblatt 1983
<i>C. rufescens</i> Lag.	10	50 (4)	Ornduff 1967, 1968; Federov 1969; Moore 1972
<i>C. subdolichostachya</i> N.J.C.Muell.	10	20, c.33	Federov 1969
<i>C. subdolichostachya</i> N.J.C.Muell. (as <i>Chloris</i> <i>latisquamea</i> Nash)	10	20, c.28, c.31, c.63/2, c.32, c.36, c.42	Federov 1969; Moore 1970

<i>C. submutica</i> Kunth	10	c.65/2, 40 (7)	Ornduff 1967, 1968; Federov 1969; Moore 1973
<i>C. truncata</i> R.Br.	10	20 (2)	Federov 1969
<i>C. verticillata</i> Nutt.	10	20 (3), c.28, 63/2	Federov 1969; Moore 1970; Goldblatt 1981
<i>C. villosa</i> Pers.	10	10	Goldblatt 1981
<i>C. virgata</i> Sw.	10	7, 10 (18), 13, 15, 18	Ornduff 1968; Federov 1969; Moore 1971, 1972, 1973, 1974; Goldblatt 1983; Goldblatt and Johnson 1990, 1998, 2000
<i>Cottea pappophoroides</i> Kunth	10	10 (4)	Federov 1969; Moore 1970
<i>Crypsis aculeata</i> Aiton	8, 9	8 (2), 9 (4), 27	Moore 1977; Federov 1969; Goldblatt 1981; Goldblatt and Johnson 1996
<i>C. alopecuroides</i> Schrad.	8, 9	8, 9 (2)	Federov 1969; Goldblatt 1983; Goldblatt and Johnson 1994
<i>C. schoenoides</i> (L.) Lam.	8, 9	16, 18 (2)	Moore 1971; Goldblatt 1981, 1983
<i>C. vaginiflora</i> Opiz	8	24	Goldblatt 1983
<i>Ctenium canescens</i> Benth.	9	9	Moore 1977
<i>C. elegans</i> Kunth	9	18	Moore 1977
<i>C. newtonii</i> Hack.	9	12, 18	Moore 1977; Goldblatt 1981
<i>Cynodon aethiopicus</i> Clayton & J.R.Harlan	10, 9	9-18	Moore 1973
<i>C. alterniflorus</i>	10	21	Federov 1969
<i>C. barberi</i> Rang. & Tadul.	9	9 (3)	Moore 1977; Goldblatt 1981
<i>C. dactylon</i> (L.) Pers.	10, 9	9 (9), 9+0-3B (3), 27/2 (6), 13+I, 15 (2), 18 (30), 18+0-2B (3), 20 (6), 27 (3)	Ornduff 1968, 1969; Federov 1969; Moore 1970, 1972, 1977; Goldblatt 1981, 1983, 1985; Goldblatt and Johnson 1990, 1994, 1996, 1998
<i>C. dactylon</i> var. <i>afghanicus</i> J.R.Harlan & DeWet	9	9 (2), 18 (2)	Moore 1972, 1974
<i>C. dactylon</i> var. <i>aridus</i> J.R.Harlan & DeWet	9	9 (3)	Moore 1972, 1974; Goldblatt 1985
<i>C. dactylon</i> var. <i>coursii</i> (A.Camus) J.R.Harlan & DeWet	9	18	Moore 1972
<i>C. dactylon</i> var. <i>dactylon</i>	9	9 (2)	Moore 1972; Goldblatt and Johnson 1994
<i>C. dactylon</i> var. <i>elegans</i> Rendle	9	18	Moore 1972



<i>C. dactylon</i> var. <i>polevansii</i> (Stent) J.R.Harlan & DeWet	9	18	Moore 1972
<i>C. dactylon</i> var. <i>villosus</i> Regel	9	9	Goldblatt and Johnson 1994
<i>C. diploideus</i>	9	9	Federov 1969
<i>C. incompletes</i> Nees var. <i>hirsutus</i> (Stent) DeWet (as <i>Cynodon hirsutus</i> Stent)	9	9	Federov 1969
<i>C. michauxioides</i>	9	14	Federov 1969
<i>C. nlemfuensis</i> Vanderyst var. <i>nlemfuensis</i> Clayton & J.R.Harlan	10, 9	9-18	Moore 1973
<i>C. nlemfuensis</i> var. <i>robustus</i> Clayton & J.R.Harlan	9	18	Goldblatt and Johnson 1990
<i>C. plectostachyum</i> Pilg.	9	9	Federov 1969
<i>C. radiatus</i> Roth (as <i>Cynodon</i> <i>arcautus</i> J. & C.Presl.)	10, 9	9, 10, 16, 18 (5)	Federov 1969; Moore 1977; Goldblatt and Johnson 1990
<i>C. transvaalensis</i> Burt Davy	9	9	Moore 1973
<i>Dactyloctenium aegyptium</i> (L.) Willd.	10, 6	9 (3), 9+, 10 (6), 11, 11+, 23/2, 27/2, 17, 18 (3), 19 (3), 20 (11), 21 (2), 22 (3), 23 (9), 24 (7), 26	Ornduff 1967, 1968; Federov 1969; Moore 1971, 1972, 1973, 1974, 1977; Goldblatt 1981, 1983, 1985, 1988; Goldblatt and Johnson 1990, 1991, 1994, 1998
<i>D. aegyptium</i> (L.) Willd. (as <i>Eleusine aegyptica</i> (L.) Roxb.)	9	17, 18 (2), 45/2 (2)	Federov 1969
<i>D. aristatum</i> Link	10	10 (3), 20 (3)	Goldblatt 1981, 1983
<i>D. scindicum</i> Boiss.	10, 6	9, 10 (3), 24	Federov 1969; Goldblatt 1981, 1983; Goldblatt and Johnson 1990
<i>Desmostachya bipinnata</i> Stapf	10	10 (5), 20	Moore 1977; Goldblatt 1981, 1985; Goldblatt and Johnson 1990
<i>Dinebra arabica</i> Jacq.	10	10	Federov 1969
<i>D. retroflexa</i> Panz.	10	10 (4), 20, 30	Federov 1969; Goldblatt 1981; Goldblatt and Johnson 1990, 1998
<i>Diplachne bulgarica</i> Bornm.	10	20 (2)	Moore 1977
<i>D. fusca</i> (L.) Roem. & Schult.	10	10, 20 (2)	Federov 1969; Goldblatt and Johnson 1990, 1998

<i>D. serotina</i> Link	10	20 (4)	Ornduff 1968; Federov 1969; Moore 1977
<i>D. serotina</i> var. <i>sinensis</i> Hance	10	20	Ornduff 1968
<i>Distichilis palmeri</i> (Vasey) Fassett ex I.M.Johnst.	10	20	Ornduff 1968
<i>D. spicata</i> (L.) Greene var. <i>borealis</i> (J.Presl.) Beetle	10	20	Moore 1977
<i>D. spicata</i> var. <i>spicata</i> (as <i>Distichilis spicata</i> (L.) Greene)	10	19, 20 (7), 21, 36	Federov 1969; Ornduff 1969; Moore 1973; Goldblatt 1981
<i>D. stricta</i> Rydb.	10	20 (4), c.36	Federov 1969; Ornduff 1969; Goldblatt 1983
<i>D. texana</i> (Vasey) Scribn.	10	20	Federov 1969
<i>Eleusine brevifolia</i> R.Br.	9	18	Federov 1969
<i>E. compressa</i> (Forssk.) Asch. & Sweinf. ex. C. Chr.	10, 9	9, 20, 45/2	Federov 1969; Goldblatt 1985; Goldblatt and Johnson 1990
<i>E. coracana</i> (L.) Gaertn.	9	18 (24), 18+1-2B	Ornduff 1968; Federov 1969; Moore 1971, 1974, 1977; Goldblatt 1981, 1985; Goldblatt and Johnson 1990, 1994, 2000
<i>E. coracana</i> subsp. <i>africana</i> (Kenn.-O'Byrne) Hilu & DeWet (as <i>Eleusine africana</i> Kenn.- O'Byrne)	9	18 (11)	Ornduff 1967, 1968; Federov 1969; Moore 1977; Goldblatt 1985; Goldblatt and Johnson 1990, 1994, 2000
<i>E. coracana</i> subsp. <i>africana</i> (Kenn.-O'Byrne) Hilu & DeWet (as <i>Eleusine indica</i> subsp. <i>africana</i> (Kenn.-O'Byrne) S.M.Phillips)	9	9, 18 (2)	Goldblatt 1981, 1985; Goldblatt and Johnson 1990
<i>E. flagellifera</i> Nees	9	45/2	Federov 1969
<i>E. floccifolia</i> Spreng.	9	9 (5), 18	Moore 1977; Goldblatt 1985; Goldblatt and Johnson 1994, 1998, 2000
<i>E. indica</i> (L.) Gaertn.	9	8, 9 (51), 18 (4), 27 (2)	Ornduff 1967, 1968, 1969; Federov 1969; Moore 1971, 1972, 1974, 1977; Goldblatt 1981, 1983, 1985; Goldblatt and Johnson 1990, 1991, 1994, 1998, 2000
<i>E. indica</i> subsp. <i>indica</i>	9	9	Goldblatt 1985
<i>E. intermedia</i> (Chiov.) S.M.Phillips	9	9 (2)	Goldblatt and Johnson 1994, 1998
<i>E. jaegeri</i> Pilg.	10	10 (3)	Goldblatt 1985; Goldblatt and Johnson 1994, 2000
<i>E. kigeziensis</i> S.M.Phillips	9	19	Goldblatt and Johnson 1994

<i>E. lagopoides</i> Merr.	9	18 (2)	Federov 1969
<i>E. multiflora</i> Hochst.	9	8 (4), 9	Goldblatt 1985, 1988; Goldblatt and Johnson 1994, 1998, 2000
<i>E. oligostachya</i> Link	9	9 (2)	Federov 1969
<i>E. reniformis</i> Divak.	9	18, 21	Goldblatt 1985; Goldblatt and Johnson 1990
<i>E. tocussa</i> Fresen.	9	39/2	Federov 1969
<i>E. tristachya</i> (Lam.) Lam.	9	8, 9 (12)	Ornduff 1968; Federov 1969; Moore 1970, 1977; Goldblatt 1981, 1985, Goldblatt and Johnson 1994, 1998, 2000
<i>E. verticillata</i> Roxb.	9	9 (3), 18	Federov 1969; Goldblatt and Johnson 1994, 1998
<i>Enneapogon borealis</i> (Griseb.) Honda	10	10	Goldblatt and Johnson 1998
<i>E. desvauxii</i> P. Beauv.	10	10 (3)	Ornduff 1968, 1969; Federov 1969
<i>E. elegans</i> Stapf	10	10 (3)	Federov 1969; Moore 1977; Goldblatt 1981
<i>E. persicus</i> Boiss.	10	10	Goldblatt 1981
<i>E. wrightii</i> (S. Watson) C.E. Hubb.	10	10	Federov 1969
<i>Enteropogon gracilior</i> Rendle	10	20	Federov 1969
<i>E. monostachyus</i> K. Schum.	10	10 (2)	Federov 1969; Goldblatt 1981
<i>Eragrostiella bifaria</i> (Vahl) Bor	10,9	9, 20	Moore 1977; Goldblatt 1981
<i>E. brachyphylla</i> (Stapf) Bor	9	18	Federov 1969
<i>E. leioptera</i> (Stapf) Bor	10	7, 10 (3)	Goldblatt 1981, 1985
<i>Eragrostis acutiflora</i> Nees	10	20 (2)	Moore 1972, 1977
<i>E. airoides</i> Nees	10	18	Moore 1977
<i>E. albida</i> Hitchc.	10	20	Federov 1969
<i>E. amabilis</i> Wight & Arn. (as <i>Eragrostis tenella</i> (L.) Roem. & Schult.)	10	9, 10 (9), 20, 30	Ornduff 1967; Federov 1969; Moore 1971, 1972, 1977; Goldblatt 1981, 1983; Goldblatt and Johnson 1990
<i>E. amurensis</i> Prob.	10	20	Goldblatt 1988
<i>E. aquatica</i> Honda	10	30	Ornduff 1969
<i>E. atrovirens</i> (Desf.) Trin.	10	10 (3), 20 (2), 30 (8)	Federov 1969; Moore 1977; Goldblatt 1981; Goldblatt and Johnson 1990, 1991
<i>E. barrelieri</i> Daveau	10	30 (4)	Ornduff 1968, 1969; Federov 1969; Goldblatt and Johnson 1994
<i>E. barteri</i> C.E. Hubb.	10	10	Goldblatt 1981
<i>E. beyrichii</i> J.G. Sm.	10	20	Federov 1969
<i>E. bifaria</i> (Vahl) Steud.	10	20	Federov 1969
<i>E. boriiana</i> Launert	10	30	Moore 1973

<i>E. brownii</i> (Kunth) Nees	10	20	Federov 1969
<i>E. bulbifera</i> Steud.	10	20	Federov 1969
<i>E. burmanica</i> Bor	10	20	Federov 1969
<i>E. cambessediana</i> Steud.	10	10	Federov 1969
<i>E. capensis</i> (Thunb.) Trin.	10	20	Federov 1969
<i>E. capillaris</i> Nees	10	50	Federov 1969
<i>E. chapellieri</i> (Kunth) Nees	10	10 (4)	Ornduff 1967; Goldblatt 1981, 1983
<i>E. cilianensis</i> (All.) Vignolo ex Janch.	10	10 (15), 20 (4)	Ornduff 1968; Federov 1969; Moore 1972, 1973; Goldblatt 1983, 1988; Goldblatt and Johnson 1990, 1991, 1994, 1996, 1998, 2000
<i>E. cilianensis</i> (All.) Vignolo ex Janch. (as <i>Eragrostis megastachya</i> Link)	10	10 (3), 20	Federov 1969
<i>E. curtipendicellata</i> Buckley	10	20 (2)	Federov 1969; Moore 1970
<i>E. curvula</i> (Schrad.) Nees	10	20 (3), 27 (2)	Goldblatt and Johnson 1990, 1991, 1994
<i>E. decannensis</i> Bor	10	10	Goldblatt 1988
<i>E. diarrhena</i> Steud.	10	10 (3), 20 (2), 30 (2)	Ornduff 1967; Goldblatt and Johnson 1990, 1991
<i>E. diplachnoides</i> Steud.	10	10 (2)	Moore 1977
<i>E. elongata</i> (Willd.) Jacq.	10	20	Federov 1969
<i>E. ferruginea</i> P. Beauv.	10	40 (3)	Federov 1969; Ornduff 1969
<i>E. frankii</i> Steud.	10	20, 40	Goldblatt 1981
<i>E. gangetica</i> Steud.	10	10, 40	Ornduff 1967; Goldblatt 1983
<i>E. gangetica</i> Steud. (as <i>Eragrostis stenophylla</i> Hochst. ex Miq.)	10	30	Goldblatt and Johnson 1990
<i>E. glutinosa</i> Trin.	10	30	Moore 1970
<i>E. grandis</i> Hillebr.	10	22	Federov 1969
<i>E. guianensis</i> Hitchc.	10	10	Moore 1977
<i>E. hirsuta</i> Nash	10	50	Federov 1969
<i>E. holacantha</i> Torr.	10	15	Goldblatt 1988
<i>E. hondurensis</i> R.W. Pohl	10	c.20, 15	Goldblatt 1983
<i>E. hypnoides</i> (Lam.) B.S.P.	10	10 (2), 20	Moore 1974; Goldblatt 1983
<i>E. intermedia</i> Hitchc.	10	30, 36, c.37, 38, 40 (2), 50, 60 (2), c.104	Moore 1970, 1973

<i>E. japonica</i> (Thunb.) Trin.	10	10 (6), 20 (3), 30	Federov 1969; Moore 1977; Goldblatt 1981, 1985; Goldblatt and Johnson 1990, 1991, 1998
<i>E. japonica</i> (Thunb.) Trin. (as <i>Eragrostis glomerata</i> (Walter) L.H.Dewey)	10	10 (2)	Ornduff 1969; Moore 1973
<i>E. lehmanniana</i> Nees	10	20, 30	Federov 1969
<i>E. lugens</i> Nees	10	20, 40, c.54	Federov 1969
<i>E. macilenta</i> Steud.	10	20	Ornduff 1967
<i>E. maderaspatana</i> Bor	10	30	Goldblatt 1985
<i>E. maypurensis</i> Steud.	10	10 (3), 20	Ornduff 1969; Moore 1972, 1973, 1977
<i>E. megalosperma</i> F.Muell. ex Benth.	10	10 (2)	Ornduff 1968; Federov 1969
<i>E. mexicana</i> (Hornem.) Link	10	30 (5)	Ornduff 1968; Federov 1969; Moore 1973; Goldblatt 1981
<i>E. mildbraedii</i> Pilg.	10	10	Ornduff 1967
<i>E. minor</i> Host	10	10 (2), 15, 20 (8), 30 (3)	Federov 1969; Goldblatt 1981, 1983, 1988; Goldblatt and Johnson 1990, 1991, 1994, 1996, 1998, 2000
<i>E. minor</i> Host (as <i>Eragrostis</i> <i>poaeoides</i> P.Beauv.)	10	15, 20 (5), 22, 30	Federov 1969; Moore 1972, 1977; Goldblatt 1981, 1983; Goldblatt and Johnson 1990
<i>E. minor</i> Host (as <i>Eragrostis</i> <i>suaveolens</i> A.K.Becker ex Claus)	10	15	Goldblatt 1983
<i>E. mexicana</i> subsp. <i>virescens</i> (J.Presl) S.D.Koch & Sánchez Vega (as <i>Eragrostis virescens</i> J. & C.Presl.)	10	30	Goldblatt and Johnson 1994
<i>E. montufari</i> Steud.	10	35	Federov 1969
<i>E. multicaulis</i> Steud.	10	20 (3)	Federov 1969; Ornduff 1969
<i>E. namaquensis</i> Nees	10	10 (2)	Ornduff 1967; Goldblatt 1983
<i>E. neomexicana</i> Vasey	10	28 (3), 30 (4)	Ornduff 1968; Federov 1969; Moore 1970, 1973; Goldblatt 1983; Goldblatt and Johnson 1991
<i>E. nigra</i> Nees ex Steud.	10	30 (2)	Goldblatt 1981, 1985

<i>E. nutans</i> Nees ex Steud.	10	20 (2), 21+f, 27+1, 30 (3)	Federov 1969; Moore 1977; Goldblatt 1981, 1985; Goldblatt and Johnson 1990, 1991
<i>E. nutans</i> Nees ex Steud. (as <i>Eragrostis chariis</i> (Schult.) Hitchc.)	10	20	Federov 1969
<i>E. obtusiflora</i> (E.Fourn.) Scribn.	10	20	Goldblatt 1981
<i>E. olivacea</i> K.Schumm. ex Engl. (as <i>Eragrostis lasiantha</i> Stapf)	10	10	Ornduff 1967
<i>E. pallescens</i> Hitchc.	10	40	Federov 1969
<i>E. palmeri</i> S.Watson	10	20	Moore 1973
<i>E. paniciformis</i> (A.Braun) Steud.	10	30	Ornduff 1967
<i>E. papposa</i> Steud.	10	10	Moore 1972
<i>E. patens</i> Oliv.	10	10 (2)	Goldblatt 1983
<i>E. pectinacea</i> Nees	10	20, 30 (5), 40	Ornduff 1968; Federov 1969; Goldblatt 1981, 1983
<i>E. pectinacea</i> Nees (as <i>Eragrostis diffusa</i> Buckley)	10	30 (3)	Ornduff 1967; Moore 1970, 1973
<i>E. pilosa</i> (L.) P.Beauv.	10	10 (3), 15, 35/2, 18 (3), 20 (8), 25 (4), 30 (1), 36	Ornduff 1967; Federov 1969; Moore 1971, 1972, 1977; Goldblatt 1981, 1983, 1988; Goldblatt and Johnson 1990, 1991, 1994, 1998
<i>E. pilosa</i> var. <i>pilosa</i>	10	20	Goldblatt 1981
<i>E. pilosa</i> subsp. <i>pilosa</i>	10	20	Goldblatt 1981
<i>E. plumbea</i> Scribn. ex Beal	10	20, 30	Ornduff 1967
<i>E. polytricha</i> Nees	10	c.30	Ornduff 1969
<i>E. polytricha</i> Nees (as <i>Eragrostis</i> <i>trichocolea</i> Arechav.)	10	30, 40	Moore 1972, 1974
<i>E. prolifera</i> Steud.	10	10, 20	Moore 1974; Goldblatt 1981
<i>E. prolifera</i> Steud. (as <i>Eragrostis</i> <i>domingensis</i> Steud.)	10	20	Moore 1972
<i>E. racemosa</i> (Thunb.) Steud.	10	20	Ornduff 1967
<i>E. secundiflora</i> J. & C.Presl.	10	20	Federov 1969
<i>E. secundiflora</i> subsp. <i>oxylepis</i> (Torr.) S.D.Koch (as <i>Eragrostis</i> <i>oxylepis</i> (Torr.) Torr.)	10	20	Moore 1970
<i>E. sessilispica</i> Buckley	10	20 (3)	Ornduff 1967; Federov 1969; Moore 1970
<i>E. simpliciflora</i> Steud.	10	20	Moore 1973
<i>E. spectabilis</i> Steud.	10	10, 20, 21	Federov 1969; Goldblatt 1985

<i>E. spicata</i> Vasey	10	20	Federov 1969
<i>E. squamata</i> Steud.	10	20	Goldblatt 1981
<i>E. superba</i> Peyr.	10	10, 20 (4)	Goldblatt 1981, 1983
<i>E. swallenii</i> Hitchc.	10	42	Moore 1970
<i>E. tef</i> (Zucc.) Trotter	10	20 (3)	Federov 1969
<i>E. tenella</i> (L.) P.Beauv. var. <i>insularis</i> C.E.Hubb.	10	10, 20, 30	Moore 1977; Goldblatt and Johnson 1991
<i>E. tenella</i> var. <i>tenella</i>	10	10 (2), 20, 30	Moore 1977; Goldblatt and Johnson 1991
<i>E. tenuifolia</i> (A.Rich.) Steud.	10	20 (6)	Ornduff 1967; Moore 1973, 1977; Goldblatt 1983; Goldblatt and Johnson 1996
<i>E. tephrosantos</i> Schult.	10	30	Goldblatt 1981
<i>E. tremula</i> Hochst.	10	10 (4), 14 (2), 14+, 20 (2)	Ornduff 1967; Federov 1969; Moore 1977; Goldblatt 1983, 1988; Goldblatt and Johnson 1990, 1991
<i>E. trichodes</i> Wood	10	20	Goldblatt 1981
<i>E. uniolooides</i> Nees ex Steud.	10	9 (2), 10 (6), c.15, 30 (2)	Federov 1969; Moore 1977; Goldblatt 1981, 1985; Goldblatt and Johnson 1990, 1991
<i>E. usambarensis</i> Napper	10	40	Ornduff 1967
<i>E. variabilis</i> Gaudich.	10	20	Goldblatt 1981
<i>E. viscosa</i> Trin.	10	8, 20, 30 (4)	Federov 1969; Moore 1972, 1977; Goldblatt 1988; Goldblatt and Johnson 1994
<i>E. zeylanica</i> Nees & Meyen	10	18, 20, 30, c.40	Federov 1969
<i>Erioneuron avenaceum</i> (Kunth) Tateoka	8	8 (2), 16	Moore 1970; Goldblatt 1988
<i>E. grandiflorum</i> (Vasey) Tateoka	8	8 (2), 16 (2)	Moore 1970, 1973; Goldblatt 1981
<i>E. nealleyi</i> (Vasey) Tateoka	8	8	Ornduff 1968
<i>E. pilosum</i> (Buckley) Nash	8	8 (4), 16	Ornduff 1968; Moore 1970, 1973; Goldblatt 1981
<i>E. pulchellum</i> (Kunth) Tateoka	8	8 (3), 16	Moore 1970, 1973; Goldblatt 1981
<i>Eustachys petraea</i> (Sw.) Desv.	10	20 (3)	Ornduff 1968; Goldblatt 1981
<i>E. uliginosa</i> (Hack.) Herter (as <i>Chloris uliginosa</i> Hack.)	10	20	Federov 1969
<i>Gouinia gautemalensis</i> (Hack.) Swallen	10	38	Moore 1974
<i>G. latifolia</i> Vasey	10	20 (2)	Federov 1969; Goldblatt 1981

<i>G. longiramea</i> Swallen (as <i>Gouinia virgata</i> (J.Presl.) Scribn.)	10	20 (4)	Federov 1969; Moore 1970, 1973; Goldblatt and Johnson 1998
<i>Griffithsochloa multifida</i> (Griffiths) G.J.Pierce	10	10	Goldblatt 1981
<i>Gymnopogon ambiguus</i> (Michx.) B.S.P.	10	20	Moore 1970
<i>G. fastigiatus</i> Nees	10	20 (2)	Moore 1973; Goldblatt 1981
<i>G. foliosus</i> Nees	10	10	Goldblatt 1981
<i>G. spicatus</i> (Spreng.) Kuntze	10	10	Goldblatt 1981
<i>Harpachne schimperii</i> Hochst.	10	20	Ornduff 1967
<i>Hilaria annua</i> Reeder & C.Reeder	9	18	Goldblatt and Johnson 1991
<i>H. belangeri</i> (Steud.) Nash	9	18 (3), 36 (3), 37 (3), 38, 41, 45	Ornduff 1967; Federov 1969; Moore 1970
<i>H. cencroides</i> Kunth	9	22, 36 (4), 37	Ornduff 1967, 1968, 1969; Federov 1969; Moore 1973
<i>H. ciliata</i> Nash	9	36 (4), 37	Ornduff 1967, 1968; Moore 1970; Goldblatt 1988
<i>H. jamesii</i> Benth.	9	9, 18, 19, 36	Ornduff 1967; Goldblatt 1981, 1985
<i>H. mutica</i> Benth.	9	18 (3), 19, c.27 (2), 36 (2), c.90	Federov 1969; Moore 1970, 1973; Goldblatt 1981
<i>H. rigida</i> Scribn.	9	18, 54	Goldblatt 1981, 1988
<i>H. swallenii</i> Cory	9	27, 36, 45, c.60	Federov 1969; Goldblatt 1988
<i>Jouvea pilosa</i> Scribn.	10	10 (4)	Ornduff 1969; Moore 1973, 1974
<i>J. straminea</i> E.Fourn.	10	10 (2)	Ornduff 1969; Moore 1973
<i>Kengia hackellii</i> (Honda) Packer	10	20	Ornduff 1969
<i>K. serotina</i> (L.) Packer	10	20	Federov 1969
<i>Leptochloa caerulescens</i> Steud.	10	10 (3)	Moore 1977; Goldblatt 1981
<i>L. capillacea</i> P.Beauv.	10	10	Federov 1969
<i>L. chinensis</i> Nees	10	10, 18, 20 (7), 27	Federov 1969; Moore 1977; Goldblatt 1981, 1985; Goldblatt and Johnson 1990, 1996
<i>L. cognatum</i> (Schult.) Chase	10	18, 36	Moore 1970
<i>L. dubia</i> Nees	10	20 (3), 30 (2), 40 (2)	Ornduff 1967, 1968; Federov 1969; Moore 1970; Goldblatt 1988



<i>L. dubia</i> Nees (as <i>Diplachne dubia</i> Scribn.)	10	20	Federov 1969
<i>L. fusca</i> (L.) Kunth subsp. <i>fascicularis</i> (Lam.) N.Snow (as <i>Leptochloa fascicularis</i> A.Gray)	10	10, 20	Ornduff 1968; Moore 1973
<i>L. fusca</i> subsp. <i>uninervia</i> (J.Presl) N.Snow (as <i>Leptochloa uninervia</i> (J.Presl.) Hitchc. & Chase)	10	10 (2)	Federov 1969; Moore 1970
<i>L. nealleyi</i> Vasey	10	20	Federov 1969
<i>L. neesii</i> (Thwaites) Benth.	10	20	Moore 1977
<i>L. obtusiflora</i> Trin. ex Steud.	10	10	Federov 1969
<i>L. panicea</i> (Retz.) Ohwi	10	10 (3), 10+1B, 20	Federov 1969; Goldblatt 1983, 1988; Goldblatt and Johnson 1990
<i>L. panicea</i> subsp. <i>brachiata</i> (Steud.) N.Snow (as <i>Leptochloa filiformis</i> (Lam.) P.Beauv.)	10	10 (4)	Federov 1969; Moore 1970; Goldblatt 1981; Goldblatt and Johnson 1990
<i>L. panicoides</i> (J.Presl.) Hitchc.	10	10	Goldblatt 1985
<i>L. phleoides</i> (Vill.) Rchb.	10	6, 10	Moore 1972; Goldblatt and Johnson 1990
<i>L. polystachya</i> Benth.	10	10	Federov 1969
<i>L. scabra</i> Nees	10	30 (2)	Moore 1974, 1977
<i>L. uniflora</i> Hochst. ex A.Rich.	10	10, 18, 20	Moore 1977; Goldblatt 1981
<i>L. virgata</i> (L.) P.Beauv.	10	20 (4)	Federov 1969; Ornduff 1969; Moore 1973
<i>L. viscida</i> (Scribn.) Beal	10	20	Moore 1973
<i>Leptochloopsis virgata</i> (Poir.) Yates	10	20	Moore 1974
<i>Lepturus cylindricus</i> (Willd.) Trin.	7	13, 26	Federov 1969
<i>L. filiformis</i> Trin.	7, 9	7 (2), 18	Federov 1969
<i>L. incurvatus</i> (L.f.) Trin.	7, 9	7, 18 (3)	Federov 1969; Goldblatt 1981
<i>L. pannonicus</i> Kunth	7	7 (2)	Federov 1969
<i>L. radicans</i> A.Camus	9	9 (2)	Federov 1969; Moore 1977
<i>L. repens</i> (G.Forst.) R.Br.	9	27 (2)	Federov 1969; Moore 1973
<i>Lycurus phleoides</i> Kunth	10	14, 20 (6)	Ornduff 1967, 1969; Federov 1969; Moore 1973; Goldblatt 1981
<i>Microchloa kunthii</i> Desv.	10, 6	12 (3), 20 (2), 22	Ornduff 1967, 1969; Moore 1974; Goldblatt 1981, 1985
<i>M. indica</i> (L.f.) P.Beauv.	6	6 (2), 12, c.18, 24	Federov 1969; Moore 1977; Goldblatt and Johnson 1998
<i>Monanthochloë littoralis</i> Engelm.	10	20 (2)	Ornduff 1969; Goldblatt 1988

<i>Mosdenia phleoides</i> Stent	10	20	Federov 1969
<i>Muhlenbergia aguascalientensis</i> Y.Arrieta & Lacerda-Lemus	10	10	Goldblatt and Johnson 2000
<i>M. andina</i> Hitchc.	10	5, 10 (2)	Ornduff 1967; Federov 1969; Moore 1971
<i>M. annua</i> (Vasey) Swallen	10	10 (2)	Goldblatt and Johnson 1991, 1994
<i>M. arizonica</i> Scribn.	10	10	Moore 1970
<i>M. articulata</i> Scribn.	10	20	Moore 1970
<i>M. asperifolia</i> (Trin.) Parodi	10	10 (3), 11, 14	Ornduff 1967; Federov 1969; Moore 1970; Goldblatt 1983
<i>M. biloba</i> Hitchc.	10	8	Goldblatt and Johnson 1991
<i>M. brachyphylla</i> Bush	10	20 (2)	Ornduff 1967; Federov 1969
<i>M. brevis</i> C.O.Goodd.	10	10 (4)	Moore 1970; Goldblatt and Johnson 1991, 1994
<i>M. bushii</i> R.W.Pohl	10	20	Moore 1971
<i>M. californica</i> Vasey	10	40	Ornduff 1967
<i>M. ciliata</i> (Kunth) Trin.	10, 9	9 (2), 10 (3)	Moore 1970; Goldblatt and Johnson 1991, 1994
<i>M. confusa</i> (E.Fourn.) Swallen	10	30	Moore 1970
<i>M. crispiseta</i> Hitchc.	10	10 (3)	Moore 1970; Goldblatt and Johnson 1991, 1994
<i>M. curviaristata</i> (Ohwi) Ohwi	10	20	Goldblatt 1981
<i>M. curviaristata</i> var. <i>curviaristata</i>	10	20	Goldblatt and Johnson 1991
<i>M. curviaristata</i> var. <i>nipponica</i> Ohwi	10	20	Goldblatt and Johnson 1991
<i>M. cuspidata</i> Rydb.	10	10	Goldblatt 1983
<i>M. depauperata</i> Scribn.	10	10 (4)	Ornduff 1969; Moore 1970; Goldblatt and Johnson 1991, 1994
<i>M. distans</i> Swallen ex Hitchc.	10	20	Ornduff 1969
<i>M. diversiglumis</i> Trin.	10	10 (3)	Ornduff 1969; Goldblatt and Johnson 1991, 1994
<i>M. eludens</i> C.Reeder	10	20 (2)	Goldblatt and Johnson 1991, 1994
<i>M. emersleyi</i> Vasey	10	20 (3), 30, 32	Federov 1969; Ornduff 1969; Moore 1970
<i>M. filiformis</i> Rydb.	9	9 (3)	Federov 1969; Goldblatt and Johnson 1991, 1994
<i>M. filiformis</i> var. <i>fortis</i> E.H.Kelso	9	9	Moore 1970
<i>M. flaviseta</i> Scribn.	10	10	Moore 1970
<i>M. fragilis</i> Swallen	10	10 (2)	Goldblatt and Johnson 1991, 1994
<i>M. frondosa</i> (Poir. ex Lam.) Fernald	10	20	Moore 1971

<i>M. frondosa</i> f. <i>commutata</i> (Scribn.) Fernald	10	20	Ornduff 1967
<i>M. frondosa</i> f. <i>frondosa</i>	10	20	Ornduff 1967
<i>M. gigantea</i> Hitchc.	10, 6	10, 12	Federov 1969; Ornduff 1969
<i>M. glabrifloris</i> Scribn.	10	20	Moore 1971
<i>M. glauca</i> Mez	10	30 (2)	Ornduff 1969; Moore 1970
<i>M. glomerata</i> (Willd.) Trin.	10	10 (3), 20 (2)	Ornduff 1967; Federov 1969; Moore 1971; Goldblatt 1983
<i>M. grandis</i> Vasey	10	10	Moore 1970
<i>M. gypsophila</i> C.Reeder & Reeder	10	10	Ornduff 1968
<i>M. hakonensis</i> (Hack.) Makino	10	20 (2)	Federov 1969; Goldblatt and Johnson 1991
<i>M. himalayensis</i> Hack.	10	20 (2)	Moore 1974; Goldblatt 1981
<i>M. huegelii</i> Trin.	10	20 (3), 21	Federov 1969; Goldblatt 1981; Goldblatt and Johnson 1990
<i>M. implicata</i> (Kunth) Trin.	10	10 (5)	Ornduff 1968; Moore 1970, 1973, 1977; Goldblatt and Johnson 1991, 1994
<i>M. involuta</i> Swallen	6	12	Moore 1970
<i>M. japonica</i> Steud.	10	20 (2), 21 (2)	Federov 1969; Ornduff 1969; Goldblatt and Johnson 1991
<i>M. lehmanniana</i> Henr.	10	10+2B	Goldblatt 1981
<i>M. leptoura</i> Hitchc.	10	20	Moore 1970
<i>M. lindheimeri</i> Hitchc.	10, 6	10 (3), 13	Ornduff 1968; Federov 1969; Moore 1970
<i>M. longistolon</i> Ohwi	10	20, 21	Federov 1969; Goldblatt and Johnson 1991
<i>M. macrostis</i> Hitchc.	10	20, 30	Ornduff 1969; Moore 1970
<i>M. macroura</i> (Kunth) Hitchc.	10, 6	10, 12, c.14, 20 (2)	Ornduff 1968, 1969; Federov 1969; Moore 1970
<i>M. mexicana</i> (L.) Trin.	10	20 (4)	Federov 1969; Moore 1971; Goldblatt 1983
<i>M. mexicana</i> (L.) Trin. f. <i>ambigua</i> (Torr.) Fernald	10	20	Ornduff 1967
<i>M. mexicana</i> f. <i>mexicana</i>	10	20	Ornduff 1967
<i>M. microsperma</i> (DC.) Kunth	10	10 (2)	Goldblatt and Johnson 1991, 1994
<i>M. minutissima</i> (Steud.) Swallen	10	15, 20, 30 (4)	Ornduff 1968; Moore 1970; Goldblatt 1983; Goldblatt and Johnson 1991, 1994
<i>M. montana</i> Hitchc.	10	20 (2)	Moore 1970; Goldblatt and Johnson 2000
<i>M. parviglumes</i> Vasey	10	20	Moore 1970
<i>M. pectinata</i> C.O.Goodd.	10	10 (2)	Goldblatt and Johnson 1991, 1994
<i>M. peruviana</i> (P.Beauv.) Steud.	10	15	Goldblatt and Johnson 1994
<i>M. plumbea</i> Hitchc.	10	20	Ornduff 1969

<i>M. polycaulis</i> Scribn.	10	10, 20 (2)	Federov 1969; Ornduff 1969; Moore 1970
<i>M. porteri</i> Scribn.	10, 6	10, 23/2, 12	Ornduff 1968; Federov 1969; Moore 1970
<i>M. pubescens</i> Hitchc.	10	20 (3), 23, c.30	Ornduff 1967, 1969; Moore 1970
<i>M. pubigluma</i> Swallen	10	10, 15	Ornduff 1969; Moore 1970
<i>M. pulcherrima</i> Scribn.	10	15	Moore 1970
<i>M. pungens</i> Thurb.	10	21, 30	Federov 1969
<i>M. purpusii</i> Mez	10	10	Moore 1970
<i>M. pusilla</i> Steud.	10	15	Goldblatt and Johnson 1991
<i>M. quadridentata</i> (Kunth) Trin.	10	10	Goldblatt and Johnson 2000
<i>M. racemosa</i> B.S.P.	10	20 (3)	Ornduff 1967; Moore 1971; Goldblatt 1983
<i>M. ramosa</i> (Hack.) Makino	10	20	Goldblatt and Johnson 1991
<i>M. ramulosa</i> (Kunth) Swallen	10	10 (4)	Ornduff 1968; Moore 1973; Goldblatt and Johnson 1991, 1994
<i>M. reederorum</i> Soderstr.	10	26, 34 (2)	Ornduff 1969; Moore 1970
<i>M. repens</i> Hitchc.	10, 9	30, c.35, c.36 (2)	Federov 1969; Ornduff 1969; Moore 1970
<i>M. reverchonii</i> Vasey & Scribn.	10	10, 20	Federov 1969; Moore 1970
<i>M. richardsonis</i> Rydb.	10	20 (2)	Federov 1969; Goldblatt 1983
<i>M. rigens</i> Hitchc.	10	20 (2)	Federov 1969; Ornduff 1969
<i>M. rigida</i> (Kunth) Kunth	10	20 (3), c.22	Ornduff 1967; Federov 1969; Moore 1970
<i>M. rigida</i> complex	10	20	Ornduff 1969
<i>M. robusta</i> (E.Fourn.) Hitchc.	10	20	Moore 1970
<i>M. schmitzii</i> Hack.	10	20 (2)	Goldblatt and Johnson 1991, 1994
<i>M. schreberi</i> J.F.Gmel.	10	10, 20 (2)	Ornduff 1967; Federov 1969; Moore 1971
<i>M. setarioides</i> E.Fourn.	10	20 (2)	Moore 1973; Goldblatt 1981
<i>M. setifolia</i> Vasey	10	20	Moore 1970
<i>M. sheperdii</i> (Vasey) Swallen	10	8	Goldblatt and Johnson 1991
<i>M. sinuosa</i> Swallen	10, 6	10 (2), 12 (2)	Goldblatt and Johnson 1991, 1994
<i>M. sobolifera</i> (Muhl. ex Willd.) Trin.	10	10, 20	Ornduff 1967; Moore 1971
<i>M. spiciformis</i> Trin.	10	20	Moore 1970
<i>M. squarrosa</i> Rydb.	10	20	Federov 1969
<i>M. straminea</i> Hitchc.	10, 6	10, 21/2, 11, 12	Goldblatt and Johnson 2000
<i>M. straminea</i> Hitchc. (as <i>Muhlenbergia virescens</i> (Kunth) Trin.)	10, 6	10 (4), 10+1s, 12, 20 (3), 21/2	Ornduff 1969; Moore 1970; Goldblatt and Johnson 2000
<i>M. stricta</i> (J.Presl.) Kunth	10	c.14; 16	Moore 1970

<i>M. strictior</i> Beal	10	10 (3)	Moore 1970; Goldblatt and Johnson 1991, 1994
<i>M. silvatica</i> (Torr.) Torr. ex Gray	10	20 (3)	Ornduff 1967; Moore 1971; Goldblatt 1981
<i>M. tenella</i> (Kunth) Trin.	10	10 (4)	Moore 1972, 1973; Goldblatt and Johnson 1991, 1994
<i>M. tenuifolia</i> (Kunth) Kunth	10	20 (5)	Ornduff 1968, 1969; Moore 1970; Goldblatt and Johnson 1991, 1994
<i>M. tenuifolia</i> (Kunth) Kunth (as <i>Muhlenbergia monticola</i> Buckley)	10	10 (2), 20 (2)	Ornduff 1968, 1969; Federov 1969; Moore 1970
<i>M. tenuiflora</i> B.S.P.	10	10, 20 (2)	Ornduff 1967; Federov 1969; Moore 1971
<i>M. tenuissima</i> (J.Presl.) Kunth	10	10 (2)	Goldblatt and Johnson 1991, 1994
<i>M. texana</i> Buckley	10	20 (3)	Moore 1970; Goldblatt and Johnson 1991, 1994
<i>M. umbrosa</i> Scribn.	10	20	Federov 1969
<i>M. uniflora</i> Fernald	10	c.21	Federov 1969
<i>M. utilis</i> Hitchc.	10	10	Ornduff 1968
<i>M. vaginata</i> Swallen	9	9 (3)	Moore 1970; Goldblatt and Johnson 1991, 1994
<i>M. villiflora</i> Hitchc.	10	10, 11	Ornduff 1969
<i>M. villosa</i> Swallen	10	10	Ornduff 1969
<i>M. wolfii</i> Rydb.	10	10	Moore 1970
<i>Munroa mendocina</i> Phil.	8	8	Federov 1969
<i>M. squarrosa</i> Torr.	8	8 (2)	Moore 1973; Goldblatt 1981
<i>Neeragrostis reptans</i> (Michx.) Nicora	10	30	Moore 1970
<i>Neostapfia colusana</i> Davy	10	20 (2)	Ornduff 1967; Goldblatt 1985
<i>Neyraudia arundinacea</i> (L.) Henrard	10	20	Goldblatt and Johnson 1990
<i>N. madagascariensis</i> Hook.f.	10	20	Ornduff 1967
<i>N. neyraudiana</i> (Kunth) Keng ex Hitchc.	10	20 (4)	Federov 1969; Goldblatt 1981, 1985
<i>Oxychloris scariosa</i> (F.Muell.) Lazarides (as <i>Chloris scariosa</i> F.Muell.)	10	20 (3)	Ornduff 1968; Federov 1969

<i>Orcuttia californica</i> Vasey	10, 6	12, 16	Federov 1969; Goldblatt 1985
<i>Orcuttia californica</i> Vasey	10, 6	12, 16	Federov 1969; Goldblatt 1985
<i>O. fragilis</i> Swallen	10	20	Goldblatt 1985
<i>O. greenei</i> Vasey	6	12 (2)	Federov 1969; Goldblatt 1985
<i>O. inaequalis</i> Hoover	6	12	Goldblatt 1985
<i>O. mucronata</i> Crampton	10	20	Goldblatt 1985
<i>O. pilosa</i> Hoover	10	15, 16	Federov 1969; Goldblatt 1985
<i>O. tenuis</i> Hitchc.	10	13, 23	Federov 1969; Goldblatt 1985
<i>O. viscida</i> (Hoover) Reeder	10	14	Goldblatt 1985
<i>Oropetium capense</i> Stapf	10	20	Moore 1974
<i>O. minimum</i> (Hochst.) Pilg.	9	18	Goldblatt and Johnson 1998
<i>O. thomaeum</i> Trin.	10, 9	5, 9 (2), 10 (2)	Federov 1969; Moore 1972, 1977; Goldblatt and Johnson 1998
<i>Pappophorum bicolor</i> E.Fourn. ex Hemsl.	10	20, 30, 50 (3)	Ornduff 1968; Federov 1969; Moore 1970
<i>P. mucronulutum</i> Nees	10	30 (5)	Ornduff 1968; Federov 1969; Moore 1970
<i>P. pappiferum</i> (Lam.) Kuntze	10	30, 50	Moore 1977; Goldblatt 1981
<i>Pentarrhaphis polymorpha</i> Griffiths	10	10	Federov 1969
<i>P. scabra</i> Kunth	10	10 (2)	Moore 1974
<i>Pereilema brasilianum</i> Trin.	10	20	Goldblatt 1981
<i>P. ciliatum</i> E.Fourn. ex Hemsl.	10	10, 20	Moore 1972, 1970
<i>P. crinitum</i> J. & C.Presl.	10	10 (3)	Moore 1972, 1977; Goldblatt 1988
<i>Perotis hilderbrandtii</i> Mez	10	20	Ornduff 1968
<i>P. hordeiformis</i> Nees ex Hook. & Arn.	10, 9	9, 18, 20	Federov 1969; Goldblatt and Johnson 1990
<i>P. indica</i> (L.) Kuntze	10	10 (2), 20	Moore 1972, 1977; Goldblatt 1981
<i>P. patens</i> Gand.	10	20 (3)	Federov 1969; Goldblatt 1981, 1983
<i>P. vaginata</i> Hack.	10	20 (2)	Goldblatt 1981, 1985
<i>Pringleochloa stolonifera</i> Scribn.	10	20 (2)	Ornduff 1968, 1969
<i>Redfeldia flexuosa</i> (Thurb.) Vasey	10	25/2, 12II+1I	Goldblatt 1981
<i>Reederochloa eludens</i> Soderstr. & H.F.Decker	10	19 (2)	Ornduff 1969; Goldblatt 1988
<i>Schedonnardus paniculatus</i> Trel.	10	10 (4), 15	Federov 1969; Ornduff 1969; Moore 1970; Goldblatt 1981, 1983
<i>Scleropogon brevifolius</i> Phil.	10	14, 20 (4)	Ornduff 1969; Moore 1970, 1973; Goldblatt 1981

<i>Spartina alterniflora</i> Loisel.	10, 7	20, 21, 28, 30, 31 (7), 35 (3)	Federov 1969; Moore 1970, 1972; Goldblatt 1981
<i>S. anglica</i> C.E.Hubb.	10	30, 31, c.60, 64	Moore 1972; Goldblatt 1985
<i>S. arundinacea</i> Carmich.	10	20	Moore 1970
<i>S. bakeri</i> Merr.	10, 7	20 (2), 21	Federov 1969; Moore 1970
<i>S. caespitosa</i> A.A.Eaton	10, 7	20, 21	Federov 1969; Moore 1972
<i>S. cynosuroides</i> Roth	10, 7	14, 20, 21 (3), 40	Federov 1969; Moore 1970
<i>S. foliosa</i> Trin.	10	30	Goldblatt 1988
<i>S. glabra</i> Muhl.	10, 7	28, 31 (2)	Federov 1969; Moore 1970
<i>S. gracilis</i> Trin.	10, 7	20 (3), 21 (2)	Federov 1969; Moore 1970; Goldblatt 1981, 1983
<i>S. leiantha</i> Benth.	7	28	Federov 1969
<i>S. maritima</i> Fernald	10, 7	28, 30 (4)	Federov 1969; Moore 1970, 1972
<i>S. michauxiana</i> Hitchc.	7	14	Federov 1969
<i>S.x neyrautii</i> Foucaud	10	31	Goldblatt 1981
<i>S. patens</i> Muhl.	10, 7	14, 20 (5), 21, 28	Federov 1969; Ornduff 1969; Moore 1970, 1972
<i>S. pectinata</i> Bosc ex Link	10, 7	20 (6), 20+2B, 21 (2), 24, 35, 40	Federov 1969; Moore 1970, 1972; Goldblatt 1981, 1983
<i>S. schreberii</i> J.F.Gmel.	10	20 (2)	Federov 1969
<i>S. spartinae</i> (Trin.) Merr.	10, 7	14, 20(4), 21	Federov 1969; Moore 1970, 1973
<i>S. stricta</i> Roth	7	28	Federov 1969
<i>S. townsendii</i> H. & J.Groves	10, 7	30, 31 (4), 38, 45, 60, 61, 62 (2), 63 (3)	Federov 1969; Moore 1970, 1977; Goldblatt 1981, 1985
<i>S. townsendii-sterilis</i> f. <i>donegalensis</i>	10	60	Moore 1974
<i>Sporobolus africanus</i> (Poir.) Robyns & Tournay	6/9	18 (2)	Ornduff 1968; Federov 1969; Goldblatt and Johnson 1996
<i>S. airoides</i> Torr.	9	c.45, 54, 63	Federov 1969
<i>S. airoides</i> (Torr.) Torr. var. <i>airoides</i>	10	c.40	Ornduff 1968
<i>S. agrostoides</i> Chiov.	6	24	Goldblatt 1981
<i>S. arabicus</i> Boiss.	6/9	18	Goldblatt and Johnson 1998

<i>S. asper</i> (Michx.) Kunth var. <i>asper</i> (Michx.) Kunth	9	27	Goldblatt 1981
<i>S. asper</i> var. <i>macer</i> (Trin.) Shinners	10, 6	18, 19, 20, 21, 22, 23, 24	Goldblatt 1981
<i>S. atrovirens</i> Kunth	6	12	Goldblatt 1988
<i>S. buckleyi</i> Vasey	10	20 (2)	Federov 1969; Moore 1972
<i>S. ciliatus</i> J.Presl.	9	27	Moore 1973
<i>S. clandestinus</i> (Biehler) Hitchc.	6, 9	23, 24, 26, 27, 28	Goldblatt 1981
<i>S. compositus</i> (Poir.) Merr. (as <i>Sporobolus asper</i> Kunth)	9	27 (4), c.44, 54	Federov 1969; Moore 1970; Goldblatt 1983
<i>S. congoensis</i> Franch.	10	10	Goldblatt 1983
<i>S. contractus</i> Hitchc.	6/9	18 (3)	Federov 1969; Goldblatt 1981; Goldblatt and Johnson 1991
<i>S. coromandelianus</i> (Retz.) Kunth	6	12, 18 (6)	Moore 1974, 1977; Goldblatt 1988; Goldblatt and Johnson 1990, 1991, 1998
<i>S. cubensis</i> Hitchc.	10, 6/9	36, 40, c.45	Ornduff 1969
<i>S. crytandrus</i> A.Gray	9	9, 18 (5), 19 (6), 36	Ornduff 1968; Federov 1969; Moore 1970; Goldblatt 1981, 1983; Goldblatt and Johnson 1991
<i>S. diander</i> P.Beauv.	6	12 (13), 18 (7), 24	Federov 1969; Moore 1973, 1974, 1977; Goldblatt 1981, 1985; Goldblatt and Johnson 1990, 1991, 1998
<i>S. elongatus</i> R.Br.	6/9	18 (3)	Federov 1969
<i>S. festivus</i> A.Rich.	10, 6	10, 12	Ornduff 1967; Goldblatt 1983
<i>S. fimbriatus</i> (Trin.) Nees	6, 9	18, 24, 27	Goldblatt 1981, 1983
<i>S. flexuosus</i> Rydb.	6/9	18, 19	Federov 1969; Goldblatt 1981
<i>S. giganteus</i> Rydb.	6/9	18	Federov 1969
<i>S. helvola</i> T.Durand & Schinz	10, 6/9	8, 10, 14, 16, 18 (3)	Goldblatt and Johnson 1990, 1991
<i>S. heterolepis</i> A.Gray	6/9	36 (2)	Federov 1969; Goldblatt 1983
<i>S. indicus</i> (L.) R.Br.	10, 6, 9	9, 12 (6), 18 (9), 20 (2), 41/2, 21, 43/2, 22, 45/2, c.24	Ornduff 1967, 1968, 1969; Federov 1969; Moore 1974, 1977; Goldblatt 1988; Goldblatt and Johnson 1990, 1991, 1994



<i>S. indicus</i> (L.) R.Br. (as <i>Sporobolus beteroanus</i> (Trin.) Hitc. & Chase)	10, 6/9	18, c.22	Federov 1969; Ornduff 1967
<i>S. indicus</i> var. <i>indicus</i> Jovet & Guédès (as <i>Sporobolus poiretti</i> Hitc.)	6/9	18 (7)	Ornduff 1967, 1968, 1969; Federov 1969; Moore 1970, 1972
<i>S. indicus</i> var. <i>major</i> (Büse) Baaijens (as <i>Sporobolus fertilis</i> (Steud.) Clayton)	6	12, 18 (3), 24 (5)	Ornduff 1969; Moore 1974, 1977; Goldblatt 1981, 1983, 1985
<i>S. indicus</i> var. <i>pyramidalis</i> (P.Beauv.) Veldkamp (as <i>Sporobolus jacquemontii</i> Kunth)	6	12 (3)	Moore 1972, 1973; Goldblatt 1981
<i>S. infirmus</i> Mez	10	10	Goldblatt 1981
<i>S. interruptus</i> Vasey	10	15	Goldblatt 1981
<i>S. japonicus</i> Maxim. ex Rendle	10	20 (2)	Federov 1969
<i>S. macrospermus</i> Scribn. ex Beal	10	20	Moore 1970
<i>S. maderaspatanus</i> Bor	6	6	Goldblatt 1988
<i>S. marginatus</i> Hochst. ex A.Rich.	10, 9	9 (6), 9+, 10 (2), 18 (4)	Moore 1971, 1972; Goldblatt 1988; Goldblatt and Johnson 1990, 1991
<i>S. molleri</i> Hack.	6	6 (4)	Ornduff 1967; Goldblatt 1981, 1983
<i>S. montanus</i> Engl.	10, 9	9, 10	Goldblatt 1981; Goldblatt and Johnson 1996
<i>S. nealleyi</i> Vasey	10	20 (3)	Ornduff 1969; Moore 1973; Goldblatt 1981
<i>S. neglectus</i> Nash	6/9	18 (2)	Federov 1969; Goldblatt 1983
<i>S. nervosus</i> Hochst.	6/9	18	Goldblatt and Johnson 1998
<i>S. olivaceus</i> Napper	6	12	Moore 1972
<i>S. pectinellus</i> Mez	6	12	Moore 1977
<i>S. piliferus</i> Kunth	10, 6/9	10, 18, 20 (2)	Goldblatt 1981, 1985, 1988
<i>S. purpurascens</i> Ham.	10	30 (2)	Moore 1973, 1974
<i>S. pyramidalis</i> P.Beauv.	6	12 (5), 18	Ornduff 1967; Moore 1977; Goldblatt 1981
<i>S. pyramidatus</i> (Lam.) Hitc.	6, 9	12 (4), 18 (3), 27	Ornduff 1968, 1969; Federov 1969; Moore 1970, 1972, 1973
<i>S. rigens</i> E.Desv.	6/9	c.72	Goldblatt 1981
<i>S. robustus</i> Kunth	10	20 (2)	Ornduff 1967; Goldblatt 1981
<i>S. sanguineus</i> Rendle	10	10	Goldblatt 1983
<i>S. spiciformis</i> Swallen	10	20 (2)	Moore 1970, 1973
<i>S. subtilis</i> Kunth	10	10	Goldblatt 1981

<i>S. tenuissimus</i> Kuntze	10, 6	6 (3), 20	Ornduff 1967; Federov 1969; Moore 1974; Goldblatt 1981
<i>S. tremulus</i> Kunth	10, 6	10 (2), 24	Federov 1969; Moore 1977
<i>S. trichodes</i> Hitchc.	6	12	Ornduff 1967
<i>S. vaginiflorus</i> (Torr. ex A.Gray) A.W.Wood	9	27 (2)	Federov 1969; Moore 1970
<i>S. virginicus</i> (L.) Kunth	10, 9	9, 10 (5), 15 (2), 31/2, 20, 25, 30	Moore 1970, 1972, 1974; Goldblatt 1981; Goldblatt and Johnson 1991
<i>S. virginicus</i> var. <i>arenarius</i> (Gouan) Maire	10, 6/9	23/2-15	Moore 1977
<i>S. virginicus</i> var. <i>minor</i> F.M.Bailey	10	10 (2), 15 (2), 20 (2)	Goldblatt 1983
<i>S. virginicus</i> var. <i>virginicus</i> (L.) Kunth	10	20 (2)	Goldblatt 1983
<i>S. wallichii</i> Munro ex Trin.	10, 6	c.20, 12	Federov 1969; Moore 1977
<i>S. wrightii</i> Munro ex Scribn.	6/9	18	Federov 1969
<i>Tetrapogon spathaceus</i> Hack. ex Dur. & Schinz	9	9 (2)	Ornduff 1968; Federov 1969
<i>T. tenellus</i> Chiov.	10, 9	9 (2), 10	Ornduff 1968; Federov 1969; Goldblatt and Johnson 1998
<i>T. villosus</i> Desf.	10	10 (2)	Moore 1972; Goldblatt and Johnson 1998
<i>Tragus berteronianus</i> Schult.	10	10 (10)	Ornduff 1967, 1968, 1969; Federov 1969; Moore 1970, 1973, 1974; Goldblatt 1983; Goldblatt and Johnson 1998
<i>T. biflorus</i> Schult.	10	10 (4)	Moore 1972, 1977; Goldblatt 1981, 1983
<i>T. muricatus</i> Moench	10	18, 20	Moore 1977; Goldblatt 1981
<i>T. racemosus</i> (L.) All.	10	20 (10)	Moore 1977; Federov 1969; Goldblatt 1981, 1983, 1988; Goldblatt and Johnson 1994
<i>T. roxburghii</i> Panigrahi	10	10 (2)	Goldblatt and Johnson 1990, Johnson 1998
<i>Trichloris crinita</i> (Lag.) Parodi	10	20	Federov 1969
<i>T. mendocina</i> (Phil.) Kurtz	10	20 (2)	Federov 1969
<i>T. pluriflora</i> E.Fourn.	10	30, 40	Federov 1969; Moore 1970
<i>Tridens albescens</i> Wooton & Standl.	10	30 (3)	Moore 1970, 1973; Goldblatt 1981
<i>Tridens albescens</i> Wooton & Standl. (as <i>Triodia albescens</i> (Vasey))	8	16, 36	Federov 1969

<i>T. ambigua</i> Schult.	10	20	Moore 1970
<i>T. avenaceus</i> Hitchc.	8	16	Federov 1969
<i>T. buckleyanus</i> Nash	10	20	Goldblatt 1988
<i>T. elongatus</i> (Buckley) Nash	10	20 (2)	Federov 1969
<i>T. eragrostoides</i> Nash	10	20 (2)	Federov 1969; Moore 1970
<i>T. flavus</i> Hitchc.	10	20 (2)	Federov 1969
<i>T. grandiflorus</i> (Vasey) Wooton & Standl.	8	16	Federov 1969
<i>T. muticus</i> Nash	10	20 (5)	Federov 1969; Moore 1973; Goldblatt 1981
<i>T. muticus</i> var. <i>elongatus</i> (Buckley) Shinnery	10	20, 20+2B	Moore 1970
<i>T. nealleyi</i> (Vasey) Wooton & Standl.	8	8	Federov 1969
<i>T. pilosus</i> (Buckley) Hitchc.	8	8	Federov 1969
<i>T. pulchellus</i> (Kunth) Hitchc.	8	8	Federov 1969
<i>T. strictus</i> Nash	10	20	Moore 1970
<i>T. texanus</i> Nash	10	20 (5)	Ornduff 1968; Federov 1969; Moore 1970
<i>Triodia basedowii</i> E.Pritz.	10	c.30	Moore 1973
<i>T. buckleyana</i> Vasey	8	16	Federov 1969
<i>T. cuprea</i> Jacq.	10	21	Federov 1969
<i>T. elongata</i> (Buckley) Smyth	8	16	Federov 1969
<i>T. flava</i> Hitchc.	10	14	Federov 1969
<i>T. irritans</i> R.Br.	10	c.15	Moore 1973
<i>T. mutica</i> Benth. ex S.Watson	8	16	Federov 1969
<i>T. pilosa</i> (Buckley) Merr.	8	8 (2)	Federov 1969
<i>T. stricta</i> Vasey	8	16	Federov 1969
<i>T. texana</i> S.Watson	8	8	Federov 1969
<i>T. thomsonii</i> Petrie	8	24	Federov 1969
<i>Triplasis purpurea</i> Chapm.	10	20	Moore 1970
<i>Tripogon bromoides</i> Roth ex Roem. & Schult.	10	10 (4)	Moore 1977; Goldblatt 1981, 1985
<i>T. chinensis</i> Hack.	10	10	Goldblatt and Johnson 1994
<i>T. filiformis</i> Nees ex Steud.	10	10 (5)	Moore 1974, 1977; Goldblatt 1981, 1985
<i>T. jacquemontii</i> Stapf	10	10, 20	Goldblatt 1981, 1988
<i>T. japonicus</i> (Honda) Ohwi	10	10	Federov 1969
<i>T. minimus</i> Hochst. ex Steud.	10	10 (4)	Ornduff 1968; Federov 1969; Moore 1974, 1977
<i>T. spicatus</i> Ekman	10	10 (2)	Federov 1969; Moore 1974

<i>T. trifidus</i> Munro ex Hook.f.	10	10 (3), 20, 29, 30 (3)	Moore 1974, 1977; Goldblatt 1981, 1985
<i>T. wardii</i> Bor	10	10	Moore 1977
<i>Uniola latifolia</i> Michx.	6	12, 24 (6)	Ornduff 1968; Federov 1969
<i>U. laxa</i> (L.) B.S.P.	6	12 (3)	Ornduff 1968; Federov 1969
<i>U. nitida</i> Baldwin ex Elliott	6	12 (2)	Ornduff 1968
<i>U. ornithorhyncha</i> Steud.	6	12 (2)	Ornduff 1968
<i>U. ornithorhyncha x laxa</i>	6	12	Ornduff 1968
<i>U. paniculata</i> Roth	10	20 (2)	Ornduff 1968
<i>U. pittieri</i> Hack.	10	20 (2)	Moore 1973, 1974
<i>U. sessiliflora</i> Poir.	6	12 (3)	Ornduff 1968; Federov 1969
<i>U. virgata</i> Griseb.	10	20 (2)	Ornduff 1968
<i>Vaseyochloa multinervosa</i> (Vasey) Hitchc.	10	28?, 34	Federov 1969
<i>Willkommia texana</i> Hitchc.	10	30	Federov 1969
<i>Zoysia japonica</i> Steud.	10	20 (2)	Federov 1969
<i>Z. macrostachya</i> Franch. & Sav.	10	20 (2)	Federov 1969
<i>Z. matrella</i> (L.) Merr.	10	10, 20 (4)	Federov 1969; Moore 1977; Goldblatt 1981
<i>Z. tenuifolia</i> Trin.	10	20 (2)	Federov 1969
<i>Zoysia</i> sp.	10	20	Federov 1969

**APPENDIX D.** Aligned sequence data of chloridoid and outgroup taxa for *ITS*. Numbers indicate the consecutive positions of 1 to 668 (5' to 3') from the beginning of the *ITS1* region to the end of the *ITS2* region. Dashes denote gaps. Arrows indicate the beginning of the *ITS1*, 5.8S and *ITS2* regions. N indicates large regions of missing data where sequencing reactions failed. Due to the large size of the matrix and to keep sequences together, this appendix starts on the next page.

IMPERATACYLINDRICA  
 SORGHUMHALEPENSE  
 PANICUMLANIPES  
 PANICUMSTAFFIANUM  
 KARROOCHLOAPURPUREA6241  
 CHAETOBROMUSINVOLUCRATES6284  
 Merxmullerangei  
 Centropodiaglauca  
 Cynodondactylon5248  
 Cynodondactylon7480  
 Cynodondactylon  
 Eleusinecoracana  
 Eleusineindica  
 Chlorisvirgata6616  
 Chlorisvirgata7498  
 Eustachyspaspaloides5597  
 Harpochloafalx6530  
 Harpochloafalx5118  
 Harpochloafalx6670  
 Rendliaaltera5133  
 Leptochloafusca7622  
 Enteropogonmacrostachyus9339  
 Astreblalappacea  
 Muhlenbergiarichardsonis  
 Triodiastenostachya  
 Microchloacaffra5146  
 Dactylocteniumaegyptium9326  
 Tragusberteronianus6620  
 Tragusracemosus7108  
 Traguskoelerioides6729  
 Polevansiarigida7621  
 Perotispatens9323  
 Eragrostiscurvula6529  
 Eragrostisobtusata6623  
 Eragrostisobtusata6732  
 Eragrostisechinochloidea7485  
 Eragrostispseudobtusata7508  
 Eragrostisechinochloidea5933  
 Eragrostislehmanniana20  
 Eragrostiscurvula5191  
 Eragrostischloromelas6947  
 Eragrostischloromelas7474  
 Eragrostisracemosa6533  
 Eragrostisbiflora6705  
 Eragrostissuperba7470  
 Eragrostiscapensis6551  
 Eragrostiscapensis6532  
 Pogonarthriasquarrosa7483  
 Pogonarthriasquarrosa7504  
 Catalepisgracilis6962  
 Catalepisgracilis6653  
 Cladoraphiscyperoides5704  
 Cladoraphisspinosa6335  
 Cladoraphiscyperoides4885  
 Cladoraphiscyperoides4894  
 Fingerhuthiaafricana4349  
 Fingerhuthiaafricana7507  
 Fingerhuthiaafricana4879  
 Entoplocamiaaristulata9353  
 Enneapogonscoparius18  
 Enneapogoncenchroides7506  
 Enneapogoncenchroides5213  
 Enneapogonscoparius7487  
 Schmidtiaappophoroides6334  
 Schmidtiakalihariensis7490  
 Schmidtiakalihariensis7486  
 Sporobolusafricanus7476  
 Sporobolusconsimilis9354  
 Sporobolusfimbriatus9328

50

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 TCGTGACC-T TAAACAAAAC AGACCGTGAA CAT-GTCTCT CATGTCGTCCG  
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 TCGTGACCCT TAAACAAAAC AGACCGTGAA CAT-GTCATC CATGTCGTCCG  
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 TCGTGACCCT AACC-AAAAG AGACTGTGAA TAT-GTCATC TATCTGTCCG  
 TCGTGACCCT AACC-AAAAT AGACTGTGAA CAT-GTCATC AATTCCACCG

IMPERATACYLINDRICA	GGC-----	-----	TCCGGCCCCG	CCAAGGCCCC	CGA-GCTCCG
SORGHUMHALEPENSE	AGC-----	-----	TTTGGCTCGG	CCAAGGTCCC	CTT-GCTCCA
PANICUMLANIPES	GGC-----	-----	TACGGCCCCG	CCAAGGCCCC	CAA-CCTTCG
PANICUMSTAFFIANUM	GGC-----	-----	TACGTTCGCG	CCAAGGCCCC	CCT-TCACAT
KARROOCHLOAPURPUREA6241	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	CGA-CCTCCG
CHAETOBROMUSINVOLUCRATES6284	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Merxmuelleeracana	GACGCGGGG	CTC--GCCCC	CGTCGCCCGG	CACAGGCCCC	CGA-CCTCCG
Centropodiaglauca	GCCGGCGGGG	CTC--GCCCC	CGCCGACCGG	CACAGGCCCC	CGA-CCTCCG
Cynodondactylon5248	GTTGATGGG-	CTT--GCACC	TATCTCTCGG	TCTAGGCCAC	CGA-CCTTCT
Cynodondactylon7480	GTTGATGGG-	CTT--GCACC	TATCTCTCGG	TCTAGGCCAC	CGA-CCTTCT
Cynodondactylon	GTTGATGGG	CTT--GCACC	TATCTCTCGG	TCTAGGCCAC	CGA-CCTTCT
Eleusinecoracana	GGCGATGGG	CTT--GCACC	CATCTCTTGG	AACAGGGCCG	CCA-CCTTCT
Eleusineindica	GGCGATGGG	CTT--GCACC	CATCTCTTGG	AACAGGGCCG	CCA-CCTTCT
Chlorisvirgata6616	TTWGATGGG	CTT--GCACC	TATCTCTCGG	TTTAGGCCCG	CAA-CCTTCT
Chlorisvirgata7498	TTTGATGGG	CTT--GCACC	TATCTCTCGG	TTTAGGCCCG	CAA-CCTTCT
Eustachyospaloides5597	GTTGATGGG	CTT--GCACC	TATCTCTCGG	TCTAGGCCAC	CGA-CCTTCT
Harpochloafalx5118	GTTGATGGG-	CTT--GCACC	TATCTCTCGG	TCTAGGCCAC	TGA-CCTTCT
Harpochloafalx5118	GTTGATGGG-	CTT--GCACC	TATCTCTCGG	TCTAGGCCAC	TGA-CCTTCT
Harpochloafalx6670	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Rendliaaltera5133	GTTGATGGG	CTT--GCACC	TATCTCTTGG	TCTAGGCCAC	CAA-CCTTCT
Leptochloafusca7622	GGTGATGGG	CTT--GCACC	CGTCTCCCGG	TCTAGGCCAC	CGA-CCTTCT
Enteropogonmacrostachyus9339	GG-GATGGGG	CTT--GCACC	TATCTGTGCA	CGGTATCCCC	CGA-TCCTCT
Astreblalappacea	GGTGATGGG	CTT--GCACC	TATCCCTCGG	TCTAGGCCAA	CGA-CCTTCT
Muhlenbergiarichardsonis	ATTGACGGGG	CTT--GCACC	CGTCTCTCGG	TCTGGGGCAC	ATTACCTTCA
Triodiastenostachya	GGTGACGGGG	CTC--GCAAC	CGTCACCCGG	TACAGGCCCG	CGA-CCTTCT
Microchloacaffra5146	GTTGATGGG	CTT--GTACC	TATCTCTCGG	TCTAGGCCAC	CAA-CCTTCT
Dactylocteniumaegyptium9326	GTTGACGGGG	CTC--GCACC	CGTCTCTCGG	TAGGGCAGG-	CGA-CCTTCC
Tragusberteronianus6620	AGTGATGGG	CTT--GCACC	CGTCTCTTGG	CCCTGGGCTC	CGA-ACTTCG
Tragusracemosus7108	AGTGATGGG	CTT--GCACC	CGTCTCTTGG	CCCTGGGCTC	CGA-ACTTCG
Traguskoelerioides6729	AGTGACGGG	CTT--GCACC	CGTCTCTTGG	CCCTGGGCTC	CGA-ACTTCG
Polevansiarigida7621	AGNGATGGG	CTT--GCACC	CATCTCTCGG	TCCATGGGCTC	C-A-ACTTCT
Perotispatens9323	GGTGATGGG	-TA--GCACC	TGTCTCCTAG	TATTGGTGAC	TTA-CCTTCC
Eragrostiscurvula6529	NNNNNNNNNN	NNNNNNNNNTC	AA-CTCCCGG	CTTAG-CCCC	AGA--CTTCC
Eragrostisobtusa6623	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNGC
Eragrostisobtusa6732	GGTGACGGGG	CTTTTGCCTC	GC-CTCCCGG	CTTAGGCCCC	AGA--CTTCC
Eragrostisechinochloidea7485	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostispseudobtusa7508	GGTGACGGGG	CTTTTGCCTC	GC-CTCCCGG	CTTAGGCCCC	AGA--CTTCC
Eragrostisechinochloidea5933	GGTGACGGGG	CTT--GCACC	GC-CTCCCGG	CTTAGGCCCC	AGA--CTTCC
Eragrostislehmanniana20	GGWGACGGGG	CTT--GCCTC	GG-CTCCCGG	CTTAGGCCCC	AGA--CTTCC
Eragrostiscurvula5191	NNNNNNNNNN	NNNNNNNNNTC	GC-CTCTCGG	CTTAGGCCCC	AGA--CTTCC
Eragrostischloromelas6947	GGTGACGGGG	CTT--GCCTC	GR-CTCCCGG	CTTAGGCCCC	AGA--CTTCC
Eragrostischloromelas7474	GGTGACGGGG	CTT--GCCTC	GC-CTCCCGG	CTTAGGTCCC	AGA--CTTCC
Eragrostisracemosa6533	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNAGGCCCC	A-A--CTTTC
Eragrostisbiflora6705	GGTGACGGGG	CTT--GCCTC	GC-CTCCCGG	CTTAGGTCCC	AGA--CTTTC
Eragrostissubarba7470	GGCGACGGGG	CTC--ACCTC	GC-CTCCCGG	TTTAGGCCCC	AGA--CTTTC
Eragrostiscapensis6551	GCTGACGGGG	CTC--GCCTC	GC-CTCCCGG	CTTAGGCCCC	TGT--CTTCC
Eragrostiscapensis6532	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNCC
Pogonarthriasquarrosa7483	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Pogonarthriasquarrosa7504	AGTTACGGGG	CCT--GCCTC	GT-CACTCGG	CTTAGGCCCC	AGA--CTTCC
Catalepisgracilis6962	GGTGATGGG	CTA--GCCTC	GC-CTCCCGG	CTTAGGTCCC	AGA--CTTCC
Catalepisgracilis6653	NNNNNNNNNN	NNNNNNNNNN	NNNNTCCCGG	CTTAGGTCCC	AAA--CTTCC
Cladoraphiscyperoides5704	NNNNNNNNNN	NNNNNNNNNTC	GC-CTCTCGA	CTTAGGCCCC	AGA--CTTTC
Cladoraphisspinosa6335	NNNNNNNNNN	NNNNNNNNNTC	GC-CTCTCGA	CTTAGGCCCC	AGA--CTTTC
Cladoraphisspinosa4885	AGGGATGGGG	CTC--GCCTC	GC-CTCTCGA	CTTAGGCCCC	AGA--CTTTC
Cladoraphiscyperoides4894	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Fingerhuthiaafricana4349	GGCGACGGGG	CTT--GCTCC	GC-CTCCCGA	CATAGGCCCC	AGA--CTTCT
Fingerhuthiaafricana7507	GGCGACGGGG	CTT--GCTCC	GC-CTCCCGA	CATAGGCCCC	AGA--CTTCT
Fingerhuthiaafricana4879	GGMGACGGGG	CTT--GCTCC	GC-CTCCCGA	CATAGGCCCC	AGA--CTTCT
Entoplocamiaaristulata9353	GGCGATAGGG	CTT--GCCTT	GT-CTCCCGA	CTTAGGCCCC	CGA--TCTCC
Enneapogoncoparius18	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNCGA	TGT--CAGCC
Enneapogoncenchroides7506	GAAGACGGGG	CTC--RCCTC	GC-CTCCCGG	CATCGGCCGA	TGT--CATCC
Enneapogoncenchroides5213	GAAGACGGGG	CTC--GCCTC	GC-CTCCCGG	CATCGGCCGA	TGT--CATCC
Enneapogoncoparius7487	GAAGACGGGG	CTC--GCCTC	GC-CTCCCGG	CCTCGGCCGA	TGT--CGTCC
Schmidtiaapapphoroides6334	GGAGACGGGG	CTT--GCCTC	GC-CTCACGG	CATCGGCTGA	TGA--CCTCC
Schmidtiaakalihariensis7490	GGAGACGGGG	CTC--GCCTC	GC-CTCACGG	CATCGGCTGA	TGA--CCTCC
Schmidtiaakalihariensis7486	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNCGA	TGA--CCTCC
Sporobolusafricanus7476	GGTGATGGTC	ATT--GCACT	GT-CTCTTGG	CTTGGACCCT	TGT--CGTTC
Sporobolusconsimilis9354	GGCGACGGGT	CTT--GCACC	GT-CTCTCGG	CATAGACCCT	TAT--CTTTC
Sporobolusfimbriatus9328	GGTGATGGCC	ATA--RTGTC	GT-CTCCTGG	CATGGGCCTC	TAT--CCTTC

IMPERATACYLINDRICA  
 SORGHUMHALEPENSE  
 PANICUMLANIPES  
 PANICUMSTAPFIANUM  
 KARROOCHLOAPURPUREA6241  
 CHAETOBROMUSINVOLUCRATES6284  
 Merxmullerangei  
 Centropodiaglauca  
 Cynodondactylon5248  
 Cynodondactylon7480  
 Cynodondactylon  
 Eleusinecoracana  
 Eleusineindica  
 Chlorisvirgata6616  
 Chlorisvirgata7498  
 Eustachyaspaloides5597  
 Harpochloafalx6530  
 Harpochloafalx5118  
 Harpochloafalx6670  
 Rendliaaltera5133  
 Leptochloafusca7622  
 Enteropogonmacrostachyus9339  
 Astreblalappacea  
 Muhlenbergiarichardsonis  
 Triodiastenostachya  
 Microchlocaffra5146  
 Dactylocteniumaegyptium9326  
 Tragusberteronianus6620  
 Tragusracemosus7108  
 Traguskoelerioides6729  
 Polevansiarigida7621  
 Perotispatens9323  
 Eragrostiscurvula6529  
 Eragrostisobtusa6623  
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 Eragrostiscurvula5191  
 Eragrostischloromelas6947  
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 Pogonarthriasquarrosa7483  
 Pogonarthriasquarrosa7504  
 Catalepisgracilis6962  
 Catalepisgracilis6653  
 Cladoraphiscyperoides5704  
 Cladoraphispinosa6335  
 Cladoraphispinosa4885  
 Cladoraphiscyperoides4894  
 Fingerhuthiaafricana4349  
 Fingerhuthiaafricana7507  
 Fingerhuthiaafricana4879  
 Entoplocamiiaristulata9353  
 Enneapogonscoparius18  
 Enneapogoncenchroides7506  
 Enneapogoncenchroides5213  
 Enneapogonscoparius7487  
 Schmidtiapappophoroides6334  
 Schmidtiakalihariensis7490  
 Schmidtiakalihariensis7486  
 Sporobolusaffricanus7476  
 Sporobolusconsimilis9354  
 Sporobolusfimbriatus9328

101

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 ACCT---GGG GCAGAG---G GGCCA-CAAA A-GAACCAC GGCGCCTTA-  
 TTTCGTTGGA TG-GAG--- -GCCG-CCAA A-GAACCAC GGCGCCAAA-  
 TTTTGTGTTGGA GG-GAG--- -GCCG-CCAA A-GAACCAG GGCGCCGAA-  
 TCA---GGA GGAGAG---T GGCCG-CAAA A-GAACCAAC GGCGCCGAAC  
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 TCC---TGG GGGACG---G GGGCC-CAAA A-GAACCAC GGCGCCGTAT  
 TTC---GGA GGTACG---G GGCC-CAAA A-GAACCAC GGCGCCGTAT  
 TTTT---GAA GAAAA---G TGACC-CAAA A-GAACCCAC GGCGCCGTAT  
 TCC---AGA GGGGA---G TGCCACAAA A-GAACCAC GGCGCCGTAT  
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 ATC---AGA GGGGA---G GGCC-CAAA A-GAACCAC GGCGCCGTAT  
 ATC---AGA GGGGA---G GGCC-CAAA A-GAACCAC GGCGCCGTAT  
 CATT---ASA GGC---G GGCC-CAAA A-GAACCAC GGCGCCGTAT  
 TT---AGGG ATGGTG---AG TGCCACAAA A-GAACCC-C GGCGCCGTGT  
 GTA---TGG GGGCTG---G AGCCG-CAAA A-GAACCAC GGCGCCATCT  
 TA---TGG GGGCTG---G AGCCG-CAAA A-GAACCAC GGCGCCG-AT  
 GTA---TGG GGGCTG---G AGCCG-CAAA A-GAACCAC GGCGCCG-AT  
 NTA---TGG GGGATG---G AGCCG-CAAA A-GAACCAC GGCGCCG-AT  
 GTA---TGG GGGCTG---G AGCCG-CAAA A-GAACCAC GGCGCCG-AT  
 GTA---TGG GGGATG---G AGCCG-CAAA A-GAACCAC GGCGCCG-AT  
 ATA---TGG GGGCTG---G AGCCG-CAAA A-GAACCAC GGCGCCA-TC  
 GTA---TGG GGGTTG---G AGCCG-CAAA A-GAACCAC GGCGCCA-TC  
 GTA---TGG GGGGTTG---G AGCCG-CAAA A-GAACCAC GGCGCCA-TC  
 GTA---TGG GGGCTG---G GGCCG-CAAA A-GAACCAC GGCGCCA-TG  
 GTA---CSGA GGGATG---G GGCCG-CAAA A-GAACCAC GGCGCCA-TC  
 GTA---TGG GGTGTTG---G GGCCG-CAAA A-GAACCAC GGCGCCA-TC  
 GTA---TGG GGGATG---G GGCC-CAAC A-GAACCAC GGCGCCG-AT  
 TT---TGG GGGCTA---G GGCCG-CAAA A-GAACCAC GGCGCCG-AT  
 TT---TGG GGGGTA---G GGCCG-CAAA A-GAACCAC GGCGCCG-AT  
 NNNNNNNNNN NNNNNNNNNN NNCCA-CAAA A-GAACCAC AGCGCCG-AT  
 GTA---TGG GGGCTG---G AGCCG-CAAA A-GAACCAC GGCGCCG-AT  
 TTA---TGG GGGATG---G GGCCG-CAAA A-GAACCAC GGCGCCA-TC  
 TTA---TGG GGGATG---G GGCCG-CAAA A-GAACCAC GGCGCCA-TC  
 GTA---TGG TGGCTG---G GGCCG-CAAC A-AAACCAC GGCGCCG-AT  
 GTA---TGG TGGCTG---G GGCCG-CAAC A-AAACCAC GGCGCCG-AT  
 NNA---TGG TGGCTG---G GGCCG-CAAC A-AAACCAC GGCGCCG-AT  
 TTA---TGG GGGATG---G GGCCG-CAAC A-GAACCAC GGCGCCGTGT  
 TTA---TGG GGGATG---G GGCCG-CAAC A-GAACCAC GGCGCCGTAT  
 TTA---TGG GGGATG---G GGCCG-CAAC A-GAACCAC GGCGCCGTGT  
 GCT---TGGT GGTGTTG---G GGCCG-CAAC A-GAACCAC GGCGCCGTAT  
 TCA---CGGA GGGCTT---C GGCCA-CAAA A-GAACCAC GGCGCCGTT  
 TCA---CGGA GGGCTT---C GGCCA-CAAA A-GAACCAC GGCGCCGTT  
 TCA---CGGA GGGCTT---C GGCCA-CAAA A-GAACCAC GGCGCCGTT  
 TCA---CGGA GGGCTT---C GGCCA-CAAA A-GAACCAC GGCGCCGTT  
 TCA---CGGA GGGCTT---C GGCCA-CAAA A-GAACCAC GGCGCCGTT  
 TCA---TGAA GGGGAA---G GGTCT-CAAA CAGAACCAC GGCGCTGGAT  
 TTG---TGG GGGGA---G GGTCCCAAC A-GAACCAC GGCGCCGGAT  
 TTA---T-GA CGGGTG---T GGTCCCAAA A-GAACCAC GACGTCGGAT

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IMPERATACYLINDRICA	GG-CGTCAAG	G-AACACTT-	---CTATTGC	C--TTGC-TC	GGCGGAGCGG
SORGHUMHALEPENSE	GG-CGTCAAG	G-AACACTC-	---ATGTTGC	C--TTGC-AC	AGCGGAGTGG
PANICUMLANIPES	GG-CGTCAAG	G-AACACTG-	---ATATTGC	C--TTGC-TT	GGGGCTTGT
PANICUMSTAFFIANUM	GG-CGTCAAG	G-AACACTG-	---ATATTGC	C--TTGC-TT	GGGGCTTGT
KARROOCHLOAPURPUREA6241	GG-CGTCAAG	G-AACACTT-	---ATATTGC	C--TTGC-GC	GCGGCGGTGG
CHAETOBROMUSINVOLUCRATES6284	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Merxmullerangei	GG-CGTCAAG	G-AACACCG-	---ATATTGC	C--TTGC-GA	GGCGCCGCGG
Centropodiaglauca	GG-CGTCAAG	G-AACACTA-	---GTATTGC	C--TCGC-GC	GCGGCCGCGG
Cynodondactylon5248	GG-CGTCAAG	G-AGAACTA-	---ATGTTGC	C--TTGC-TT	GGGGCCTCGG
Cynodondactylon7480	GG-CGTCAAG	G-AGAACTA-	---ATGTTGC	C--TTGC-TT	GGGGCCTCGG
Cynodondactylon	GG-CGTCAAG	G-AGAACTA-	---ATGTTGC	C--TTGC-TT	GGGGCCTCGG
Eleusinecoracana	GG-CGTCAAG	G-AAAACAT-	---ATGTTGC	C--TTGC-CT	GGGGCTGTGA
Eleusineindica	GG-CGTCAAG	G-AAAACAT-	---ATGTTGC	C--TTGC-CT	GGGGCTGTGA
Chlorisvirgata6616	GG-CGTCAAG	G-AAAACAT-	---ATGTTGC	C--TTGC-CT	GGGGCTGTGA
Chlorisvirgata7498	GG-CGTCAAG	G-AAAACAT-	---ATGTTGC	C--TTGC-CT	GGGGCTGTGA
Eustachyspaspaloides5597	GG-CGTCAAG	G-ATGACTT-	---ATATTGC	C--TTGC-TT	GGGGCCACGA
Harpochloafalx6530	GG-CGTCAAG	G-AAAAGTA-	---ATATTGA	C--TTGC-CT	GGGGCCACAA
Harpochloafalx5118	GG-CGTCAAG	G-AAAAGTA-	---ATATTGA	C--TTGC-CT	GGGGCCACAA
Harpochloafalx6670	GG-CGTCAAG	G-AAAAGTA-	---ATATTGA	C--TTGC-CT	GGGGCCACAA
Rendliaaltera5133	AG-CGTCAAG	G-AAAACAT-	---ATATTGC	C--TTGC-AT	GGGGCCATGG
Leptochloafusca7622	GG-CGTCAAG	G-AAAACAT-	---ATATTGC	C--TTGC-AT	GGGGCCATGG
Entopogonmacrostachyus9339	GG-CGTCAAG	G-AAAACAT-	---ATATTGC	C--TTGC-AT	GGGGCCATGG
Astreblalappacea	GG-CGTCAAG	G-AACACTA-	---AAATTGC	C--TTGC-TC	GGGATCACGA
Muhlenbergiarichardsonis	GG-CGTCAAG	G-AACACTT-	---GTATTG-	C--TTGC-TC	GGGGCAAAGA
Triodiastenostachya	GG-CGTCAAG	G-AACACTC-	---ATATTAC	C--TTGC-TC	GGGGCCACGA
Microchloacaffra5146	GG-CGTCAAG	G-AAAACAA-	---ATATTGC	C--TTGC-TT	GGGGCAGTGG
Dactylocteniumaegyptium9326	GG-CGTCAAG	GAGACCAAT-	---ATTTGCC	AATTTCG-	GGTGTAAATGT
Tragusberteronianus6620	GG-CGTCAAG	G-AACACTT-	---GTTTTGC	A--GTAC-TC	GGGGAACGA
Tragusracemosus7108	GG-CGTCAAG	G-AACACTT-	---GTTTTGC	A--GTAC-TC	GGGGAACGA
Traguskoelerioides6729	GG-CGTCAAG	G-AACACTT-	---GTTTTGC	A--GTAC-TC	AGAGAAACGA
Polevansiarigida7621	GG-CGTCAAG	G-AACACTT-	---GTTTTGC	A--TTGC-TT	GGGGAAGCTA
Perotispatens9323	GG-CGTCAAG	G-AACACAA-	---ATGTTGC	C--TTGC-TT	GGTGTAAATGT
Eragrostiscurvula6529	GG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCTCA	AGGATTGTGA
Eragrostisobtusa6623	GG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCCCA	AGGTC-GCGA
Eragrostisobtusa6732	GG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCCCA	AGGTC-GCGA
Eragrostisechinochloidea7485	GG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCCCA	AGGTC-GCGA
Eragrostispseudobtusa7508	GG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCCCA	AGGTC-GCGA
Eragrostisechinochloidea5933	GG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCCCA	AGGTC-GCGA
Eragrostislehmanniana20	GG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCTCA	AGGTTGTGA
Eragrostiscurvula5191	GG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCTCA	AGGTTGTGA
Eragrostischloromelas6947	TG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCTCA	AGGATTGTGA
Eragrostischloromelas7474	GG-CGTCAAG	G-AATACTA-	---ATATTGC	C--TTGCCCA	AG-TTCACGA
Eragrostisracemosa6533	GG-CGTCAAG	G-AACACTA-	---ATGTTGC	C--TTGCCCA	AG-TTCATGA
Eragrostisbiflora6705	GG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCCCA	AG-TTCACGA
Eragrostissuperba7470	GG-CGTCAAG	G-AACACTA-	---CTATTGC	C--TTGCCCA	AGG-TCCGGA
Eragrostiscapensis6551	GG-CGTCAAG	G-AACACTA-	---TTGTTGC	A--TTGCCCA	AGG-TCCGCT
Eragrostiscapensis6532	GG-CGTCAAG	G-AACACTA-	---TTGTTGC	A--TTGCCCA	AGG-TCCGCT
Pogonarthriasquarrosa7483	GG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCCCA	AGG-TCACGA
Pogonarthriasquarrosa7504	GG-CGTCAAG	G-AACATA-	---ATATTGC	C--TTGCCCA	AGG-TCACGA
Catalepisgracilis6962	GG-CGTCAAG	G-AATACCA-	---ATATTGC	C--TTGCCCA	AG-TTTATGA
Catalepisgracilis6653	GG-CGTCAAG	G-AATACCA-	---ATATTGC	C--TTGCCCA	AG-TTTATGA
Cladoraphiscyperoides5704	GG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCCTA	AGG-CCGTGT
Cladoraphisspinosa6335	GG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCCCA	AGG-CCGTGT
Cladoraphisspinosa4885	GG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCCCA	AGG-CCGTGT
Cladoraphiscyperoides4894	GG-CGTCAAG	G-AACACTA-	---ATATTGC	C--TTGCCTA	AGG-CCGTGT
Fingerhuthiaafricana4349	GG-CGTCAAG	G-AACACTT-	---CTATTGC	A--ATGCCCA	AG-ATTGTGA
Fingerhuthiaafricana7507	GG-CGTCAAG	G-AACACTT-	---CTATTGC	A--ATGCCCA	AG-ATTGTGA
Fingerhuthiaafricana4879	GG-CGTCAAG	G-AACACTT-	---CTATTGC	A--ATGCCCA	AG-ATTGTGA
Entoplocamiaaristulata9353	GG-CGTCAAG	G-AACACTA-	---TTATTGC	T--TTGCCCA	AG-ACCGTGA
Enneapogonscoparius18	GG-CGTCAAG	G-AACACTT-	---ATATTGC	C--TTGCCCA	AGG-TGCGG
Enneapogonenchroides7506	GG-CGTCAAG	G-AACACTT-	---ATATTGC	C--TTGCCCA	AGG-TGCGG
Enneapogonenchroides5213	GG-CGTCAAG	G-AACACTT-	---ATATTGC	C--TTGCCCA	AGG-TGCGG
Enneapogonscoparius7487	GG-CGTCAAG	G-AACACTT-	---ATATTGC	C--TTGCCCA	AGG-TGCGG
Schmidtiaappophoroides6334	GG-CGTCAAG	G-AACACTA-	---ATGTTGC	C--TTGCCCA	AAG-TGCGG
Schmidtiaakaliariensis7490	GG-CGTCAAG	G-AACACTG-	---ATATTGC	C--TTGCCCA	AGG-CTGCGG
Schmidtiaakaliariensis7486	GG-CGTCAAG	G-AACACTG-	---ATATTGC	C--TTGCCCA	AGG-CTGCGG
Sporobolusafricanus7476	GGGCGTCAAG	G-AACACTG-	---ATGTTGC	---TTGCTTG	GGG-TTTTGG
Sporobolusconsimilis9354	GG-CGTCAAG	G-AATACTA-	---ATGTTGC	---TTGCTCG	GGG-TTACGG
Sporobolusfimbriatus9328	GA-CGTCAAG	G-AACACTTT	TATTTATTGC	---TTGCACA	ATG-GTGTGA

IMPERATACYLINDRICA	TCCGGCTGCC	TTC--CGCTC	CCC-GCGCAG	CGAT-GATA-	-TCTTAATCC
SORGHUMHALEPENSE	TCCGGCATGCC	TTC--CGCTC	CCT-GAGCAG	CGAT-GATA-	-TCTTAATCC
PANICUMLANIPES	TCCGGCTGCC	CGG--CAAGC	CCC-GTGCAA	TGTT-GCTA-	-TCTTAATCA
PANICUMSTAFFIANUM	TCCGGCTGCC	GCG--CATGT	CCC-ATGCAA	TGTT-GCTA-	-TCCAAATCC
KARROOCHLOAPURPUREA6241	CCCGGCTGCC	GGA--CGCTC	CGT-GCGCAG	CGAT-TGTA-	-TACTAATCC
CHAETOBROMUSINVOLUCRATES6284	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Merxmullerangei	CCGGCTCGCC	GGA--CGCGG	CCC-GCGCAG	CGAT-GCTA-	-TCTTAATCC
Centropodiaglauca	CCGGCTGCC	GGG--CGCGC	CGC-GCGCGG	CGAT-GACA-	-CCTTAATCC
Cynodondactylon5248	CCGGCTGCC	GGA--TGCAC	CCC-TTGCAG	CGAT-GCTA-	-TGGAAAATC
Cynodondactylon7480	CCGGCTGCC	GGA--TGCAC	CCC-TTGCAG	CGAT-GCTA-	-TGGAAAATC
Cynodondactylon	CCGGCTGCC	GGA--TGCAC	CCC-TTGCAG	CGAT-GCTA-	-TGGAAAATC
Eleusinecoracana	TTGGCATGCC	CTA--TGTAC	CCC-GTGCAG	TGAT-GCAT-	-TGTTAAGTC
Eleusineindica	TTGGCATGCC	GTA--TGTAC	CCC-GTGCAG	TGAT-GCAT-	-TGTTAAGTC
Chlorisvirgata6616	TCCGGCTGCC	GGA--CACAC	CTC-TCGCAG	CGAT-GCTA-	-TGAAATTTT
Chlorisvirgata7498	TCCGGCTGCC	GGA--CACAC	CTC-TCGCAG	CGAT-GCTA-	-TGAAATTTT
Eustachyaspaloides5597	TCCGGCTGCC	GGA--CGTAC	TCC-TAGCAG	CGAT-GCKT-	-TGGAATTC
Harpochloafalx6530	CTGGCTTGCT	GGA--CGCAC	CCG-GTGCAG	TGAT-GCTA-	-TAAAAATTT
Harpochloafalx5118	CTGGCTTGCT	GGA--CGCAC	CCG-GTGCAG	TGAT-GCTA-	-TAAAAATTT
Harpochloafalx6670	CTGGCTTGCT	GGA--CGCAC	CCG-GTGCAG	TGAT-GCTA-	-TAAAAATTT
Rendliaaltera5133	CCGGCTTGCT	GGA--CATA	CCC-TTGCAG	CGAT-GCTA-	-TATAAATTC
Leptochloafusca7622	CCGGCTTGCT	GGA--TGCAC	CCC-CTGCAG	CGAT-GCTA-	-TGTAGATAC
Enteropogonmacrostachyus9339	CCGGCTTGCC	GGACGTGCAC	CCG--TGAC	AGATTGATA-	-TGGAAATTC
Astreblalappacea	CCGGCTTGCC	GGA--CGCAC	CCG--TGAC	CGAT-GCTA-	-TGTTAATTC
Muhlenbergiarichardsonis	TCCGGCTGCC	GAAC-GGAAC	CCC-GGACAA	CGAT-GATA-	-TGCTAATCA
Triodiastenostachya	TCCGGCTGCT	GGT--CATGC	CCC-GTGCAG	TGAT-GCTT-	-TGTTAATCC
Microchloacaffra5146	CCGGCTTGCT	GGA--CATA	CCT-TTGCAG	CGAT-TCTA-	-TGGAAATTC
Dactylocteniumaegyptium9326	CCGGCTAGCC	GGA--GAAC	CCC-GCGCAG	CGAT-GCTAN	NNNNNNNNNN
Tragusberteronianus6620	TCCGGCTGCC	GAA--CGAAC	CTC-GGGTTG	CACT-GATA-	---TTAATCA
Tragusracemosus7108	TCCGGCTGCC	GAA--CGAAC	CTC-GGGTTG	CACT-GATA-	---TTAATCA
Traguskoelerioides6729	TCCGGCTGCC	GAA--CGTWC	CTC-GGGTTG	CRCT-GATA-	---TTAATCA
Polevansiarigida7621	TCCGGCTGCC	GGA--CGCAC	CCC-AGGCTG	CATT-GATA-	---TTAATCA
Perotispates9323	CTGGCTTGCT	GGA--TGTGA	ATC-AAGCAT	TGAT-GCTAT	ATATTAAATCC
Eragrostiscurvula6529	CCGGCTTGCC	GGA--AACGT	CTT-GAGCAG	CGATACCTA-	-TCGTAATCC
Eragrostisobtusa6623	CCGGCTTGCC	GGA--CGTGT	CTT-GAGCAG	CGATACCTA-	-TCATAATCC
Eragrostisobtusa6732	CCGGCTTGCC	GGA--CGTGT	CTT-GAGCAG	CGATACCTA-	-TCATAATCC
Eragrostisechinocloidea7485	CCGGCTTGCC	GGA--CGTGT	CTT-GAGCAG	CGATACCTA-	-TCATAATCC
Eragrostispseudobtusa7508	CCGGCTTGCC	GGA--CGTGT	CTT-GAGCAG	CGATACCTA-	-TCATAATCC
Eragrostisechinocloidea5933	CCGGCTTGCC	GGA--CGTGT	CTT-GAGCAG	CGATACCTA-	-TCATAATCC
Eragrostislehmanniana20	CCGGCTTGCC	GGT--AACGT	CTTTGAGCAG	CGATACCTA-	-TTGTAATCC
Eragrostiscurvula5191	TCCGGCTTGCC	GGA--AACGT	CTT-GAGCAG	CGATACCTA-	-TCGTAATCC
Eragrostischloromelas6947	CCGGCTTGCC	GGA--AACAT	CTT-GAGCAG	CGATACCTA-	-TCGTAATCC
Eragrostischloromelas7474	TCCGGCTTGCC	GGA--CGCGT	CTT-GAGCGG	CGATACCTA-	-TCTTAATCC
Eragrostisracemosa6533	CCGGCTTGCC	GGA--CGTGT	CTT-GAGCGG	CGATACCTA-	-TTGTAATCC
Eragrostisbiflora6705	CCGGCTTGCC	GGA--CGCGA	CTT-GAGCGG	CGATACCTA-	-TCTTAATCC
Eragrostissuperba7470	TCCGGCTTGCC	GAG--CGCGT	CTT-GAGCAG	CGATACCTA-	-TTGTAATCC
Eragrostiscapensis6551	TTGGCTTGCC	GGA--CACGT	CTT-GAGCAG	CGATACCTA-	-CCATAATCC
Eragrostiscapensis6532	TTGGCTTGCC	GGA--CACGT	CTT-GAGCAG	CGATACCTA-	-CCATAATCC
Pogonarthriasquarrosa7483	ACGGCTTGTC	GGA--CGTGT	CTT-GAGCAG	CGATACATA-	-TCATAATCC
Pogonarthriasquarrosa7504	TCCGGCTTGCC	GGA--CGTGT	CTT-GAGCAG	CGATACATA-	-TCATAATCC
Catalepisgracilis6962	TCCGGCTTGCC	GGA--CAAGT	CTT-GAGCAG	CGATACCTA-	-TCTTAATCC
Catalepisgracilis6653	TCCGGCTTGCC	GGA--CAAGT	CTT-GAGCAG	CGATACCTA-	-TCTTAATCC
Cladoraphiscyperoides5704	TTGGCTTGCC	AAA--CGCGT	CTT-GGGCAG	CGATACCAA-	-TTATAATCC
Cladoraphisspinosa6335	TTGGCTTGCC	GGA--CGCGT	CTT-GGGCAG	CGATACCCA-	-TTATAATCC
Cladoraphisspinosa4885	TTGGCTTGCC	GGA--CGCGT	CTT-GGGCAG	CGATACCCA-	-TTATAATCC
Cladoraphiscyperoides4894	TTGGCTTGCC	AAA--CGCGT	CTT-GGGCAG	CGATACCAA-	-TTATAATCC
Fingerhuthiaafricana4349	TCCGGCTTGCC	GAA--CGCGT	CTT-GGGCAG	CGAT-GATA-	-TCTTAATCC
Fingerhuthiaafricana7507	TCCGGCTTGCC	GAA--CGCGT	CTT-GGGCAG	CGAT-GATA-	-TCTTAATCC
Fingerhuthiaafricana4879	TCCGGCTTGCC	GAA--CGCGT	CTT-GGGCAG	CGAT-GATA-	-TCTTAATCC
Entoplocamiaaristulata9353	TCCGGCTTGCC	GGA--CGCGT	CTT-GAGCAG	CGAT-GCTA-	-TCTTAATCC
Enneapogonscoparius18	TCCGGCTTGCC	GGA--CGCGC	CTT-GAGCAG	CGAT-GCTA-	-TGTTAATCC
Enneapogoncenchroides7506	TCCGGCTTGCC	GGA--CGCGT	CTT-GAGCAG	CGAT-GCTA-	-ATGTTAATCC
Enneapogoncenchroides5213	TCCGGCTTGCC	GGA--CGCGT	CTT-GAGCAG	CGAT-GCTA-	-TGTTAATCC
Enneapogonscoparius7487	TCCGGCTTGCC	GGA--CGCGC	CTT-GAGCAG	CGAT-GCTA-	-TGTTAATCC
Schmidtiaapappophoroides6334	TCCGGCTTGCC	GGA--CGCGT	TTG-GAGCAG	GGAT-GCTA-	-TCTTAASCR
Schmidtialihariensis7490	CCGGCTTGCC	GTT--CGCGT	CTT-GAGCAG	CGAT-GCTA-	-TCTCAATCC
Schmidtialihariensis7486	CCGGCTTGCC	GTT--CGCGT	CTT-GAGCAG	CGAT-GCTA-	-TCTCAATCC
Sporobolusafricanus7476	CCAGCCTGCT	GGC--AGATC	CCT-GAGCTG	CGAT-GGTA-	-CCTTAATCC
Sporobolusconsimilis9354	CCGGCTTGCT	GGA--CGAGC	CCT-GTGCAG	CGAT-GCTAT	ACCTGAATCC
Sporobolusfimbriatus9328	ACTTCTTGTT	TGA--TACAC	CTT-GTGCAG	TGAT-GCTA-	-CCTTAATCC

IMPERATACYLINDRICA	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
SORGHUMHALEPENSE	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
PANICUMLANIPES	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
PANICUMSTAPFIANUM	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
KARROOCHLOAPURPUREA6241	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGC
CHAETOBROMUSINVOLUCRATES6284	NNNNNNNNNN	NNGGAAGGA	TATTTAGTTT	TTTCGCATTGA	TGAAGATCGT
Merxmuellerrangei	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Centropodiaglauca	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Cynodondactylon5248	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTTGCATCGA	TGAAGAACGT
Cynodondactylon7480	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTTGCATCGA	TGAAGAACGT
Cynodondactylon	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTTGCATCGA	TGAAGAACGT
Eleusinecoracana	ACATGACTCT	CGGCAACGGA	TATCTTGGCT	CTCGCATCGA	TGAAGAACGT
Eleusineindica	ACATGACTCT	CGGCAACGGA	TATCTTGGCT	CTCGCATCGA	TGAAGAACGT
Chlorisvirgata6616	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTTGCATCGA	TGAAGAACGT
Chlorisvirgata7498	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTTGCATCGA	TGAAGAACGT
Eustachyspaspaloides5597	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTTGCATCGA	TGAAGAACGT
Harpochloafalx6530	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTTGCATCGA	TGAAGAACGT
Harpochloafalx5118	ACATGACTCT	TGGCAACGGA	TATCTCGGCT	CTTGCATCGA	TGAAGAACGT
Harpochloafalx6670	ACATGACTCT	TGGCAACGGA	TATCTCGGCT	CTTGCATCGA	TGAAGAACGT
Rendliaaltera5133	TCATGACTCT	CGGCAACGGA	TATCTTGGCT	CTTGCATCGA	TGAAGAACGT
Leptochloafusca7622	ACATGACTCT	CGGCAACGGA	TATCTTGGCT	CTCGCATCGA	TGAAGAACGT
Enteropogonmacrostachyus9339	ACATGACTCT	TGGCAACGGA	TATCTCGGCT	CTTGCATCGA	TGAAGAACGT
Astreblalappacea	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTTGCATCGA	TGAAGAACGT
Muhlenbergiarichardsonis	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Triodiastenostachya	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Microchloacaffra5146	ACATGACTCT	CGGCAACGGA	TATCTTGGCT	CTTGCATCGA	TGAAGAACGT
Dactylocteniumaegyptium9326	NNNNNNNNNN	NNGCAACGGA	TATCTTGGCT	CTCGCATCGA	TGAAGAACGT
Tragusberteronianus6620	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Tragusracemosus7108	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Traguskoelerioides6729	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Polevansiarigida7621	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Perotispatens9323	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAATAACGT
Eragrostiscurvula6529	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Eragrostisobtusa6623	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Eragrostisobtusa6732	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Eragrostisechinochloidea7485	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Eragrostispseudobtusa7508	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAASGT
Eragrostisechinochloidea5933	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Eragrostislehmanniana20	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Eragrostiscurvula5191	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Eragrostischloromelas6947	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Eragrostischloromelas7474	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Eragrostisracemosa6533	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAANNNNNN
Eragrostisbiflora6705	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Eragrostissuperba7470	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Eragrostiscapensis6551	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Eragrostiscapensis6532	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Pogonarthriasquarrosa7483	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Pogonarthriasquarrosa7504	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Catalepisgracilis6962	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAAAGT
Catalepisgracilis6653	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Cladoraphiscyperoides5704	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAANNNNNN
Cladoraphisspinosa6335	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Cladoraphisspinosa4885	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Cladoraphiscyperoides4894	ACATGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Fingerhuthiaafricana4349	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAAGGT
Fingerhuthiaafricana7507	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Fingerhuthiaafricana4879	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAAGGT
Entoplocamiaaristulata9353	ACACGACTCT	CGGCAACGGA	TATCTCGGCT	CTCGCATCGA	TGAAGAACGT
Enneapogonscoparius18	AAACGACTCT	CGGCAACGGA	TATCTTGGCT	CTCGCATCGA	TGAAGAACGT
Enneapogoncenchroides7506	AAACGACTCT	CGGCAACGGA	TATCTTGGCT	CTCGCATCGA	TGAAGAACGT
Enneapogoncenchroides5213	AAACGACTCT	CGGCAACGGA	TATCTTGGCT	CTCGCATCGA	TGAAGAACGT
Enneapogonscoparius7487	AAACGACTCT	CGGCAACGGA	TATCTTGGCT	CTCGCATCGA	TGAAGAACGT
Schmidtiaappophoroides6334	ACACGACTTT	TGGCAACGGA	TATCTTGGCT	CTCGCATCGA	TGAAGAACGT
Schmidtiaakalihariensis7490	ACACGACTCT	CGGCAACGGA	TATCTTGGCT	CTTGCATCGA	TGAAGACNNN
Schmidtiaakalihariensis7486	ACACGACTCT	CGGCAACGGA	TATCTTGGCT	CTTGCATCGA	TGAAGAACGT
Sporobolusafricanus7476	ATAAGACTCT	CGGCAACGGA	TATCTTGGCT	CTCGCATCGA	TGAAGAACGT
Sporobolusconsimilis9354	ATAAGACTCT	CGGCAACGGA	TATCTTGGCT	CTCGCATCGA	TGAANNNNNN
Sporobolusfimbriatus9328	ACAAGACTCT	CGGCAACGGA	TATCTTGGCT	CTCGCATCGA	TGAAGAACGT

IMPERATACYLINDRICA	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
SORGHUMHALEPENSE	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
PANICUMLANIPES	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
PANICUMSTAPFIANUM	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
KARROOCHLOAPURPUREA6241	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
CHAETOBROMUSINVOLUCRATES6284	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	GAACCATGAG
Merxmuelleraangei	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Centropodiaglauca	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Cynodondactylon5248	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Cynodondactylon7480	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Cynodondactylon	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Eleusinecoracana	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Eleusineindica	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Chlorisvirgata6616	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCAAG
Chlorisvirgata7498	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCAAG
Eustachyspaspaloides5597	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Harpochloafalx6530	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCAAG
Harpochloafalx5118	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCAAG
Harpochloafalx6670	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCAAG
Rendliaaltera5133	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Leptochloafusca7622	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Enteropogonmacrostachyus9339	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Astreblalappacea	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Muhlenbergiarichardsonis	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Triodiastenostachya	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCAAG
Microchloaaffra5146	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Dactylocteniumaegyptium9326	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Tragusberteronianus6620	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Tragusracemosus7108	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Traguskoelerioides6729	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Polevansiarigida7621	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Perotispatens9323	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Eragrostiscurvula6529	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Eragrostisobtusa6623	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Eragrostisobtusa6732	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Eragrostisechinochloidea7485	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Eragrostispseudobtusa7508	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Eragrostisechinochloidea5933	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Eragrostislehmanniana20	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Eragrostiscurvula5191	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Eragrostischloromelas6947	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Eragrostischloromelas7474	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Eragrostisracemosa6533	NNCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Eragrostisbiflora6705	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Eragrostissuperba7470	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Eragrostiscapensis6551	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Eragrostiscapensis6532	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Pogonarthriasquarrosa7483	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Pogonarthriasquarrosa7504	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Catalepisgracilis6962	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Catalepisgracilis6653	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Cladoraphiscyperoides5704	NNCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Cladoraphisspinosa6335	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Cladoraphisspinosa4885	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Cladoraphiscyperoides4894	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Fingerhuthiaafricana4349	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Fingerhuthiaafricana7507	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Fingerhuthiaafricana4879	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Entoplocamiaaristulata9353	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Enneapogonscoparius18	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Enneapogoncenchroides7506	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Enneapogoncenchroides5213	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Enneapogonscoparius7487	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Schmidtiaappophoroides6334	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGCG	AACCATCGAG
Schmidtiaakalihariensis7490	NNNNNNNNNN	NNNNNNCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Schmidtiaakalihariensis7486	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCATG	AACCATCGAG
Sporobolusafricanus7476	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG
Sporobolusconsimilis9354	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNATCCCGTG	AACCATCGAG
Sporobolusfimbriatus9328	AGCAAAATGC	GATACCTGGT	GTGAATTGCA	GAATCCCGTG	AACCATCGAG



IMPERATACYLINDRICA	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCAA	CCCACCCTC-	GGG-----
SORGHUMHALEPENSE	CTGGGCGTCA	CGCCAACAGA	CA-CTCCCAA	CCCACCCTC-	GGG-----
PANICUMLANIPESES	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCAA	CCCATCCTT-	GGG-----
PANICUMSTAFFIANUM	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCTAC	CCCATACTAT	GGG-----
KARROOCHLOAPURPUREA6241	CTGGGCGTCA	CGCCAAAAGA	CG-CTCCRC	CCTACCTTT-	GGG-----
CHAETOBROMUSINVOLUCRATES6284	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCGAA	CCCCCAA--	CGCAAGTTGT
Merxmuellearangei	CTGGGCGTCA	CGCCAAAAGA	CG-CTCCGC	CCACCCTC--	GGC-----
Centropodiaglauca	CTGGGCGTCA	CGCCAAAAGA	CG-CTCCAC	CCCACCCC--	GGT-----
Cynodondactylon5248	CTGGGCGTCA	CGCCAAAAGA	CA-CT-CCAC	CGCAATCCC-	GGT-----
Cynodondactylon7480	CTGGGCGTCA	CGCCAAAAGA	CA-CT-CCAC	CGCAATCCC-	GGT-----
Cynodondactylon	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCAC	CGCAATCCC-	GGT-----
Eleusinecoracana	CTGGGCGTCA	CGTCAAAAAGA	CA-CTCCCTA	CCATTCCTT-	GGT-----
Eleusineindica	CTGGGCGTCA	CGTCAAAAAGA	CA-CTCCCTA	CCATTCCTT-	GGT-----
Chlorisvirgata6616	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCACA	CCAACCTC--	GGT-----
Chlorisvirgata7498	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCACA	CCAACCTC--	GGT-----
Eustachyspaspaloides5597	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCATA	CCCAACCT--	GGT-----
Harpochloafalx530	TTGGGCGTCA	CGCCAAAAGA	CA-CTCCACA	CCTGTCTC--	GGT-----
Harpochloafalx5118	TTGGGCGTCA	CACCAAAAAGA	CA-CTCCACA	CCTGTCTC--	GGT-----
Harpochloafalx6670	TTGGGCGTCA	CACCAAAAAGA	CA-CTCCACA	CCTGTCTC--	GGT-----
Rendliaalferusa5133	CTGGGCGTCA	TGTGAAAAGA	CA-CTCCACA	CCAACTC--	AGT-----
Leptochloafusca7622	CTGGGCGTCA	CGCCAAAAGA	CA-CTCTAGA	CCTAACCT--	GGT-----
Enteropogonmacrostachyus9339	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCAAA	CCAATCCC--	GGT-----
Astreblalappacea	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCACA	TCAATCCT--	GGT-----
Muhlenbergiarichardsonis	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCAACTC--	AGT-----
Triodiastenostachya	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCCTC-	GGT-----
Microchloacaffra5146	CTGGGCGTCA	CGTCAAAAAGA	CA-CTCCACA	CCAAACTT--	GGT-----
Dactylocteniumaegyptium9326	CTGGGCGTCA	CGCCAGTATA	CAACTCCCCA	CCCATCCC--	GGT-----
Tragusberteronianus6620	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCAATCCT--	GGT-----
Tragusracemosus7108	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCAATCCT--	GGT-----
Traguskoelerioides6729	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCAATCCT--	GGT-----
Polevansirigida7621	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCAAACTT--	GGT-----
Perotispatens9323	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCTA	CCCAATTT--	GGT-----
Eragrostiscurvula6529	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCAT--	GGT-----
Eragrostisobtusa6623	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCAATCAT--	GGT-----
Eragrostisobtusa6732	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCAATCAT--	GGT-----
Eragrostisochinochloidea7485	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCAATCAT--	GGT-----
Eragrostispseudobtusa7508	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCAATCAT--	GGT-----
Eragrostisechinochloidea5933	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCAATCAT--	GGT-----
Eragrostislehmanniana20	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCAT--	GGT-----
Eragrostiscurvula5191	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCAACTC--	GGT-----
Eragrostischloromelas6947	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCAT--	GGT-----
Eragrostischloromelas7474	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCAT--	GGT-----
Eragrostisracemosa6533	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCAT--	GGT-----
Eragrostisbiflora6705	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCAT--	GGT-----
Eragrostissuperba7470	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCTA	CCCATCGT--	GGT-----
Eragrostiscapensis6551	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCACA	CCCATCGT--	GGT-----
Eragrostiscapensis6532	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCACA	CCCATCGT--	GGT-----
Pogonarthriasquarrosa7483	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCAT--	GGT-----
Pogonarthriasquarrosa7504	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCAT--	GGT-----
Catalepisgracilis6962	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCAT--	GGT-----
Catalepisgracilis6653	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCAT--	GGT-----
Cladoraphiscyperoides5704	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCAACTC--	GGT-----
Cladoraphisspinosa6335	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCAACTC--	GGT-----
Cladoraphisspinosa4885	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCAACTC--	GGT-----
Cladoraphiscyperoides4894	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCAACTC--	GGT-----
Fingerhuthiaafricana4349	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCGC--	GGT-----
Fingerhuthiaafricana7507	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCGC--	GGT-----
Fingerhuthiaafricana4879	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCGC--	GGT-----
Entoplocamiaaristulata9353	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCAAA	CCCATCCC--	GGT-----
Enneapogoncoparius18	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATACA--	GGT-----
Enneapogoncenchroides7506	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATACC--	GGT-----
Enneapogoncenchroides5213	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATACC--	GGT-----
Enneapogoncoparius7487	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATACA--	GGT-----
Schmidtiaappophoroides6334	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCAACTC--	GGT-----
Schmidtiaakalihariensis7490	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCTC--	GGT-----
Schmidtiaakalihariensis7486	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCA	CCCATCTC--	GGT-----
Sporobolusafricanus7476	CTGGGCGTCA	CGCCAAAGTGA	CA-CTTCACT	ACCATTCT--	GGT-----
Sporobolusconsimilis9354	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCTC	GCCATCCT--	GGT-----
Sporobolusfimbriatus9328	CTGGGCGTCA	CGCCAAAAGA	CA-CTCCCCC	ACCCTTCA-T	GGA-----

IMPERATACYLINDRICA	-----GAGG	GACGTGGTGT	TTGGACCCCC	GCG----	CCG	-CAGGGCGCG
SORGHUMHALEPENSE	-----GAGG	GATGTGGTTT	TTGGCTCCCC	GTG----	CCG	-CATGGCGCG
PANICUMLANIPES	-----TACG	GACGTGGTGT	TTGGCTCCCC	ATG----	CCT	-CATGGTGTG
PANICUMSTAPFIANUM	-----GTACG	GATGTGGTGT	TTGGCTCCCC	TTG----	CCT	-TGCGGCATG
KARROOCHLOAPURPUREA6241	-----TGAG	GATGCGGTGT	ATGGCTCCTC	GTG----	CCG	--TGAGGCGC
CHAETOBROMUSINVOLUCRATES6284	AACGGGAACG	GACGCGGCAT	ATGGCCCCCC	GTG----	TCC	GCAAGGCGCG
Merxmuelleraangei	-----GAG	GACGCGCGT	TTGGCTCCCC	GCG----	CCC	GCGGGGCGCG
Centropodiaglauca	-----GCG	GACGCGGTGT	CTGGCCCCCC	GTG----	CC-	GCAGGGGCGCG
Cynodondactylon5248	-----CTG	GACGTGGTGT	TTGGCCCCCTC	ATT----	CCA	TAGTGT-ATG
Cynodondactylon7480	-----CTG	GACGTGGTGT	TTGGCCCCCTC	ATT----	CCA	TAGTGT-ATG
Cynodondactylon	-----CTG	GACGTGGTGT	TTGGCCCCCTC	ATT----	CCA	TAGTGT-ATG
Eleusinecoracana	-----GTG	GACGTGGA-T	TTGGCTCCTC	ATG----	CCT	TAGGGCGTGG
Eleusineindica	-----GTG	GACGTGGA-T	TTGGCTCCTC	ATG----	CCT	TCGGGCGTGG
Chlorisvirgata6616	-----ATG	GACGTGGTGT	TTGGCCCCCC	ATA----	CCA	CATGGTTATG
Chlorisvirgata7498	-----ATG	GAYTGGTGT	TTGGCCCCCC	ATA----	CCA	CATGGTTATG
Eustachysaspaloides5597	-----ATG	GATGTGGTGT	TTGGCCCCCTC	ATC----	CCA	TAGGT-ATG
Harpochloafalx6530	-----TTG	GATGTGGTGC	ATGGCCCCCTC	ATA----	CCA	TCGGGTT-TG
Harpochloafalx5118	-----TTG	GATGTGGTGC	ATGGCCCCCTC	ATA----	CCA	TCGGGTT-TG
Harpochloafalx6670	-----TTG	GATGTGGTGC	ATGGCCCCCTC	ATA----	CCA	TCGGGTT-TG
Rendliaaltera5133	-----CTG	GACGTGGTAT	TTGGCCCCCTC	ATC----	CTA	TAAGGT-ATG
Leptochloafusa7622	-----GTG	GACGTGGTGT	TTGGCCCCCTC	ATG----	CCG	CAGGGT-GTG
Enteropogonmacrostachyus9339	-----CTG	GACGTGGTGT	TTGGCCCCCTC	ATG----	CCC	CAGGGT-GTG
Astreblalappacea	-----GTG	GACGAGGTGT	TTGGCTCTC	ATG----	CCG	CAGGGT-GTG
Muhlenbergiarichardsonis	-----GGG	GACGTGGTAT	TTGGCTCCTC	GTG----	TCG	TTATGC-ACG
Triodiastenostachya	-----GAG	GACGTGGAGT	CTGGCTCCTC	GTG----	CCG	---AGC-ACG
Microchloacaffra5146	----GGTCAG	GATGTGGTGT	TTGGCCCCCTC	ATC----	CTA	AATGGGTATG
Dactylocteniumaegyptium9326	-----GCG	GACGTGGTGT	CCGGCCCCCTC	GTG----	CCT	CCGGGC-GCG
Tragusberteronianus6620	-----GTG	GATGTGGTAT	TTGGCCCCCTC	GTT----	CCT	TATGGC-ACG
Tragusracemosus7108	-----GTG	GATGTGGTAT	TTGGCCCCCTC	RTT----	CCT	TATGGC-ACG
Traguskoelerioides6729	-----GTG	GATGTGGTAT	TTGGCCCCCTC	GTT----	CCT	TATGGC-ACG
Polevansiarigida7621	-----GAG	GACGTGGTAT	TTGGCCCCCTC	GTT----	CCG	TATGGC-RCG
Perotispatens9323	-----GGTG	GATGTGGTGT	TTGGCTCCTC	GTG----	CCG	TC-GGT-GCG
Eragrostiscurvula6529	-----GTG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCA	CATGGT-GCG
Eragrostisobtusa6623	-----GCG	GATGTGGCAT	TTGGCTCTCC	GTG----	CCA	TACGGC-GCG
Eragrostisobtusa6732	-----GCG	GATGTGGCAT	TTGGCTCTCC	GTG----	CCA	TACGGC-GCG
Eragrostisechinochloidea7485	-----GCG	GATGTGGCAT	TTGGCTCCCC	GTG----	CCA	CACGGC-GCG
Eragrostispseudobtusa7508	-----GCG	GATGTGGCAT	TTGGCTCTCC	GTG----	CCA	TACGGC-GCG
Eragrostisechinochloidea5933	-----GCG	GATGTGGCAT	TTGGCTCCCC	GTG----	CCA	CACGGC-GCG
Eragrostislehmanniana20	-----GTG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCA	CATGGT-GCG
Eragrostiscurvula5191	-----GTG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCA	CATGGT-GCG
Eragrostischloromelas6947	-----GTG	GACGTGGCAT	TTGGCTCYCC	GTG----	CCA	CATGGT-GCG
Eragrostischloromelas7474	-----GTG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCT	CACGGT-GCG
Eragrostisracemosa6533	-----GTG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCA	CACGGG-GCG
Eragrostisbiflora6705	-----GTG	GATGTGGCAT	TTGGCTCCCC	GTG----	CCT	CACGGT-GCG
Eragrostissuperba7470	-----GTG	GATGTGGCAT	TTGGCTCCCC	GTG----	CCA	CATGGT-GCG
Eragrostiscapensis6551	-----GCG	GATGTGGCAT	TTGGCTCCCC	GTG----	CYG	CATGGT-GCG
Eragrostiscapensis6532	-----GCG	GATGTGGCAT	TTGGCTCCCC	GTG----	CTG	CATGGT-GCG
Pogonarthriasquarrosa7483	-----GCG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCT	CACGGT-GCG
Pogonarthriasquarrosa7504	-----GCG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCT	CACGGT-GCG
Catalepisgracilis6962	-----GTG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCT	CACGGT-GCG
Catalepisgracilis6653	-----GTG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCT	CACGGT-GCG
Cladoraphiscyperoides5704	-----GTG	GATGTGGCAT	TTGGCTCCCC	GTG----	CCG	CACGGT-GCG
Cladoraphisspinosa6335	-----GCG	GATGTGGCAT	TTGGCTCCCC	GTG----	CCG	CACGGT-GCG
Cladoraphisspinosa4885	-----GCG	GATGTGGCAT	TTGGCTCYCC	GTG----	CCG	CACGGT-GCG
Cladoraphiscyperoides4894	-----GTG	GATGTGGCAT	TTGGCTCCCC	GTG----	CCG	CACGGT-GCG
Fingerhuthiaafricana4349	-----GCG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCA	AACGGC-GCG
Fingerhuthiaafricana7507	-----GCG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCA	AACGGC-GCG
Fingerhuthiaafricana4879	-----GCG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCA	AACGGC-GCG
Entoplocamiaaristulata9353	-----GCG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCA	AACGGC-ACG
Enneapogonscoparius18	-----GTG	GACGTGGTGT	TTGGCTCCCC	GTG----	CCA	CAAGG-GCG
Enneapogoncenchroides7506	-----GTG	GACGTGGTGT	TTGGCTCCCC	GTG----	CCA	CGAGG-GCG
Enneapogoncenchroides5213	-----GTG	GACGTGGTGT	TTGGCTCCCC	GTG----	CCA	CGAGG-GCG
Enneapogonscoparius7487	-----GTG	GACGTGGTGT	TTGGCTCCCC	GTG----	CCA	CAAGG-GCG
Schmidtiaappophoroides6334	-----GAG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCA	CATGG-ATG
Schmidtiaakalihariensis7490	-----GCG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCA	CATGGT-GCG
Schmidtiaakalihariensis7486	-----GCG	GACGTGGCAT	TTGGCTCCCC	GTG----	CCA	CATGGT-GCG
Sporobolusafricanus7476	-----ATTG	GATGTGGATT	CTGACCTTCC	ATGCC--	TGA	AAGGGC-GTG
Sporobolusconsimilis9354	-----GCTG	GATGTGGCGT	CTGGTCTTCC	ATGCCACTAG	AAAGGA	-GTG
Sporobolusfimbriatus9328	-----GCGG	GATGTGGTGT	TTGGCTTCC	GTGTCATGTG	AATGGT	-TCG



IMPERATACYLINDRICA	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCGAA	CCGCGCCGGG	CAC-AGCACG
SORGHUMHALEPENSE	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCGAT	TCGTGTCCGG	CAC-AGCACG
PANICUMLANIPES	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCATA	CCGTGCCGGG	CACCTGCATG
PANICUMSTAFFIANUM	GTGGGCCGAA	-GTT-GTGGC	TGCCGGCATA	CCGTGTCCGG	CACCTGCATG
KARROOCHLOAPURPUREA6241	GTGGGCCGAA	-GTT-GCGGC	TGCCGGCGTA	CCGTGCCGGG	CAC-AGCACG
CHAETOBROMUSINVOLUCRATES6284	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCGCA	CCCGGCTGGG	CAC-AGCACG
Merxmuellearangei	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCGTA	CCGTGCCGGG	CAC-AGCACG
Centropodiaglauca	GCGGGCCGAA	-GTT-GGGGC	TGCCGGCGTA	CCGTGCCGGG	CAC-AGCACG
Cynodondactylon5248	GTGGGCCAAA	-GAG-GCGGC	TGCCGGCG--	--GTGCCGGT	CAC-AGCAGA
Cynodondactylon7480	GTGGGCCAAA	-GAG-GCGGC	TGCCGGCG--	--GTGCCGGT	CAC-AGCAGA
Cynodondactylon	GTGGGCCAAA	-GAG-GAGGC	TGCCGGCG--	--GTGCCGGT	CAC-AGCAGA
Eleusinecoracana	TGGACCAAAG	-ATG-GAG-C	TGCCGGTT--	--GTGCCGAT	CGC-AGCACG
Eleusineindica	TGGACCAAAG	-ATG-GAG-C	TGCCGGTT--	--GTGCCGAT	CGC-AGCACG
Chlorisvirgata6616	ATGGGCTTAA	-GAT-GTGGC	TATCGGCA--	--GGTGCCGAT	CAG-AGCACG
Chlorisvirgata7498	ATGGGCTTAA	-GAT-GTGGC	TATCGGCA--	--GGTGCCGAT	CAK-AGCACG
Eustachyspaspaloides5597	ATGGGCCAAA	-GAT-GCGGC	TGCCGGT---	--GGTGCTGAT	CAC-AGCACG
Harpochoafalx6530	ATGGGCCAAA	-GTT-GCGGA	TGGCGGC---	--GGTGCTGAA	CAC-AGCACG
Harpochoafalx5118	ATGGGCCAAA	-GTT-GCGGA	TGGCGGC---	--GGTGCTGAA	CAC-AGCACG
Harpochoafalx6670	ATGGGCCAAA	-GTT-GCGGA	TGGCGGC---	--GGTGCTGAA	CAC-AGCACG
Rendliaaltera5133	ATGGGCCAAA	-AAT-GTGGC	TGCCGGT---	--GGTGCCGAT	CAC-AGCACG
Leptochloafusca7622	GTGGGCCAAA	-TTT-GGGGC	TGCCGGT---	--GGTGCCGAT	CAC-AGCACG
Enteropogonmacrostachyus9339	GTAGGCCAAA	AGAT-GGGGC	TGCCGGC---	--GGTGCCGAT	CAC-AGCACG
Astreblalappacea	GTGGGCCAAA	-GAC-GGGGC	TGCCGGT---	--GGTGCTGAT	CAC-AGCACG
Muhlenbergiarichardsonis	GTGGGCCAAA	-GTT-GGGGC	TGCCGGC---	--GGTGCCGAT	CAC-AGCACG
Triodiastenostachya	GTGAGCCAAA	-GTT-TGGCC	TGCCGAC---	--GGTGCCGAT	CAC-AGCACG
Microchloacaffra5146	GTGGGCCAAA	-GAT-GCGGC	TGGCGGC---	--AGTGCCGAT	CAC-AGCACG
Dactylocteniumaegyptium9326	GTGGGCCAAA	-GTT-GGGGC	TGCCGGC---	--CGTGCCGAT	CAC-AGCACG
Tragusberteronianus6620	GTGGGCCGAA	-GTT-AGGGC	TGCCGGC---	--GGTGCCGAT	CAC-AGCACG
Tragusracemosus7108	GTGGGCCGAA	-GTT-ACGGC	TGCCGGC---	--GGTGCCGAT	CAC-AGCACG
Traguskoelerioides6729	GTGGGCCGAA	-GTT-AGGGC	TGCCGGC---	--GGTGCCGAT	CAC-AGCACG
Polevansiarigida7621	GTGGGCCGAA	-GTT-AGGGC	TGCCGGC---	--GGTGCCGAT	CAC-AGCACG
Perotispatens9323	GTGGGCCAAA	-GTA-GGGGA	TACCGGC---	--AGTGCATCT	CAC-AGCACG
Eragrostiscurvula6529	GTGGGCCGAA	-GTT-GGGGC	TGCCGGTTTA	CGATACCGGT	CAC-AGCACG
Eragrostisobtusata6623	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	AGGTACCGGT	CAC-AGCACG
Eragrostisobtusata6732	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	AGGTACCGGT	CAC-AGCACG
Eragrostisechinochloidea7485	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	AGSTACCGGT	CAC-AGCACG
Eragrostispseudobtusata7508	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	AGGTACCGGT	CAC-AGCACG
Eragrostisechinochloidea5933	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	AGSTACCGGT	CAC-AGCACG
Eragrostislehmanniana2C	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	CGATACCGGT	CAC-AGCACG
Eragrostiscurvula5191	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	CGATACCGGT	CAC-AGCACG
Eragrostischloromelas6947	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	CGATACCGGT	CAC-AGCACG
Eragrostischloromelas7474	GTGGGCCGAA	-GAT-GGGGC	TGCCGGCTTA	TGGTGCCGGT	CAC-AGCACG
Eragrostisracemosa6533	GTGGG-CCAA	-ATT-GGGCT	GCCCGCTTA	CGGGGCCGGT	CAC-ATCACG
Eragrostisbiflora6705	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	CGGTGTCCGGT	CAC-AGCACG
Eragrostissuperba7470	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	TGGTACCGGT	CAC-AGCACG
Eragrostiscapensis6551	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	TGGTACCGGT	CAC-AGCACG
Eragrostiscapensis6532	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	TGGTACCG-T	CAC-AGCACT
Pogonarthriasquarrosa7483	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	CGGTACCGGT	CAC-AGCACG
Pogonarthriasquarrosa7504	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	CGGTACCGGT	CAC-AGCACG
Catalepisgracilis6962	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	AGGTGCCGGT	CAC-AGCACG
Catalepisgracilis6653	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	AGGTGCCGGT	CAC-AGCACG
Cladoraphiscyperoides5704	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	CGGTACCGGT	CAC-AGCACG
Cladoraphis spinosa6335	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	CGGTACCGGT	CAC-AGCACG
Cladoraphis spinosa4885	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	CGGTACCGGT	CAC-AGCACG
Cladoraphiscyperoides4894	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	CGGTACCGGT	CAC-AGCACG
Fingerhuthiaafricana4349	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	TGGTACCGGT	CAC-AGCACG
Fingerhuthiaafricana7507	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	TGGTACCGGT	CAC-AGCACG
Fingerhuthiaafricana4879	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	TGGTACCGGT	CAC-AGCACG
Entollocamia aristulata9353	GTGGGCCGAA	-GTT-GGGGC	TGCCGGCTTA	TGGTACCGGT	CAC-AGCACG
Enneapogon scoparius18	GTGGGCCGAA	-GTT-GGGGC	TGCCGGTGAA	AGGTACCGGT	CAC-AGCGCA
Enneapogon cenchroides7506	GTGGGCCGAA	-GTT-GGGGC	TGCCGGTGAA	AGGTACCGGT	CAC-AGCGCA
Enneapogon cenchroides5213	GTGGGCCGAA	-GTT-GGGGC	TGCCGGTGAA	AGGTACCGGT	CAC-AGCGCA
Enneapogon scoparius7487	GTGGGCCGAA	-GTT-GGGGC	TGCCGGTGAA	AGGTACCGGT	CAC-AGCGCA
Schmidtia pappophoroides6334	GTGGGCCGAA	-GTT-GGGGC	TGCCGGTGAA	AGGTACCGGT	CAG-AGCACG
Schmidtia kalihariensis7490	GTGGGCCGAA	-GTT-GGGGC	TGCCGGTGAA	AGGTACCGGT	CAC-AGCACG
Schmidtia kalihariensis7486	GTGGGCCGAA	-GTT-GGGGC	TGCCGGTGAA	AGGTACCGGT	CAC-AGCACG
Sporobolus africanus7476	GTAGGCTAAA	GTAAGAGGC	TGCT-GC-GA	TGGTGCTAGT	CAC-AGCACT
Sporobolus consimilis9354	GTAGGCTAAA	GTAAGAGGC	TGCTGGC-TA	TGGTGCCGGT	CAC-AGCACT
Sporobolus fimbriatus9328	GTGACTAAA	-GTAT-AGGC	TGCTAGC-TA	TGGGGCTGGT	CAT-AGCACT



	551			600
IMPERATACYLINDRICA	TGGTGGGCGA	CA-TCAGTTG	TTCTCGGTGC	A-GCGCCCN NNNNNNNNN
SORGHUMHALEPENSE	TGGTGGGCGA	CACTTAGTTG	TTCTCGGTGC	A-GCGCCCN NNNNNNNNN
PANICUMLANIPES	TGGTGGGCGA	CA-TAAGTTG	TTCTCGGTGC	A--GTGTCC-- --TGTCACG-
PANICUMSTAPFIANUM	TGGTGGGCGA	CA-TTAGTTG	TTCTCGGTGC	A--GTGTCC-- --TGACACG-
KARROOCHLOAPURPUREA6241	TGGTGGTCTGA	CAAAAGTTG	TCA-CGGTGT	T--GTGCCTC G-GTGCCTA-
CHAETOBROMUSINVOLUCRATES6284	AGGTGGGCGT	AACA--TCCC	TTCAACGTGT	M--GTGCCTC G-GCCGGTAG
Merxmullerangei	TGGTGGGCGA	CACAA-GTTG	TTCTCGGTGC	T--GCGCTC G-GCCCGCAG
Centropodiaglauca	TGGTGGGCGA	CACGA-GTTG	TTCTCGGTGC	T--GTGCCTC G-GACCAGCAG
Cynodondactylon5248	TGGTGGATGG	CACAA-GTTA	TTCTCGGTGT	T--GTGATCC T-GACAAGTC
Cynodondactylon7480	TGGTGGATGG	CACAA-GTTA	TTCTCGGTGT	T--GTGATCC T-GACAAGTC
Cynodondactylon	TGGTGGATGG	CACAA-GTTA	TTCTCGGTGT	T--GTGATCC T-GACAAGTC
Eleusinecoracana	AGGTGGATGA	CGAAT-GTTG	TTCTCACTGC	T--TTGATCG A-AACAGCTC
Eleusineindica	AGGTGGATGA	CGAAT-GTTG	TTCTCACTGC	T--TTGATCG A-AACAGCTC
Chlorisvirgata6616	AGGTGGATGA	CACAT-GTTA	TTCTCGGTGT	T--ATGATCT G-GAGAAGTC
Chlorisvirgata7498	AGGTGGATGA	CACAT-GTTA	TTCTCGGTGT	T--ATGATCT G-GAGAAGTC
Eustachyspaspaloides5597	TGGTGGATAA	CACAA-GTTA	TTCTCGGTGT	TTTATTGAAT --CATGACAC
Harpochloafalx6530	TGGTGGATGA	CACAT-GTTA	TTCTCGGTGT	TTT----CGT T--CGGACAC
Harpochloafalx5118	TGGTGGATGA	CACAT-GTTA	TTCTCGGTGT	TTT----CGT T--CGGACAC
Harpochloafalx6670	TGGTGGATGA	CACAT-GTTA	TTCTCGGTGT	TTT----CGT T--CGGACAC
Rendliaaltera5133	TGGTGGATGA	CACAA-GTTA	TTCTCGGTGC	T--ATTATCT G-GACAAGTC
Leptochloafusca7622	AGGTGGATGA	CGCAA-GTTG	TTCTCGGTGT	T--ATGATCC G-GACCACTC
Enteropogonmacrostachyus9339	AGGTGGATGA	CGCAA-GTTG	TTCTCGGTGT	T--AGGATCC G-GTGCACTA
Astrelblappacea	AGGTGGATGA	CGCAA-GTTG	TTCTCGGTGT	T--ACGATCT G-GACCACTC
Muhlenbergiarichardsonis	AGGTGGATGA	CGCAA-GTTG	TTCTCGGTGC	T--ATGATCC G-AGCCGTGT
Triodiastenostachya	AGGTGGATGA	CTCGT-GTCG	TTCTCGATGC	T--ATGATCC G-GACCGTA
Microchloacaffra5146	TGGTGGATGA	CGCAA-GTTA	TTCTCAGTGT	T--ATGATCT G-GACAAGTC
Dactylocteniumaegyptium9326	AGGTGGATGA	CGCAT-GTTG	TTCTCAGTGC	T--AAAATCG G-TACAGCTT
Tragusberteronianus6620	TGGTGGATAA	CGCGA-GTTG	TTCTCGGTGC	T--ATGATCC G-GACTCCTT
Tragusracemosus7108	TGGNNNNNNN	NNNNNNNNN	NNNNNNNNN	NNNNNNNNN
Traguskoelerioides6729	TGGTGGATGA	CGCGA-GTTG	TTCTCGGTGC	T--ATGATCC G-GACTCCTT
Polevansiarigida7621	TGGTG-ATGA	CTTTT-GTTG	TTCTCGGTGC	T--ATGGTTC G-GATGCCTT
Perotispatens9323	AGGTGGATGG	CACAG-GTTA	TTCTCGGTGC	T--GTGATGC TGTATTTGAT
Eragrostiscurvula6529	AGGTGGGTGA	CACAT-AGTG	TTCTCGCTGC	T--GTGGCT G-GATCATTG
Eragrostisobtusa6623	AGGTGGGTGA	CACAT-AGTG	TTCTCGCTGT	T--GTGGCT G-GACTATTG
Eragrostisobtusa6732	AGGTGGGTGA	CACAT-AGTG	TTCTCGCTGT	T--GTGGCT G-GACTATTG
Eragrostisechinochloidea7485	AGGTGGGTGA	CACGT-AGTG	TTCTCGCTGT	T--GTGGCT G-AGCTATTG
Eragrostispseudobtusa7508	AGGTGGGTGA	CACAT-AGTG	TTCTCGCTGT	T--GTGGCT G-GACTATTG
Eragrostisechinochloidea5933	AGGTGGGTGA	CACGT-AGTG	TTCTCGCTGT	T--GTGGCT G-AGCTATTG
Eragrostislehmanniana20	AGGTGGGTGA	CACAT-AGTG	TTCTCGCTGT	T--GTGGCT G-GACTATTG
Eragrostiscurvula5191	AGGTGGGTGA	CACAT-AGTG	TTCTCGCTGC	T--GTGGCT G-GACTATTG
Eragrostischloromelas6947	AGGTGGGTGA	CACAT-AGTG	TTCTCGCTGC	T--GTGGCT G-GACTATTG
Eragrostischloromelas7474	AGGTGGGTGA	CACTC-GGTG	TTCTCGCTGC	T--GTGACTC T-AACCGT-G
Eragrostisracemosa6533	AGGGGGTGA	CACAC-GGTG	TTCTCGCTGC	T--GTGACTC G-AATCGTTG
Eragrostisbiflora6705	AGGTGGGTGA	CACAT-GGTG	TTCTCGCTGC	T--GTGACTC G-AACCGT-G
Eragrostissuperba7470	AGGTGGGTGA	CACAT-AGTG	CTCTCGCTGC	T--GTGGCT G-GATCAATG
Eragrostiscapensis6551	AGGTGGGTGA	CACAC-GGTG	TTCTCGCTGT	T--GTGACTC C-TACCGCTG
Eragrostiscapensis6532	TGGTGGACAA	CATGC-GTTG	TTCTCGGAGT	T--GTGACTC G-TACTGCTT
Pogonarthriasquarrosa7483	AGGTGGGTGA	CACAT-GGTG	TTCTCGCTGC	T--GTGACTC G-GACCCTG
Pogonarthriasquarrosa7504	AGGTGGGTGA	CACAT-GGTG	TTCTCGCTGC	T--GTGACTC G-GACCCTG
Catalepisgracilis6962	AGGTGGGTGA	CACTC-GGTG	TTYTCGCTGC	T--GTGACTC G-AACTGT-G
Catalepisgracilis6653	AGGTGGGTGA	CACTC-GGTG	TTCTCGCTGC	T--GTGACTC G-AACTGT-G
Cladoraphiscyperoides5704	AGGTGGGTGA	CACAT-GTTG	TTCTCGCTGC	T--GTGACTC G-GCCGCT-G
Cladoraphisspinosa6335	AGGTGGGTGA	CACAT-GTTG	TTCTCGCTGC	T--GTGACTC G-GCCGCT-G
Cladoraphisspinosa4885	AGGTGGGTGA	CACAT-GTTG	TTCTCGCTGC	T--GTGACTC G-GCCGCT-G
Cladoraphiscyperoides4894	AGGTGGGTGA	CACAT-GTTG	TTCTCGCTGC	T--GTGACTC G-GCCGCT-G
Fingerhuthiaafricana4349	AGGTGGGTGA	CACAT-GATG	TTCTCGCTGC	T--GCGACAC G-GACCCTT-
Fingerhuthiaafricana7567	AGGTGGGTGA	CACAT-GWTG	TTCTCGCTGC	T--GCGACAC G-GACCCTT-
Fingerhuthiaafricana4879	AGGTGGGTGA	CACAT-GATG	TTCTCGCTGC	T--GCGACAC G-GACCCTT-
Entoplocamiaaristulata9353	AGGTGGGTGA	CACAT-AGTG	TTCTCGCTGC	T--GCGACCT G-GACCCTT-
Enneapogoncoparius18	AGGTGGGTGA	CACAA-GGTG	TTCTCGCCGC	A--GTGGCTC G-GATCACAT
Enneapogoncenchroides7506	AGGTGGGTGA	CACAC-GGTG	TTCTCGCCGC	A--GTGGCTC G-GATCACAT
Enneapogoncenchroides5213	AGGTGGGTGA	CACAC-GGTG	TTCTCGCCGC	A--GTGGCTC G-GATCACAT
Enneapogoncoparius7487	AGGTGGGTGA	CACAA-GGTG	TTCTCGCCGC	A--GTGGCTC G-GATCACAT
Schmidtiaappophoroides6334	AGGTGGGTGA	CACGC-GGTG	TTCTCGCTGC	T--CTGACTT G-GATCACTG
Schmidtiaalihariensis7490	AGGTGGGTGA	CACAA-GGTG	TTCTCGCTGC	C--GTGACTC G-GATCCATG
Schmidtiaalihariensis7486	AGGTGGGTGA	CACAA-GGTG	TTCTCGCTGC	C--GTGACTC G-GATCCATG
Sporobolusafricanus7476	TGGTGGACAA	CATGC-GTTG	TTCTCGGAGT	NNNNNNNNN NNNNNNNNN
Sporobolusconsimilis9354	TGGTGGACAA	CGTCG-GTTG	TTCTCGGAGT	T--GTGACTT G-TACTGGTT
Sporobolusfimbriatus9328	TGGTGGATGA	CAAT-GTTG	TTCTCGGAGT	T--GTGACTT G-TATTGCTT

IMPERATACYLINDRICA  
 SORGHUMHALEPENSE  
 PANICUMLANIPES  
 PANICUMSTAPFIANUM  
 KARROOCHLOAPURPUREA6241  
 CHAETOBROMUSINVOLUCRATES6284  
 Merxmuelเลอร์arangei  
 Centropodiaglauca  
 Cynodondactylon5248  
 Cynodondactylon7480  
 Cynodondactylon  
 Eleusinecoracana  
 Eleusineindica  
 Chlorisvirgata6616  
 Chlorisvirgata7498  
 Eustachyspaspaloides5597  
 Harpochloafalx6530  
 Harpochloafalx5118  
 Harpochloafalx6670  
 Rendliaaltera5133  
 Leptochloafusca7622  
 Enteropogonmacrostachyus9339  
 Astreblalappacea  
 Muhlenbergiarichardsonis  
 Triodiastenostachya  
 Microchloacaffra5146  
 Dactylocteniumaegyptium9326  
 Tragusberteronianus6620  
 Tragusracemosus7108  
 Traguskoelerioides6729  
 Polevansiarigida7621  
 Perotispates9323  
 Eragrostiscurvula6529  
 Eragrostisobtusa6623  
 Eragrostisobtusa6732  
 Eragrostisechinochloidea7485  
 Eragrostispseudobtusa7508  
 Eragrostisechinochloidea5933  
 Eragrostislehmanniana20  
 Eragrostiscurvula5191  
 Eragrostischloromelas6947  
 Eragrostischloromelas7474  
 Eragrostisracemosa6533  
 Eragrostisbiflora6705  
 Eragrostissuperba7470  
 Eragrostiscapensis6551  
 Eragrostiscapensis6532  
 Pogonarthriasquarrosa7483  
 Pogonarthriasquarrosa7504  
 Catalepisgracilis6962  
 Catalepisgracilis6653  
 Cladoraphiscyperoides5704  
 Cladoraphisspinosa6335  
 Cladoraphisspinosa4885  
 Cladoraphiscyperoides4894  
 Fingerhuthiaafricana4349  
 Fingerhuthiaafricana7507  
 Fingerhuthiaafricana4879  
 Entoplocamiaaristulata9353  
 Enneapogonscoparius18  
 Enneapogoncenchroides7506  
 Enneapogoncenchroides5213  
 Enneapogonscoparius7487  
 Schmidtiaappophoroides6334  
 Schmidtiakalihariensis7490  
 Schmidtiakalihariensis7486  
 Sporobolusafricanus7476  
 Sporobolusconsimilis9354  
 Sporobolusfimbriatus9328

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 NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN  
 --CAGCTAGC TCCTTGGCCT AA--AGGACC CATTACGAC CGTAGC-CCT  
 --CAGCTAGC TCCATAGCCT CA--AGGACC CATTACGAC CGTAGCGCCT  
 C-CAG---CG ATAC-GGCC TTT-ACGACC CTT--GAGAC CGCAGCGCTC  
 C-CGG---CA TCAT-GGCC TT--CCGACC CTT--ACACC GCTAGCGCTC  
 C-CGG---CG ACAC-GGCC TT--GGGACC CT----CCAT CGGAGCGCGA  
 C-CGG---CG CGAG-GGCC TC--GGACC CAT--GCGAC CGTAGCGCGC  
 C-CGGTCATG TTAT-GCCCC TTTT-GGACC CAT--GGTT TGGAGCGTAC  
 C-CGGTCATG TTAT-GCCCC TTTT-GGACC CAT--GGTT TGGAGCGTAT  
 C-CGGTCATG TTAT-GCCCC TTTT-GGACC CAT--GGTT TGGAGCGTAT  
 C--GGTGATG CAAT-GGC-- TCTATGGACC CAT--GGAT TAAAGTGCAT  
 C--GGTGATG CAAT-GGC-- TCTATGGACC CAT--GGAT TGAAGTGCAT  
 C-TTGCTATG TTAT-GGCC TTAATGACC CAT--GGTT TTGAGTGCAT  
 C-TTGCTATG TTAT-GGCC TTAATGACC CAT--GGTT TTGAGTGCAT  
 T-CGGTCATG TCAA-GCCCC TTT--AGACC CAT--GAAT TTGAGCGCAT  
 A--GTCATG TTAT-GCCCT TT--GGACC CAT--GGTT TTGAGTGCAT  
 A--GTCATG TTAT-GCCCT TT--GGACC CAT--GGTT TTGAGTGCAT  
 A--GTCATG TTAT-GCCCT TT--GGACC CAT--GGTT TTGAGTGCAT  
 C-CGGTCACC TGAT-GCCCC CTTT-GGACC CAA--GGTT TGGTGCACAT  
 C-TGGTGATG TTAT-GCCCC TTT--GGACC CAT--CGAG TGGAGCACGT  
 A-CGGTGATG TTTGCGGCC TTT--GGACC CWT--GGTT AGGAGCGCAC  
 C-CGGTAATG TTAT-GGCC TTT--GGACC CAT--GGTT TCGAGTGCAT  
 A-CGGCGATA CAAT-GGCC TTT--GGACC CAT--TAAT CGAAGCGCAC  
 C-CGGCGTTG ATAC-GGCC TTT--GGACC CAT--GGAC CGAAGCACAA  
 C-CGGTCATG TGAT-GCCCC TTT--GGACC CAT--GGTT TGGAGCGCTT  
 --CGTGTAGC CCTC-GGG-- ----ACCCC CATTT-GACC GAAGTGCAT  
 T-TGGCTGTG TTCT-GGCC TCT--GGACC CATT--AGAC TGAAGCACAC  
 NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN  
 T-CGGCTGTG TTAT-GGCC TCT--GGACC CATT--AGAC TGAAGCACAC  
 T-CGGCTAAG TTAG-GGCC TCT--GGACC CATT--GAC TGAAGCACAC  
 TCTGGTGTTA CAAC--ATCC CAT--GGACT CAT--ACAT CGAGATGCAA  
 CTATGGCATG AAAT-GGCC TCAT--GACC CAT--CAAC CGATGCGCAT  
 C-----CGTG CAAT-GGCTC TCAT--GACC CAT--CAAC CGAAGCGCAT  
 C-----CGTG CAAT-GGCTC TCAT--GACC CAT--CAAC CGAAGCGCAT  
 C-----CGTG CAAT-GGCTC TCAT--GACC CAT--CAAC CGAAGCGCAT  
 C-----CGTG CAAT-GGCTC TCAT--GACC CAT--CAAC CGAAGCGCAT  
 CTATGGCATG AAAT-GGCC TCAC--GACC CAT--CAAC CGATGCGCAT  
 CTATGGCATG AAAT-GGCC TCAC--GACC CAT--CAAC CGATGCGCAT  
 CTATGGCATG AAAT-GGCC TCAT--GACC CAT--CAAC CGAAGCGCAT  
 CA--GGCATA CAAT-GGCC TCAC--GACC CAT--CAAC CGAAGCGCAT  
 CA--TGATG CAAT-GGNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN  
 CA--GGTATG CAAT-GGCC TCAC--GACC CAT--CAAT CGAAGCGTAC  
 C--TGGTACA CAAT-GGCC TCAT--GACC CAT--CAAC CGAAGCGCAC  
 CC---TATG CAAT-AGTCC TCAG--GACC CAT--CAAC CGAAGCGCAT  
 CC--GGCG-A CATT-GGCAC TTT--GGAC CCAT--GAAT TG-AGCAAAT  
 CC-----ATG CAAG-GGCC TTGT--GACC CAT--CAAC C-AAGCGCAT  
 CC-----ATG CAAG-GGCC TTGT--GACC CAT--CAAC C-AAGCGCAT  
 CA--GGCATG CAAT-GGCC TCAT--GACC CAT--GAAC CAAAGAGCAT  
 CA--GGCATG CAAT-GGCC TCAT--GACC CAT--GAAC CAAAGAGCAT  
 CC---TATG CAAT-GGCC TCAT--GACC CAT--CAAC CGATGCGCAC  
 CC---TATG CAAT-GGCC TCAT--GACC CAT--CAAC CGATGCGCAC  
 CC---TATG CAAT-GGCC TCAT--GACC CAT--CAAC CGATGCGCAC  
 CC---TATG CAAT-GGCC TCAT--GACC CAT--CAAC CGATGCGCAC  
 CC---AAGG CAAT-TACCC TCAT--GACC CAT--CGAC CGAAGCGCAC  
 CC---AAGG CAAT-TACCC TCAT--GACC CAT--CGAC CGAAGCGCAC  
 CC---AAGG CAAT-TACCC TCAT--GACC CAT--CGAC CGAAGCGCAC  
 GC---AAGG CAAC-GACCC TCAT--GACC CAT--CGAC CGAAGCGCAC  
 CCA-GGCAAG CA-T-GGCC TCAA--GACC CAT--CGAC CGTAGCGCAC  
 CCA-GGCAAG CA-T-GGCC TCAA--GACC CAT--CGAC CGTAGCGCAC  
 CCA-GGCAAG CA-T-GGCC TCAA--GACC CAT--CGAC CGTAGCGCAC  
 CCT-GGCAAG CAAT-GGCC TCAC--GACC CAT--CAAC CGGAGCGCAC  
 ACT-GGCAAG CAAT-GGCC TCAT--GACC CTT--CGAC CGTAGCGCAC  
 ACT-GGCAAG CAAT-GGCC TCAT--GACC CTT--CGAC CGTAGCGCAC  
 NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN  
 T-CCGCG--A CACT-GGCC T-TA-GGACC CAT--GAAT CT-AGCATCG  
 CTAGCGAT-A GTAT-GGCTG T-TA-TGACC CAT--GCAT CGGAGCTTTG

	651	668
IMPERATACYLINDRICA	NNNNNNNNNN	NNNNNNNN
SORGHUMHALEPENSE	NNNNNNNNNN	NNNNNNNN
PANICUMLANIPES	TGGTG-----	-CTCGGAC
PANICUMSTAFFIANUM	TGGTG-----	-CTCGGAC
KARROOCHLOAPURPUREA6241	-GACG-----	-CTCGGAC
CHAETOBROMUSINVOLUCRATES6284	-GTCACTCGG	ATCGCGAC
Merxmullerangei	-GACG-----	-CTCGGAC
Centropodiaglauca	-GCCG-----	-CTCGGAC
Cynodondactylon5248	-GTAG-----	-CTCGGAC
Cynodondactylon7480	-GTAG-----	-CTCGGAC
Cynodondactylon	-GTAG-----	-CTCGGAC
Eleusinecoracana	-GTTT-----	-ATCAAAC
Eleusineindica	-GTTT-----	-ATCAAAC
Chlorisvirgata6616	-GCCG-----	-CTCGGTC
Chlorisvirgata7498	-GCCG-----	-CTCGGTC
Eustachyspaspaloides5597	-GTTG-----	-CTTGAC
Harpochoafalx6530	-GCTT-----	-CTTGAC
Harpochoafalx5118	-GCTT-----	-CTTGAC
Harpochoafalx6670	-GCTT-----	-CTTGAC
Rendliaaltera5133	-GTTG-----	-CTCGGAC
Leptochloafusca7622	-GTTG-----	-CTCGGAC
Enteropogonmacrostachyus9339	-GTTG-----	-CTCGTAC
Astreblalappacea	-GTCG-----	-CTCGGAC
Muhlenbergiarichardsonis	-GTCG-----	-CTCGAAC
Triodiastenostachya	-GTTG-----	-CTCGGAC
Microchloacaffra5146	-GTCG-----	-CTTGAC
Dactylocteniumaegyptium9326	-GTTA-----	-CTCGGAC
Tragusberteronianus6620	-GTTG-----	-CTCGGAC
Tragusracemosus7108	NNNNNNNNNN	NNNNNNNN
Traguskoeleroides6729	-GTTG-----	-CTCGGAC
Polevansiarigida7621	-GTTG-----	-CTCGGAC
Perotispatens9323	-GTCA-----	-CTTGAC
Eragrostiscurvula6529	-GTCG-----	-CTCGGAC
Eragrostisobtusa6623	-G-----	-CTCGGAC
Eragrostisobtusa6732	-G-----	-CTCGGAC
Eragrostisechinocloidea7485	-G-----	-CTCGGAC
Eragrostispseudobtusa7508	-G-----	-CTCGGAC
Eragrostisechinocloidea5933	-G-----	-CTCGGAC
Eragrostislehmanniana20	-GTCG-----	-CTCGGAC
Eragrostiscurvula5191	-GTCG-----	-CTCGGAC
Eragrostischloromelas6947	-GTCG-----	-CTCGGAC
Eragrostischloromelas7474	-GTCG-----	-CTCGGAC
Eragrostisracemosa6533	NNNNNNNNNN	NNNNNNNN
Eragrostisbiflora6705	-GTCG-----	-CTCGGAC
Eragrostissuperba7470	-GTCG-----	-CTCGGAC
Eragrostiscapensis6551	-GTCG-----	-CTCGGAC
Eragrostiscapensis6532	-GTTG-----	-CTCGAAC
Pogonarthriasquarrosa7483	-G-----	-CTCGGAC
Pogonarthriasquarrosa7504	-G-----	-CTCGGAC
Catalepisgracilis6962	-GTTG-----	-CTTGAC
Catalepisgracilis6653	-GTTG-----	-CTTGAC
Cladoraphiscyperoides5704	-GTCG-----	-CTCGGAC
Cladoraphisspinosa6335	-GTCG-----	-CTCGGAC
Cladoraphisspinosa4885	-GTCG-----	-CTCGGAC
Cladoraphiscyperoides4894	-GTCG-----	-CTCGGAC
Fingerhuthiaafricana4349	-GTCG-----	-CTCGGAC
Fingerhuthiaafricana7507	-GTCG-----	-CTCGGAC
Fingerhuthiaafricana4879	-GTCG-----	-CTCGGAC
Entoplocamiaaristulata9353	-GTCG-----	-CTCGGAC
Enneapogonscoparius18	-GTCG-----	-CTCGGAC
Enneapogoncnchroides7506	-GTCG-----	-CTCGGAC
Enneapogoncnchroides5213	-GTCG-----	-CTCGGAC
Enneapogonscoparius7487	-GTCG-----	-CTCGGAC
Schmidtiaapappohoroides6334	-GTCG-----	-CTCGGAT
Schmidtiaakalihariensis7490	-GCCG-----	-CTCGGAC
Schmidtiaakalihariensis7486	-GCCG-----	-CTCGGAC
Sporobolusafricanus7476	NNNNNNNNNN	NNNNNNNN
Sporobolusconsimilis9354	-ATTG-----	-CTCGCAC
Sporobolusfimbriatus9328	-ATTG-----	-CTCAAC

**APPENDIX E.** Indels coded by GapCoder for *ITS* data set. Where missing data was coded in the sequence data set (N), the corresponding indel was coded as ?. The indel characters are coded with 1 for present, 0 for missing and – for inapplicable. As the gap matrix was added directly to the sequencing matrix the numbering continues from the last character in the sequence data set. Due to the large size of the matrix and to keep sequences together, this appendix starts on the next page.

IMPERATACYLINDRICA	??10001---00001000001000100-011000111100110000010-10110-0010
SORGHUMHALEPENSE	1010001---00001000001000100-011000111100110000010-10110-0010
PANICUMLANIPES	0010001---000010000000010-1--011000111100110000010-10110-0010
PANICUMSTAPFIANUM	0010001---00001000000010-1--011000111100110000010-10110-0010
KARROOCHLOAPURPUREA6241	?????????????010001--000100-011000011100110000010-10110-0010
CHAETOBROMUSINVOLUCRATES6284	??0010
Merxmuellierangei	01100000010000100001000100-011000011100110000010-10110-0010
Centropodiaglauc	011000001010000101---000100-011000011100110000010-10110-0010
Cynodondactylon5248	01100001010000101---000100-011000011100110000010-10110-0011
Cynodondactylon7480	01100001010000101---000100-011000011100110000010-10110-0011
Cynodondactylon	01100000010000101---000100-011000011100110000010-10110-0010
Eleusinecoracana	0010000001000010001--000100-011000011100110001010-10110-0010
Eleusineindica	0010000001000010001--000100-011000011100110001010-10110-0010
Chlorisvirgata6616	0110000001000010001--000100-011000011100110000010-10110-0010
Chlorisvirgata7498	0110000001000010001--000100-011000011100110000010-10110-0010
Eustachyspaspaloides5597	0000000001000010001--000100-1-0000011100110000010-10110-0010
Harpochloafalx6530	0110000101000010001--000100-011000011100110000010-10110-0010
Harpochloafalx5118	?????????????????????????????????00100-011000011100110000010-10110-0010
Harpochloafalx6670	0110000101000010001--000100-011000011100110000010-10110-0010
Rendliaaltera5133	0110000001000010001--000100-011000011100110000010-10110-0010
Leptochloafusa7622	0110000001000010001--000100-011000011100110000010-10110-0010
Enteropogonmacrostachyus9339	0110110001000010001--000100-0110000111001100000000-1010-0010
Astreblalappacea	0110000001000010001--000100-011000011100110000010--1110-0010
Muhlenbergiarichardsonis	011000000100000000001000100-010000011110110000000110110-0010
Triodiastenosstachya	0110000001000010001--001-00-001000011100110000010-10110-0010
Microchloacaffra5146	01100000010000101---000-01-011000011100110000010-10110-0010
Dactylocteniumaegyptium9326	0110000001001010001--00000000110000101000100001-1-101??0000
Tragusberteronianus6620	01100000010000101---001-00-011000011100110000010-101-1-0010
Tragusracemosus7108	0110000001000010001--001-00-011000011100110000010-101-1-0010
Traguskoelerioides6729	0110000001000010001--001-00-011000011100110000010-101-1-0010
Polevansiarigida7621	011000000100011000001001-00-011000011100110000010-101-1-0010
Perotispatens9323	0110000011000010010-00000001001100011100110000010-1010000010
Eragrostiscurvula6529	??????????1100-100010000100-011000011100100000010-10010-0010
Eragrostisobtusa6623	?????????????????010-0000100-011001011100100010010-10010-0010
Eragrostisobtusa6732	01100000001000-100010000100-011001011100100010010-10010-0010
Eragrostiszechinochloidea7485	?????????????????00010000100-011001011100100010010-10010-0010
Eragrostisepseudobtusa7508	01100000001000-100010000100-011001011100100010010-10010-0010
Eragrostisechinochloidea5933	01100000011000-100010000100-011001011100100010010-10010-0010
Eragrostislehmanniana20	01100000011000-100010000100-011001011100100000010-00010-0010
Eragrostiscurvula5191	??????????1100-100010000100-011001011100100000010-10010-0010
Eragrostischloromelas6947	01100000011000-100010000100-011001011100100000010-10010-0010
Eragrostischloromelas7474	01100000011000-100010000100-0110010111001010000010-10010-0010
Eragrostisracemosa6533	?????????????001-100010000100-011001011100101000010-10010-1110
Eragrostisbiflora6705	01100000011000-100010000100-0110010111001010000010-10010-0010
Eragrostis superba7470	01100000011000-100010000100-011001011100100100010-10010-0010
Eragrostiscapensis6551	01100000011000-1010-0000100-011001011100100000010-10010-0010
Eragrostiscapensis6532	?????????????????010-0000100-011001011100100100010-10010-0010
Pogonarthriasquarrosa7483	?????????????????????????????????????011001011100100100010-10010-0010
Pogonarthriasquarrosa7504	01100000011000-100010000100-011001011100100100010-10010-0010
Catalepisgracilis6962	01100000011000-100010000100-011001011100101000010-10010-0010
Catalepisgracilis6653	?????????????000-100010000100-011001011100101000010-10010-0010
Cladoraphiscyperoides5704	?????????????1000-100010000100-011001011100100100010-10010-0010
Cladoraphis spinosa6335	?????????????1000-100010000100-011001011100100100010-10010-0010
Cladoraphis spinosa4885	01110000011000-100010000100-011001011100100100010-10010-0010
Cladoraphiscyperoides4894	?????????????????00010000100-011001011100100100010-10010-0010
Fingerhuthiaafricana4349	01100000011000-100010000100-011000011100101000010-10110-0010
Fingerhuthiaafricana7507	01100000011000-100010000100-011000011100101000010-10110-0010
Fingerhuthiaafricana4879	01100000011000-100010000100-011000011100101000010-10110-0010
Entoplocamia aristulata9353	01100000011000-100010000100-011000011100101000010-10110-0010
Enneapogon scoparius18	?????????????000-100010000100-011000011100100100010-10110-0010
Enneapogon cenchroides7506	01100000011000-100010000100-011010011100100100010-1010010010
Enneapogon cenchroides5213	01100000011000-100010000100-011000011100100100010-10110-0010
Enneapogon scoparius7487	01100000011000-100010000100-011000011100100100010-10110-0010
Schmidtia pappophoroides6334	01100000011000-100010000100-010000011100100100010-10110-0010
Schmidtia kalihariensis7490	01100000011000-100010000100-011000011100100100010-10110-0010
Schmidtia kalihariensis7486	?????????????000-100010000100-011000011100100100010-10110-0010
Sporobolus africanus7476	01100000011000-100010000100-01000001101-00100010-10110-0010
Sporobolus consimilis9354	01100000011000-100010001-00-001000011101-00100010-1010000010
Sporobolus fimbriatus9328	01100000011000-1000101000100-001000011001-00100010-10110-0010

IMPERATACYLINDRICA	0011-0-01-0100000011000000001010000100????????????????????
SORGHUMHALEPENSE	0011-0-01-0100000011000000001000000100????????????????????
PANICUMLANIPES	0011-0-01-0100000011000000000010001-01--001---00000000000000
PANICUMSTAFFIANUM	000010-01-0100000011000000000010001-01--001---00000000000000
KARROCHLOAPURPUREA6241	0011-0-01-0-10000011000000001000011-000100010100000001000100
CHAETOBROMUSINVOLUCRATES6284	1--00001-00000001100000001001101-00010000100000001000100
Merxmuelleraangei	1----1-01-00000001100000001000101-000100000100000001000100
Centropodiaglauca	1----1-01-1000000011000000001000101-000100000100000001000100
Cynodondactylon5248	001--1-01-0000010011000001-01000101-000100000100000000000100
Cynodondactylon7480	001--1-01-0000010011000001-01000101-000100000100000000000100
Cynodondactylon	001--1-01-0000010011000001-01000101-000100000100000000000100
Eleusinecoracana	001--1-11-0000000011010001-01000101-000100000-10000000000101
Eleusineindica	001--1-11-0000000011010001-01000101-000100000-10000000000101
Chlorisvirgata6616	1----1-01-0000000011000000101000101-000100000100000000000100
Chlorisvirgata7498	1----1-01-0000000011000000101000101-000100000100000000000100
Eustachyspaspaloides5597	1----1-01-00000100110001-0-01000100-001-00000100000000000100
Harpochloafalx6530	1----1-01-00000010110001-0-010001000100-10000--10-00-0000100
Harpochloafalx5118	1----1-01-00000010110001-0-010001000100-10000--10-00-0000100
Harpochloafalx6670	1----1-01-00000010110001-0-010001000100-10000--10-00-0000100
Rendliaaltera5133	1----1-01-00000100110001-0-010001011000100000100000000000100
Leptochloafusa7622	1----1-01-00000100110001-0-010001011000100000100000000000100
Enteropogonmacrostachyus9339	1----1-01-00000100010001-0-01000101100010000010000000000000
Astrelalappacea	1----1-01-00000100110001-0-010001011000100000100000000000100
Muhlenbergiarichardsonis	1----1-01-00000100110001-0-010001011000100000100000000000100
Triodiastenostachya	001--1-01-0--1-100110001-0-010001011000100000100000000000100
Microchloacaffra5146	1--000101-0000000110001-0-010001011000100000100000000000100
Dactylocteniumaegyptium9326	1----1-01-00000100110001-0-010001011000100001-0000000000010-
Tragusberteronianus6620	1----1-01-00000100110001-0-010001011000100000100000000000100
Tragusracemosus7108	1----1-01-00000100110001-0-01????????????????????????????????
Traguskoelerioides6729	1----1-01-00000100110001-0-01000101-000100000100000000000100
Polevansiarigida7621	1----1-01-00000100110001-0-01100101-000100000100000000000100
Perotispatens9323	1--1-0-01-00001100110001-0-01000101-00000000000000000000-10
Eragrostiscurvula6529	1----1-01-0000010011000000001000101-000100000000000000000100
Eragrostisobtusa6623	1----1-01-0000010011000000001000101-000100000---1--0-0000100
Eragrostisobtusa6732	1----1-01-0000010011000000001000101-000100000---1--0-0000100
Eragrostisechinochloidea7485	1----1-01-0000010011000000001000101-000100000---1--0-0000100
Eragrostispseudobtusa7508	1----1-01-0000010011000000001000101-000100000---1--0-0000100
Eragrostisechinochloidea5933	1----1-01-0000010011000000001000101-000100000---1--0-0000100
Eragrostislehmanniana20	1----1-01-0000010011000000001000101-000100000000000000000100
Eragrostiscurvula5191	1----1-01-0000010011000000001000101-000100000000000000000100
Eragrostischloromelas6947	1----1-01-0000010011000000001000101-000100000000000000000100
Eragrostischloromelas7474	1----1-01-0000010011000000001000101-00010100000000100-0000100
Eragrostisracemosa6533	1----1-01-0000010111000000001000101-00010000000000100-000010?
Eragrostisbiflora6705	1----1-01-0000010011000000001000101-00010100000000100-0000100
Eragrostissuperba7470	1----1-01-0000010011000000001000101-000100000-10000000000100
Eragrostiscapensis6551	1----1-01-0000010011000000001000101-0001000000000-10-0000100
Eragrostiscapensis6532	1----1-01-0000010011000000001000101-0001000000000100-0010100
Pogonarthriasquarrosa7483	1----1-01-0000010011000000001000101-0001000000000--1-0000100
Pogonarthriasquarrosa7504	1----1-01-0000010011000000001000101-0001000000000--1-0000100
Catalepisgracilis6962	1----1-01-0000010011000000001000101-000101000000000100-0000100
Catalepisgracilis6653	1----1-01-0000010011000000001000101-00010100000000100-0000100
Cladoraphiscyperoides5704	1----1-01-0000010011000000001000101-0001010000000-10-0000100
Cladoraphisspinosa6335	1----1-01-0000010011000000001000101-0001010000000-10-0000100
Cladoraphisspinosa4885	1----1-01-0000010011000000001000101-0001010000000-10-0000100
Cladoraphiscyperoides4894	1----1-01-0000010011000000001000101-0001010000000-10-0000100
Fingerhuthiaafricana4349	1----1-01-0000010011000000001000101-0001000100000-10-0000100
Fingerhuthiaafricana7507	1----1-01-0000010011000000001000101-0001000100000-10-0000100
Fingerhuthiaafricana4879	1----1-01-0000010011000000001000101-0001000100000-10-0000100
Entollocamiaaristulata9353	1----1-01-0000010011000000001000101-0001000100000-10-0000100
Enneapogonscoparius18	1----1-01-0000010011000000001000101-000100000000000010001100
Enneapogoncenchroides7506	1----1-01-0000010011000000001000101-000100000000000010001100
Enneapogoncenchroides5213	1----1-01-0000010011000000001000101-000100000000000010001100
Enneapogonscoparius7487	1----1-01-0000010011000000001000101-000100000000000010001100
Schmidtiaappophoroides6334	1----1-01-0000010011000000001000101-000100000000000010000100
Schmidtiaakalihariensis7490	1----1-01-0000010011000000001000101-000100000000000010000100
Schmidtiaakalihariensis7486	1----1-01-0000010011000000001000101-000100000000000010000100
Sporobolusaffricanus7476	1--1-0-001000001001000101000100010????????????????????????
Sporobolusconsimilis9354	1--1-0-0000000010010000010001000101-0001000001000000001-0100
Sporobolusfimbriatus9328	0101-0-000000010010100010001000101-000100000000000000010100

IMPERATACYLINDRICA	????????????????????
SORGHUMHALEPENSE	????????????????????
PANICUMLANIPES	00010-0000000000001001
PANICUMSTAFFIANUM	00010-000000000000001
KARROOCHLOAPURPUREA6241	0100010000001000000101
CHAETOBROMUSINVOLUCRATES6284	00010-0000001000000100
Merxmullerangei	00010-00001---0-000101
Centropodiaglauca	000-1--0-0001000000101
Cynodondactylon5248	000000001001--0-000101
Cynodondactylon7480	000000001001--0-000101
Cynodondactylon	000000001001--0-000101
Eleusinecoracana	0-0000000001--0-000101
Eleusineindica	0-0000000001--0-000101
Chlorisvirgata6616	000000000001--0-000101
Chlorisvirgata7498	000000000001--0-000101
Eustachyspaspaloides5597	00000-10-001--0-000101
Harpochloafalx6530	000-1--0-001--0-000101
Harpochloafalx5118	000-1--0-001--0-000101
Harpochloafalx6670	000-1--0-001--0-000101
Rendliaaltera5133	000000001001--0-000101
Leptochloafusca7622	00000-10-001--0-000101
Enteropogonmacrostachyus9339	00000-10-001--0-000101
Astreblalappacea	00000-10-001--0-000101
Muhlenbergiarichardsonis	00000-10-001--0-000101
Triodiastenostachya	00000-10-001--0-000101
Microchloacaffra5146	00000-10-001--0-000101
Dactylocteniumaegyptium9326	1-----0-0000001000101
Tragusberteronianus6620	00000-10-000010-000101
Tragusracemosus7108	????????????????????
Traguskoelerioides6729	00000-10-000010-000101
Polevansiarigida7621	00000-10-0000-1-000101
Perotispatens9323	00000-10-001--0-000101
Eragrostiscurvula6529	00000000-101--0-000101
Eragrostisobtusa6623	00000000-101--0-00011-
Eragrostisobtusa6732	00000000-101--0-00011-
Eragrostisechinochloidea7485	00000000-101--0-00011-
Eragrostispseudobtusa7508	00000000-101--0-00011-
Eragrostisechinochloidea5933	00000000-101--0-00011-
Eragrostislehmanniana20	00000000-101--0-000101
Eragrostiscurvula5191	00000000-101--0-000101
Eragrostischloromelas6947	00000000-101--0-000101
Eragrostischloromelas7474	00000000-101--0-000101
Eragrostisracemosa6533	????????????????????
Eragrostisbiflora6705	00000000-101--0-000101
Eragrostissuperba7470	00000000-101--0-000101
Eragrostiscapensis6551	00000000-101--0-000101
Eragrostiscapensis6532	00000--1--00010-010101
Pogonarthriasquarrosa7483	00000000-101--0-10011-
Pogonarthriasquarrosa7504	00000000-101--0-10011-
Catalepisgracilis6962	00000000-101--0-000101
Catalepisgracilis6653	00000000-101--0-000101
Cladoraphiscyperoides5704	00000000-101--0-000101
Cladoraphisspinosa6335	00000000-101--0-000101
Cladoraphisspinosa4885	00000000-101--0-000101
Cladoraphiscyperoides4894	00000000-101--0-000101
Fingerhuthiaafricana4349	00000000-101--0-000101
Fingerhuthiaafricana7507	00000000-101--0-000101
Fingerhuthiaafricana4879	00000000-101--0-000101
Entoplocamiaaristulata9353	00000000-101--0-000101
Enneapogonscoparius18	00000000-101--0-000101
Enneapogoncenchroides7506	00000000-101--0-000101
Enneapogoncenchroides5213	00000000-101--0-000101
Enneapogonscoparius7487	00000000-101--0-000101
Schmidtiaappophoroides6334	00000000-101--0-000101
Schmidtialihariensis7490	00000000-101--0-000101
Schmidtialihariensis7486	00000000-101--0-000101
Sporobolusafricanus7476	????????????????????
Sporobolusconsimilis9354	001000001001--0-010101
Sporobolusfimbriatus9328	001000001001--0-000101

**APPENDIX F.** Table of correspondence between the indels and the regions they represent (Appendix E) generated by GapCoder for the *ITS* dataset.

Indel Character	Sequence Region	Indel Character	Sequence Region	Indel Character	Sequence Region
669	9-9	717	214-216	765	584-587
670	15-15	718	215-215	766	590-592
671	34-34	719	224-224	767	591-592
672	42-42	720	224-225	768	592-592
673	43-43	721	235-235	769	592-593
674	53-53	722	240-241	770	599-599
675	54-70	723	240-243	771	600-602
676	60-60	724	240-240	772	600-600
677	61-61	725	376-376	773	601-602
678	64-65	726	391-391	774	602-602
679	73-73	727	423-423	775	602-603
680	86-86	728	426-426	776	602-604
681	90-90	729	439-440	777	602-606
682	92-92	730	439-439	778	603-604
683	94-94	731	440-440	779	603-606
684	94-95	732	444-456	780	603-607
685	103-107	733	444-455	781	604-604
686	103-106	734	444-457	782	606-608
687	104-107	735	444-454	783	608-609
688	104-106	736	469-469	784	609-609
689	105-107	737	484-487	785	613-613
690	108-108	738	486-487	786	615-615
691	113-113	739	490-490	787	615-616
692	116-119	740	491-491	788	619-620
693	117-119	741	491-492	789	619-625
694	117-121	742	491-493	790	620-620
695	117-120	743	493-493	791	622-622
696	117-118	744	497-497	792	623-624
697	125-126	745	498-498	793	623-625
698	126-126	746	506-506	794	624-624
699	132-132	747	511-511	795	624-625
700	139-139	748	515-515	796	624-626
701	143-147	749	516-516	797	625-625
702	148-148	750	519-519	798	625-626
703	150-150	751	525-525	799	633-636
704	153-153	752	528-531	800	634-635
705	162-162	753	528-528	801	634-636
706	170-173	754	529-532	802	635-636
707	180-180	755	529-531	803	635-637
708	181-183	756	539-539	804	636-636
709	182-183	757	544-544	805	642-642
710	188-188	758	556-556	806	643-643
711	193-193	759	563-563	807	647-647
712	194-194	760	565-566	808	651-651
713	196-196	761	566-566	809	653-661
714	197-197	762	574-574	810	656-661
715	198-198	763	582-583		
716	214-215	764	582-582		



**APPENDIX G.** Aligned sequence data of chloridoid and outgroup taxa for *trnL*-F. Numbers indicate the consecutive positions of 1 to 1191 (5' to 3') from the beginning of the *trnL* intron region to the end of the *trnL*-F intergenic spacer region. Dashes denote gaps. Arrows indicate the beginning of the *trnL* intron, *trnL* exon and *trnL*-F intergenic spacer regions. N indicates large regions of missing data where sequencing reactions failed.

	↓	1				50
PHRAGMITESAUSTRALIS6557		NNNNTGTAT-	GAGCCT-GGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
ARISTIDAADSCENSIONIS6540		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
KARROOCHLOAPURPUREA6241		NNT-TGTATT	GAGCCT-GGT	A-GGAAACCT	GCTAAGTGGT	AACCTCCAAA
CHAETOBROMUSINVOLUCRATES6284		AAT-TGTATT	GAGCCTTGGT	ATGGAA-CCT	GCTAAGTGGT	AACCTCCAAA
SORGHUMHALEPENSE		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNTCCAAA
IMPERATACYLINDRICA		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNTCCAAA
PANICUMLANIPES		GATATGTATT	GAGCCTTGGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
PANICUMSTAFFIANUM		GATATGTATT	GAGCCTTGGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Eleusinecoracana		GAT-TGTATT	GAGCCTTGGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Eleusineindica		GAT-TGTATT	GAGCCTTGGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Cynodondactylon		GAT-TGTATT	GAGCCTTGGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Cynodondactylon7480		NNNNNNNNNN	NNNNT-GGT	ATGGAAACCT	GCTACGTGGT	AACCTCCAAA
Chlorisvirgata6616		GAT-TGTAT-	GAGCCT-GGT	ATGGAA-CCT	GCTAAGTGGT	AACCTCCAAA
Chlorisvirgata7498		NNNNNNNNNN	NNNNNNNNNN	NNGAAA-CCT	GCTAAGTGGT	AACCTCCAAA
Eustachyaspaloides5597		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Harpochloafalx6530		GAT-TGTATT	GAGCCTTGGT	ATGGA--CCT	GCTAAGTGGT	AACCTCCAAA
Harpochloafalx6670		GAT-TGTATT	GAGCCTTGGT	ATGGA-CCT	GCTAAGTGGT	AACCTCCAAA
Harpochloafalx5118		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Rendliaaltera5133		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNCCT	GCTAAGTGGT	AACCTCCAAA
Leptochloafusca7622		NNNNNNNNNN	NNNNNNNGGT	ATGGAA-CCT	GCTAAGTGGT	AACCTCCAAA
Enteropogonmacrostachyus9339		NNNNNNNNNN	NNNNNNNNNN	ATGGAA-CCT	GCTAAGTGGT	AACCTCCAAA
Dactylocteniumaegyptium9326		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Tragusberteronianus6620		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNCAAA
Tragusracemosus7108		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Traguskoelerioides6729		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Polevansiarigida7621		NNNNNNNNNN	NNNNNNNNNN	NNGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Perotispatens9323		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostiscurvula6529		GAT-TGTATT	GAGCCTTGGT	ATGGAA--CT	GCTAAGTGGT	AACCTCCAAA
Eragrostisobtusa6623		GAT-TGTATT	GAGC-TTGGT	ATGGAAACCT	GGTAAGTGGT	AACCTCCAAA
Eragrostisobtusa6732		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostisechinochloidea7485		GAT-TGTATT	GAGC-TTGGT	ATGGAAA-CT	GCTAAGTGGT	AACCTCCAAA
Eragrostispseudobtusa7508		GAT-TGTATT	GAGCC-TGGT	ATGGAA--CT	GCTAAGTGGT	AACCTCCAAA
Eragrostislehmanniana20		NNNNGTATT	GAGCCTG-T	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Eragrostischloromelas6947		GAT-TGTATT	GAGCCTTGGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Eragrostischloromelas7474		GAT-TGTATT	GAG-CTTGGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Eragrostisracemosa6533		GAT-TGTATT	GAGCCTCGGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Eragrostisbiflora6705		NNNNNNNNNN	NNNNNNNNNN	NNGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Eragrostisuperba7470		GAT-TGTATT	GAGCCTTGGT	ATGGAA--CT	GCTAAGTGGT	AACCTCCAAA
Eragrostiscapensis6551		GAT-TGTATT	GAG-CT-GGT	ATGGAA-CCT	GCTAAGTGGT	AACCTCCAAA
Eragrostiscapensis6532		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostislehmanniana7514		GAT-TGTATT	GAGCCTTGGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Pogonarthriasquarrosa7483		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNCCT	GCTAGGTGGT	AACCTCCAAA
Pogonarthriasquarrosa7504		GAT-TGTATT	GAGCCTTGGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Catalepisgracilis6962		GAT-TGTATT	GAGCCTTGGT	ATGGAAACCT	GCTAAGTGGT	A-CTTCCAAA
Catalepisgracilis6653		NNNNNNNNNN	NNNNNNNGGT	ATGGAAACCT	GCTAAGTGGT	A-CTTCCAAA
Cladoraphiscyperoides5704		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cladoraphisspinosa6335		NNNNNNNNNN	NNNNNNNGGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Cladoraphiscyperoides4894		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Fingerhuthiaafricana7507		GAT-TGTATT	GAG-CT-GGT	ATGGAAA-CT	GCTAAGTGGT	AACCTCCAAA
Entolopocamiaaristulata9353		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Enneapogonscoparius18		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNCCT	GCTAAGTGGT	AACCTCCAAA
Enneapogonenchroides7506		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	AACCTCCAAA
Enneapogonscoparius7486		GAT-TGTATT	GAGCCTTGGT	ATGGAAA-CT	GCTAAGTGGT	AACCTCCAAA
Schmidtiaappophoroides6334		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNCTCCAAA
Schmidtiaalihariensis7490		NNNNTGTATT	GAGC--TGTT	ATGGAA--CT	GCTAAGTGGT	AACCTCCAAA
Schmidtiaalihariensis7487		GAT-TGTATT	GAGCCTTGGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Sporobolusafricanus7476		GAT--GTAT-	GAGCCT-GGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Sporobolusconsimilis9354		GAT-TGTATT	GAG-CT-GGT	ATGGGAAACCT	GTTAAGTGGT	AACCTCCAAA
Sporobolusfimbriatus7509		GAT-TGTAT-	GAGCCTTGGT	ATGGAAACCT	GCTAAGTGGT	AACCTCCAAA
Sporoboluspyramidalis6966		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Sporobolusfimbriatus9328		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN

PHRAGMITESAUSTRALIS6557	TTCAGAGAAC	CCCTGGAAT-	AAAAATGGGC	AATCCTGAGC	CAAATCC-TT
ARISTIDAADSCENSIONIS6540	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
KARROOCHLOAPURPUREA6241	TTCAGAGAAA	CCCTGGAATT	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
CHAETOBROMUSINVOLUCRATES6284	TTCAGAGAAA	CCCTGGAATT	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
SORGHUMHALEPENSE	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCACT
IMPERATACYLINDRICA	TTCAGAGAAA	CC-TGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCACT
PANICUMLANIPES	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
PANICUMSTAFFIANUM	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Eleusinecoracana	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Eleusineindica	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Cynodondactylon	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Cynodondactylon7480	TTCAGAGAAA-	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Chlorisvirgata6616	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Chlorisvirgata7498	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Eustachyaspaloides5597	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Harpochloafalx6530	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Harpochloafalx6670	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Harpochloafalx5118	NNNNNNNNNN	NNNNNNNNNN	NNNNNNGGGC	AATCCTGAGC	CAAGTCCCTT
Rendliaaltera5133	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTG
Leptochloafusca7622	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Enteropogonmacrostachyus9339	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Dactylocteniumaegyptium9326	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Tragusberteronianus6620	TTCAGACAAT	CCCTGCAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Tragusracemosus7108	NNNNNNNNNN	NNNNNNNAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Traguskoelerioides6729	NNNNNNNNNN	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Polevansiarigida7621	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Perotispates9323	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostiscurvula6529	TTCAGAGAA-	CCCTGGAAGG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Eragrostisobtus6623	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Eragrostisobtus6732	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostisechinochloidea7485	TTCAGAGAAA	CC-TGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Eragrostispseudobtus6708	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Eragrostislehmanniana20	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Eragrostischloromelas6947	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Eragrostischloromelas7474	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Eragrostisiracemosa6533	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Eragrostisbiflora6705	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Eragrostissuperba7470	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Eragrostiscapensis6551	TTCAGAGA-A	CCCTGGAATG	AAAAATGGGC	AATC-TGAGC	CAAATCCCTT
Eragrostiscapensis6532	NNNNNNNNNN	NNNNNNNNNN	NNNNNTGGGC	A-TCCCTGAGC	CAAATCCCTT
Eragrostislehmanniana7514	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Pogonarthriasquarrosa7483	T-CAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Pogonarthriasquarrosa7504	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Catalepisgracilis6962	TTCAGAGAA-	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTG
Catalepisgracilis6653	TTCAGAGAA-	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTG
Cladoraphiscyperoides5704	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cladoraphisspinosa6335	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Cladoraphiscyperoides4894	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Fingerhuthiaafricana7507	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATC-TGAGC	CAAATCCCTT
Entollocamiaristulata9353	NNNNNNNNNN	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Enneapogonscoparius18	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Enneapogoncenchroides7506	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Enneapogonscoparius7486	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Schmidtiaappophoroides6334	TTCAGACAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Schmidtiaalihariensis7490	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Schmidtiaalihariensis7487	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Sporobolusaffricanus7476	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Sporobolusconsimilis9354	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Sporobolusfimbriatus7509	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Sporoboluspyramidalis6966	TTCAGAGAAA	CCCTGGAATG	AAAAATGGGC	AATCCTGAGC	CAAATCCCTT
Sporobolusfimbriatus9328	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN

PHRAGMITESAUSTRALIS6557	TTTT----G	AAAAA---C	AGGTGGTTCT	CAAAC TAGAA	CCCAAAGGGA
ARISTIDAADSCENSIONIS6540	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	CAAAC TAGAA	CCCAAAGGAA
KARROOCHLOAPURPUREA6241	TTTT----G	AAAAA---C	ACGTGGTCCT	CAAAC TAGAA	CCCAAAGGAA
CHAETOBROMUSINVOLUCRATES6284	TTTT----G	AAAAA---C	AAGCGGTCCT	CAAAC TAGAA	CCCAAAGGAA
SORGHUMHALEPENSE	TTTTTT---C	AAAAA---C	--GTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
IMPERATACYLINDRICA	TTTTTT---G	AAAAA---C	AAGTG-----	CAAAC TAGAA	CCCAAAGGAA
PANICUMLANIPES	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
PANICUMSTAPFIANUM	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Eleusinecoracana	TTTTTCATT	TAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Eleusineindica	TTTTTCATT	TAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Cynodondactylon	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Cynodondactylon7480	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Chlorisvirgata6616	TTTT----T	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Chlorisvirgata7498	TTTT----T	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Eustachyspaspaloides5597	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Harpochloafalx6530	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Harpochloafalx6670	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Harpochloafalx5118	TTTT----G	AAAA-----	--TGGTTCT	CAAA-TATAA	CCC---GGAA
Rendliaaltera5133	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Leptochloafusca7622	TTT-----G	AAAAA---C	AGGTGGTTTT	CAAAC TAGAA	CCCAAAGGAA
Enteropogonmacrostachyus9339	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Dactylocteniumaegyptium9326	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Tragusberteronianus6620	TTTTG---G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Tragusracemosus7108	TTTTG---G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Traguskoelerioides7629	TTTTG---G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Polevansiarigida7621	TTTTTT---G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Perotispatens9323	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostiscurvula6529	TTTT----G	AAAAA---C	AAGKGGTTCT	CAAAC TAGAA	CCCAAATGAA
Eragrostisobtusata6623	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Eragrostisobtusata6732	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostisechinochloidea7485	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Eragrostispseudobtusata7508	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Eragrostislehmanniana20	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Eragrostischloromelas6947	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Eragrostischloromelas7474	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Eragrostisracemosa6533	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Eragrostisbiflora6705	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Eragrostis superba7470	TTTT----G	AAAAA---C	AAGGGGTTCT	CAAAC TAGAA	CCCAAATGAA
Eragrostiscapensis6551	TTA-----G	AAAAA---C	AAGTGGTTCT	CAA-CTAGAA	CCCAAATGAA
Eragrostiscapensis6532	TTA-----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Eragrostislehmanniana7514	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Pogonarthriasquarrosa7483	TTA-----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Pogonarthriasquarrosa7504	TTA-----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Catalepisgracilis6962	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Catalepisgracilis6653	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Cladoraphiscyperoides5704	NNNNNNNNNN	NNNNNNNNNN	NNNTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Cladoraphis spinosa6335	TTTT----G	AAAAAAAAC	AAGTGGTTCT	CAAAC TAGAA	TCCAAATGAA
Cladoraphiscyperoides4894	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NAAACTAGAA	CCCAAATGAA
Fingerhuthiaafricana7507	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Entollocamiaristulata9353	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Enneapogon scoparius18	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	TCCAAATGAA
Enneapogon cenchroides7506	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	TCCAAATGAA
Enneapogon scoparius7486	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	TCCAAATGAA
Schmidtia pappophoroides6334	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	TCCAAATGAA
Schmidtia kalihariensis7490	TTTT----G	AAAAA---C	AAACGGTTCT	CAAAC TAGAA	CCCAAATGAA
Schmidtia kalihariensis7487	TTTT----G	AAAAAAA---C	AAGCGGTTCT	CAAAC TAGAA	CCCAAATGAA
Sporobolusafricanus7476	TTTT----T	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Sporobolus consimilis9354	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA
Sporobolus fimbriatus7509	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Sporobolus pyramidalis6966	TTTT----G	AAAAA---C	AAGTGGTTCT	CAAAC TAGAA	CCCAAATGAA
Sporobolus fimbriatus9328	NNNNNNNNNN	NNNNNNNNNN	NNGTGGTTCT	CAAAC TAGAA	CCCAAAGGAA

PHRAGMITESAUSTRALIS6557	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
ARISTIDAADSCENSIONIS6540	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
KARROOCHLOAPURPUREA6241	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
CHAETOBROMUSINVOLUCRATES6284	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
SORGHUMHALEPENSE	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
IMPERATACYLINDRICA	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
PANICUMLANIPES	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
PANICUMSTAFIANUM	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Eleusinecoracana	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Eleusineindica	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Cynodondactylon	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Cynodondactylon7480	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Chlorisvirgata6616	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Chlorisvirgata7498	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Eustachyspaspaloides5597	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Harpochloafalx6530	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Harpochloafalx6670	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Harpochloafalx5118	AAGG-----	ATCTGTGCCG	ACACTCAATG	GAAGCTGTTC	TAACGAATCG
Rendliaaltera5133	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Leptochloafusca7622	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Enteropogonmacrostachyus9339	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Dactylocteniumaegyptium9326	NNNNNNNNNN	NNNNNNNNNN	NNNNNCAATG	GAAGCTGTTC	TAACGAATCG
Tragusberteronianus6620	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Tragusracemosus7108	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Traguskoelerioides6729	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Polevansiarigida7621	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Perotispatens9323	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Eragrostiscurvula6529	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Eragrostisobtusa6623	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Eragrostisobtusa6732	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostisechinochloidea7485	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Eragrostispseudobtusa7508	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Eragrostislehmanniana20	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Eragrostischloromelas6947	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Eragrostischloromelas7474	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Eragrostisracemosa6533	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Eragrostisbiflora6705	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Eragrostisuperba7470	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Eragrostiscapensis6551	AAGG-ATAGG	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Eragrostiscapensis6532	AAGGGATAGG	ATAGGTGCAG	AGGCTCAATG	GAAGCTATTG	TAACGAATCG
Eragrostislehmanniana7514	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Pogonarthriasquarrosa7483	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Pogonarthriasquarrosa7504	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Catalepisgracilis6962	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Catalepisgracilis6653	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Cladoraphiscyperoides5704	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Cladoraphisspinosa6335	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Cladoraphiscyperoides4894	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Fingerhuthiaafricana7507	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Entoplocamiaaristulata9353	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Enneapogonscoparius18	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Enneapogoncenchroides7506	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Enneapogonscoparius7486	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Schmidtiaappophoroides6334	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Schmidtiaalihariensis7490	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Schmidtiaalihariensis7487	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Sporobolusafricanus7476	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Sporobolusconsimilis9354	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTGTTC	TAACGAATCG
Sporobolusfimbriatus7509	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Sporoboluspyramidalis6966	AAGG-----	ATAGGTGCAG	AGACTCAATG	GAAGCTATTG	TAACGAATCG
Sporobolusfimbriatus9328	AAGG-----	ATAGGTGCAG	ACACTCAATG	GAAGCTGTTC	TAACGAATCG

PHRAGMITESAUSTRALIS6557	AGGT--AATT	ACGTTGTGTC	GGTAGTGAAA	C----	TCCC	TCTA-----
ARISTIDAADSCENSIONIS6540	AGGT--AATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCTC	TCTC-----
KARROOCHLOAPURPUREA6241	AGGT--AATT	ACGTTGTGTT	GGTAGTGAAA	C----	GCCC	TCTA-----
CHAETOBROMUSINVOLUCRATES6284	AGGT--AATT	CGCTTGTGCT	GGTAGTGAAA	T----	TCCT	TCTA-----
SORGHUMHALEPENSE	AAGT-----	-----	-----	-----	-----	-----
IMPERATACYLINDRICA	AAGT-----	-----	-----	-----	-----	-----
PANICUMLANIPES	AA--GTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCGA-----
PANICUMSTAPFIANUM	AA--GTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCGA-----
Eleusinecoracana	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----A
Eleusineindica	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----A
Cynodondactylon	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----A
Cynodondactylon7480	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----A
Chlorisvirgata6616	AGGTGTAATT	ACGTTGTGTT	GGTAGCGAAA	C----	TCCC	TCTC-----
Chlorisvirgata7498	AGGTGTAATT	ACGTTGTGTT	GGTAGCGAAA	C----	TCCC	TCTA-----A
Eustachyspaspaloides5597	NNNNNNNNNT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----A
Harpochloafalx6530	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TTTAATTTAA
Harpochloafalx6670	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TTTAATTTAA
Harpochloafalx5118	AGGTGTAATT	ACTTTGTGTT	GGTAGTGAAA	C----	TCCC	TTTAATTTAA
Repliaaltera5133	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----A
Leptochloafusca7622	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----A
Enteropogonmacrostachyus9339	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----A
Dactylocteniumaegyptium9326	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTACTTTAA
Tragusberteronianus6620	AGGTGTAATT	ACGTTGTGTT	GGTAGTGCAA	C----	TTCC	TCTA-----A
Tragusracemosus7108	AGGTGTAATT	ACGTTGTGTT	GGTAGTGCAA	C----	TTCC	TCTA-----A
Traguskoelerioides6729	AGGTGTAATT	ACGTTGTGTT	GGTAGTGCAA	C----	TTCC	TCTA-----A
Polevansiarigida7621	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----A
Perotispatens9323	AGGTATAATT	ACGTTGTGTG	GGTAGTGAAC	C----	TTCC	TCTA-----A
Eragrostiscurvula6529	AGGTGTAATT	ACGTTGTGTT	GGTARTGAAA	S----	TCCC	TCTA-----
Eragrostisobtusa6623	AGGTGTAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Eragrostisobtusa6732	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostisechinochloidea7485	AGGTGTAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Eragrostispseudobtusa7508	AGGTGTAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Eragrostislehmanniana20	AGGTGTAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Eragrostischloromelas6947	AGGTGTAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Eragrostischloromelas7474	AGGTGTAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Eragrostisracemosa6533	AGGTGTAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Eragrostisbiflora6705	AGGTGTAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Eragrostissuperba7470	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	G----	TCCC	TCTA-----
Eragrostiscapensis6551	AGGTGTAATA	-CGTTGTGTT	GGTAGTGAAA	C----	TTCC	TCTA-----
Eragrostiscapensis6532	AGGTGTAATA	TCGTTGTGTT	GGTAGTGAAT	C----	TTCC	TCTA-----
Eragrostislehmanniana7514	AGGTGTAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Pogonarthriasquarrosa7483	AGGTGTAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Pogonarthriasquarrosa7504	AGGTGTAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Catalepisgracilis6962	AAGTGAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Catalepisgracilis6653	AAGTGAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Cladoraphiscyperoides5704	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----
Cladoraphisspinosa6335	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----
Cladoraphiscyperoides4894	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TNCC	TCTA-----
Fingerhuthiaafricana7507	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	CTAAACTCCC		TCTA-----
Entoplocamiaaristulata9353	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----
Enneapogonscoparius18	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----
Enneapogoncenchroides7506	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----
Enneapogonscoparius7486	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----
Schmidtiaappophoroides6334	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----
Schmidtialihariensis7490	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----
Schmidtialihariensis7487	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----
Sporobolusaffricanus7476	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----A
Sporobolusconsimilis9354	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCTA-----A
Sporobolusfimbriatus7509	AGGTGTAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Sporoboluspyramidalis6966	AGGTGTAATT	ACGTTGTGTT	GGTAATGAAA	C----	TCCC	TCTA-----
Sporobolusfimbriatus9328	AGGTGTAATT	ACGTTGTGTT	GGTAGTGAAA	C----	TCCC	TCAA-----A

PHRAGMITESAUSTRALIS6557	-----	---AATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
ARISTIDAADSCENSIONIS6540	-----	---AATTAGA	GAAAGAAAAA	GAAGGGCTTT	ATACATCTAA
KARROOCHLOAPURPUREA6241	-----	---AATTAGA	GAAAGAA---	---GGGCTT-	ATACATCTAA
CHAETOBROMUSINVOLUCRATES6284	-----	---AATTAGA	GAAAGAG---	---GGGCTT-	ATACATCTAA
SORGHUMHALEPENSE	-----				
IMPERATACYLINDRICA	-----				
PANICUMLANIPES	-----	---AATTATA	GAAAGAA---	---GGGCTTT	ATACATCTAA
PANICUMSTAFFIANUM	-----	---AATTATA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Eleusinecoracana	ATTAC----	-TAAATTAGA	GAAAGAA---	---GAGCTTT	ATACATCTAA
Eleusineindica	ATTAC----	-TAAATTAGA	GAAAGAA---	---GAGCTTT	ATACATCTAA
Cynodondactylon	ATTAC----	-TAAATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Cynodondactylon7480	ATTAC----	-TAAATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Chlorisvirgata6616	-----	---TAAATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Chlorisvirgata7498	ATTAC----	-TAAATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Eustachyaspaloides5597	ATTACTAAAA	CTAAATTAGA	GAAAGAA---	---GGGCTTT	ATCCATCTAA
Harpochloafalx6530	ATTAC----	-TAAATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Harpochloafalx6670	ATTAC----	-TAAATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Harpochloafalx5118	ATTAC----	-TAAATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Rendliaaltera5133	ATTAT----	-TAAATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Leptochloaferusa7622	ATTAC----	-TAAATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Enteropogonmacrostachyus9339	ATTAC----	-TAAATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Dactylocteniumaegyptium9326	A--AC----	-GAAATTAGA	GAAAGAA---	---GGGCTTT	ATACATCGAA
Tragusberteronianus6620	ATTAC----	-TAAATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Tragusracemosus7108	ATTAC----	-TAAATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Traguskoelerioides6729	ATTAC----	-TAAATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Polevansiarigida7621	ATTAC----	-TAAATTAGA	GAAAGAA---	---AGGCTTT	ATACATCTAA
Perotispatens9323	ATTAC----	-TAAATTAGA	GAAAGAA---	---GGGCTTT	ATACACCTAA
Eragrostiscurvula6529	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	ATACAT-TAA
Eragrostisobtusa6623	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Eragrostisobtusa6732	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostisechinochloidea7485	-----	---TTTGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Eragrostispseudobtusa7508	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Eragrostislehmanniana20	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Eragrostischloromelas6947	-----	---TTTGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Eragrostischloromelas7474	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Eragrostisracemosa6533	-----	---AATTGGA	G-----		
Eragrostisbiflora6705	-----	---AATTGGA	GAAAGAG---	---GGGCTTT	ATACATCTAA
Eragrostissuperba7470	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	AT----CTAA
Eragrostiscapensis6551	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Eragrostiscapensis6532	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Eragrostislehmanniana7514	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Pogonarthriasquarrosa7483	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Pogonarthriasquarrosa7504	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Catalepisgracilis6962	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Catalepisgracilis6653	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Cladoraphiscyperoides5704	-----	---AATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Cladoraphisspinosa6335	-----	---AATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Cladoraphiscyperoides4894	-----	---AATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Fingerhuthiaafricana7507	-----	---AATTAGA	GAAAGAA---	---GGGCTTT	ATATATCTAA
Entoplocamiaaristulata9353	-----	---AATTAGA	GAAAGAA---	---GC-CTTT	ATACATCTAA
Enneapogonscoparius18	-----	---AATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Enneapogoncenchroides7506	-----	---AATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Enneapogonscoparius7486	-----	---AATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Schmidtiaappophoroides6334	-----	---AATTCGA	GAAAGAA---	---GGGCTGT	ATACATCTAA
Schmidtiaalihariensis7490	-----	---AATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Schmidtiaalihariensis7487	-----	---AATTAGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Sporobolusafricanus7476	ATTAC----	-TAAATTAGA	GAAAGAA---	---GAACCTT	ATACATCTAA
Sporobolusconsimilis9354	ATTAC----	-TAAATTAGA	GAAAGAA---	---GGACTTT	ATACATCTAA
Sporobolusfimbriatus7509	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Sporoboluspyramidalis6966	-----	---AATTGGA	GAAAGAA---	---GGGCTTT	ATACATCTAA
Sporobolusfimbriatus9328	ATTAC----	-TAAATTAAA	GAAAGAA---	---GGACTTT	ATACATTTAA

<i>PHRAGMITESAUSTRALIS</i> 6557	TACACACCTA	TAGATACTGA	CATAGCAAAT	GATTAATCAC	AGAACCCATA
<i>ARISTIDAADSCENSIONIS</i> 6540	TACACACGTA	TAGATACTGA	CATAGCAAAC	GACTAATCAC	AAAACCCATA
<i>KARROOCHLOAPURPUREA</i> 6241	TACACACGTA	TAGATACTGA	CATAGGAAAC	GATTAATCAC	CGAACCTATA
<i>CHAETOBROMUSINVOLUCRATES</i> 6284	TACACACGTA	TAGATACTGA	CATAGGAAAC	GATTAATCAC	AGAACCCATA
<i>SORGHUMHALEPENSE</i>	-----	-----	----AATAAC	GATTAATCAC	AGAACCCATA
<i>IMPERATACYLINDRICA</i>	-----	-----	----AATAAC	GATTAATCAC	AGAACCCATA
<i>PANICUMLANIPES</i>	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAT	AGAACCCATA
<i>PANICUMSTAPFIANUM</i>	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAT	AGAACCCATA
<i>Eleusinecoracana</i>	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Eleusineindica</i>	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Cynodondactylon</i>	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Cynodondactylon</i> 7480	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Chlorisvirgata</i> 6616	TACACACGTA	TACATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Chlorisvirgata</i> 7498	TACACACGTA	TACATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Eustachyspaspaloides</i> 5597	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Harpochloafalx</i> 6530	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Harpochloafalx</i> 6670	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Harpochloafalx</i> 5118	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Rendliaaltera</i> 5133	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Leptochloafusca</i> 7622	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Enteropogonmacrostachyus</i> 9339	TACACACGTA	TAGATACTGA	CATGGCAAAC	GATTAATCAC	AGAACCCATA
<i>Dactylocteniumaegyptium</i> 9326	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Tragusberteronianus</i> 6620	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Tragusracemosus</i> 7108	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Traguskoelerioides</i> 6729	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Polevansiarigida</i> 7621	TACACACGTA	TAGATACTGA	GATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Perotispatens</i> 9323	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCCC	AGAACCCATA
<i>Eragrostiscurvula</i> 6529	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCMTA
<i>Eragrostisobtusata</i> 6623	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Eragrostisobtusata</i> 6732	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
<i>Eragrostisechinochloidea</i> 7485	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Eragrostispseudobtusata</i> 7508	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Eragrostislehmanniana</i> 20	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Eragrostischloromelas</i> 6947	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Eragrostischloromelas</i> 7474	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Eragrostisracemosa</i> 6533	-----	-----	-----	-----AC	AGAACCCGTA
<i>Eragrostisbiflora</i> 6705	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Eragrostisssuperba</i> 7470	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCCTA
<i>Eragrostiscapensis</i> 6551	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAT	AGAACCCATA
<i>Eragrostiscapensis</i> 6532	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAT	AGAACCCATA
<i>Eragrostislehmanniana</i> 7514	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Pogonarthriasquarrosa</i> 7483	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Pogonarthriasquarrosa</i> 7504	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Catalepisgracilis</i> 6962	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Catalepisgracilis</i> 6653	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Cladoraphiscyperoides</i> 5704	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Cladoraphis spinosa</i> 6335	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Cladoraphiscyperoides</i> 4894	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Fingerhuthiaafricana</i> 7507	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Entoplocamia aristulata</i> 9353	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCC- TA
<i>Enneapogon scoparius</i> 18	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Enneapogon cenchroides</i> 7506	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Enneapogon scoparius</i> 7486	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Schmidtia pappophoroides</i> 6334	TACACACGTA	TAGATACTGA	AATAG----	GATTAATCAC	AGAACCCATA
<i>Schmidtia kalihariensis</i> 7490	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Schmidtia kalihariensis</i> 7487	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Sporobolus africanus</i> 7476	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCGTA
<i>Sporobolus consimilis</i> 9354	TACACACGTA	TAGATGCTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Sporobolus fimbriatus</i> 7509	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Sporobolus pyramidalis</i> 6966	TACACACGTA	TAGATACTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA
<i>Sporobolus fimbriatus</i> 9328	TACACACGTA	TAGATGCTGA	CATAGCAAAC	GATTAATCAC	AGAACCCATA

PHRAGMITESAUSTRALIS6557	TCATAATAT- ----AGGTT	CTTTATTTAT	TTATTTTTT	AGAATGAAAT
ARISTIDAADSCENSIONIS6540	TCATAATAT- ----AGGTT	CTTT-----	-TTTTTTTTT	AGAATGAAAT
KARROOCHLOAPURPUREA6241	TCATAATAT- ----AGGTT	CTTTATTTT-	-TTTTTTTTT	AGAATGAAAT
CHAETOBROMUSINVOLUCRATES6284	TCATAATAT- ----AGGTT	CTTTATTTT-	----TTTTTT	AGAATGAAAT
SORGHUMHALEPENSE	TTATAATAT- ----AGGTT	CTTTATTTT-	----ATTTTG	AGAATGAAAT
IMPERATACYLINDRICA	TTATAATAT- ----AGGTT	CTTTATTTT-	----ATTTTT	AGAATGAAAT
PANICUMLANIPES	TCATAATAT- ----AGGTT	CTTTATTTT-	----ATTTTT	AGAATGAAAT
PANICUMSTAPFIANUM	TCATAATAT- ----AGGTT	CTTTATTTT-	----ATTTTT	AGAATGAAAT
Eleusinecoracana	CCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Eleusineindica	CCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Cynodondactylon	CCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Cynodondactylon7480	CCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Chlorisvirgata6616	CGATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Chlorisvirgata7498	CGATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Eustachyaspaspaloides5597	CCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Harpochloafalx6530	CCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AAAATGAAAT
Harpochloafalx6670	CCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTG	AAAATGAAAT
Harpochloafalx5118	CCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTA	AAAATGAAAT
Rendliaaltera5133	CCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Leptochloafusca7622	CCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Entopogonmacrostachyus9339	CCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Dactylocteniumaegyptium9326	CCATAATAT- ----AGGTT	CTTYATTTA-	----TTTTTT	AGAATGAAAT
Tragusberteronianus6620	CCATAATAT- ----AGGTT	CTTTATTTT-	----TTTTTT	AGAATGAAAT
Tragusracemosus7108	CCATAATAT- ----AGGTT	CTTTATTTT-	----TTTTTT	AGAATGAAAT
Traguskoelerioides6729	CCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Polevansiariigida7621	CCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Perotispates9323	CCATAATAT- AGTATAGGTT	CCTTATTTA-	----TTTTTT	AGAATGAAAT
Eragrostiscurvula6529	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Eragrostisobtusa6623	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Eragrostisobtusa6732	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostisechinochloidea7485	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Eragrostispseudobtusa7508	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Eragrostislehmanniana20	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Eragrostischloromelas6947	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Eragrostischloromelas7474	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Eragrostisracemosa6533	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Eragrostisbiflora6705	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	TGAATGAAAT
Eragrostissuperba7470	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Eragrostiscapensis6551	TCATAATAT- ----AGGTT	CCTTATTTA-	----TTTTTT	AGAATGAAAT
Eragrostiscapensis6532	TCATAATAT- ----AGGTT	CCTTATTTA-	----TTTTTT	AGAATGAAAT
Eragrostislehmanniana7514	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Pogonarthriasquarrosa7483	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Pogonarthriasquarrosa7504	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Catalepisgracilis6962	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Catalepisgracilis6653	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Cladoraphiscyperoides5704	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Cladoraphisspinosa6335	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Cladoraphiscyperoides4894	TCATAATAT- ----AGGTT	CTTTATTTA-	----ATTTTT	AGAATGAAAT
Fingerhuthiaafricana7507	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Entoplocamiae aristulata9353	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Enneapogonscoparius18	TCATAATATA TAT--AGGTT	CTTTATTTA-	-TTTTTTTTT	A-AATGAAAT
Enneapogoncenchroides7506	TCATAATATA T---AGGTT	CTTTATTTAT	TTTTTTTTTT	AGAATGAAAT
Enneapogonscoparius7486	TCATAATATA TAT--AGGTT	CTTTATTTA-	-TTTTTTTTT	A-AATGAAAT
Schmidtia pappophoroides6334	TCATAATATA TAT--AGGTT	CTTTATTTT-	----TTTTTT	AGAATGAAAT
Schmidtia kalihariensis7490	TCATAATATA TATATAGGTT	CTTTATTTA-	----TTTTTT	AAAATGAAAT
Schmidtia kalihariensis7487	TCATAATATA TATATAGGTT	CTTTATTTA-	----TTTTTT	AAAATGAAAT
Sporobolusafricanus7476	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Sporobolus consimilis9354	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Sporobolus fimbriatus7509	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Sporobolus pyramidalis6966	TCATAATAT- ----AGGTT	CTTTATTTA-	----TTTTTT	AGAATGAAAT
Sporobolus fimbriatus9328	TCATAATAT- ----AGGTT	TTTTATTTA-	----TTTTTT	AGAATGAAAT



PHRAGMITESAUSTRALIS6557	TAGGAATGAT	TATGAAATAG	AAAATTCTGA	A-TTTTTT-TT	-AGAATTGT-
ARISTIDAADSCENSIONIS6540	TAGGGATGAT	TATGAAATAG	AAAATTCAGA	A-TTTTTT---	-GGAATTAT-
KARROOCHLOAPURPUREA6241	TAGAAATGAA	TATGAAATAG	AAAATTCTGA	AATTTTTT-TT	-AGAATTAT-
CHAETOBROMUSINVOLUCRATES6284	TAGAAATGAA	TATGAAATAG	AAAATTCAGA	A-TTTTTT-TT	-CGAATTAT-
SORGHUMHALEPENSE	TAGGAATGAT	TATGAAATAG	AAAATTCATA	A-TTTTTT-TT	-AGAATTAT-
IMPERATACYLINDRICA	TAGGAATGAT	TATGAAATAG	AAAATTCCTA	A-TTTTTT-TT	-AGAATTAT-
PANICUMLANIPES	TAGGAATGAT	TATGAAATAT	AAAATTCCTGA	A-TTTTTT-TT	TAGAATTAT-
PANICUMSTAPFIANUM	TAGGAATGAT	TATGAAATAT	AAAATTCCTGA	A-TTTTTT-TT	TAGAATTAT-
Eleusinecoracana	TTGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-AGAATTAT-
Eleusineindica	TTGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-AGAATTAT-
Cynodondactylon	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-AGAATTAT-
Cynodondactylon7480	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-AGAATTAT-
Chlorisvirgata6616	TAGGAATGAT	TATGAAATAG	AAAATTCCTAA	A-TTTTTT-T-	-AGAATTATA
Chlorisvirgata7498	TAGGAATGAT	TATGAAATAG	AAAATTCCTAA	A-TTTTTT-T-	-AGAATTATA
Eustachyospaloides5597	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-AGAATTAT-
Harpochloafalx6530	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-AGAATTAT-
Harpochloafalx6670	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-AGAATTAT-
Harpochloafalx5118	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-AGAATTAT-
Rendliaaltera5133	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	TAGAATTAT-
Leptochloafusca7622	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-GGAATTAT-
Enteropogonmacrostachyus9339	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-AGAATTAT-
Dactylocteniumaegyptium9326	TAGGAATGAT	TATGAAATAG	AAAATTTTGA	A-TTTTTT-TT	-AGAATTAT-
Tragusberteronianus6620	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-CGAATTAT-
Tragusracemosus7108	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-CGAATTAT-
Traguskoelerioides6729	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-CGAATTAT-
Polevansiarigida7621	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-AGAATTAT-
Perotispatis9323	TAGGAATGAT	TATGAAATAG	GAAATTCCTGA	A-TTTTTT-TT	-AGAATTAT-
Eragrostiscurvula6529	TAGGAATGAT	TATGAAATAK	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Eragrostisobtusa6623	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Eragrostisobtusa6732	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostisechinocloidea7485	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Eragrostispseudobtusa7508	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Eragrostislehmanniana20	TAGGAATGAT	TATGAAATAK	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Eragrostischloromelas6947	TAGGAATGAT	TATGAAATAT	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Eragrostischloromelas7474	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Eragrostisracemosa6533	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Eragrostisbiflora6705	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Eragrostissuperba7470	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Eragrostiscapensis6551	TAGGAGTGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	TAGAATTAT-
Eragrostiscapensis6532	TAGGAGTGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	TAGAATTAT-
Eragrostislehmanniana7514	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Pogonarthriasquarrosa7483	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Pogonarthriasquarrosa7504	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Catalepisgracilis6962	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Catalepisgracilis6653	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Cladoraphiscyperoides5704	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Cladoraphisspinosa6335	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Cladoraphiscyperoides4894	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Fingerhuthiaafricana7507	TAGGAATGAT	TATGAAATAG	CAAATTCCTGC	A-TTTTTTATT	-AGAATTAT-
Entoplocamiaaristulata9353	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Enneapogonscoparius18	TAGGAATGAT	TATGAAATAG	AAAATTCCTGC	A-TTTTTTATT	-AGAATTAT-
Enneapogoncenchroides7506	TAGGAATGAT	TATGAAATAG	AAAATTCCTGC	A-TTTTTTATT	-AGAATTAT-
Enneapogonscoparius7486	TAGGAATGAT	TATGAAATAG	AAAATTCCTGC	A-TTTTTTATT	-AGAATTAT-
Schmidtiaappophoroides6334	TAGGAATGAT	TATGAAATAG	AAAATTCCTTA	A-TTTTTTATT	-AGAATTAT-
Schmidtiaakalihariensis7490	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Schmidtiaakalihariensis7487	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Sporobolusafricanus7476	TTGGAATGAT	TATGAAATAA	AAAATTCCTGA	A-TTTTTT-TT	-AGAATTAG-
Sporobolusconsimilis9354	TAGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-TT	-AGAATTAT-
Sporobolusfimbriatus7509	TAGGAATGAT	TATGAAATAT	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Sporoboluspyramidalis6966	TAGGAATGAT	TATGAAATAT	AAAATTCCTGA	A-TTTTTTATT	-AGAATTAT-
Sporobolusfimbriatus9328	TGGGAATGAT	TATGAAATAG	AAAATTCCTGA	A-TTTTTT-T-	-AGAATTAT-

PHRAGMITESAUSTRALIS6557	-----TGTGA	ATCCATTCCA	ATCGAATATT	GAGTAATCAA	ATCCTTCAAT
ARISTIDAADSCENSIONIS6540	-----TGTGA	-----TTCCA	ATCAAATATT	GAGTAATCAA	ATCCTTCAAT
KARROOCHLOAPURPUREA6241	-----TGTGA	ATCCATTCCA	ATTGAATATT	GAGTAATCAA	ATCCTTCAAT
CHAETOBROMUSINVOLUCRATES6284	-----TGTGA	ATCTATTCCA	ATTGAATATT	GAGTAATCAA	ATCCTTCAAT
SORGHUMHALEPENSE	-----TGTGA	ATCTATTCCA	ATCGAATATT	GAGTAATCAA	ATCCTTCAAT
IMPERATACYLINDRICA	-----TGTGA	ATCTATTCCA	ATCGAATATT	GAGTAATCAA	ATCCTTCAAT
PANICUMLANIPES	-----TGTGA	ATCCATTCCA	ATCGAATATT	GAGTAATCAA	ATCCTTCAAT
PANICUMSTAFFIANUM	-----TGTGA	ATCCATTCCA	ATCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eleusinecoracana	-----TGTGA	ATCTATTCCCT	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eleusineindica	-----TGTGA	ATCTATTCCCT	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Cynodondactylon	-----TGTGA	ATCTATTCCA	CTCGAATATT	GAGTAATCAA	ACCCTTCAAT
Cynodondactylon7480	-----TGTGA	ATCTATTCCA	CTCGAATATT	GAGTAATCAA	ACCCTTCAAT
Chlorisvirgata6616	ATTATTGTGA	ATCTATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Chlorisvirgata7498	ATTATTGTGA	ATCTATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eustachyaspaspaloides5597	-----TGTGA	ATCTATTCCA	CTCGAATATT	GAGTAATCAA	ACCCTTCAAT
Harpochloafalx6530	-----TGTGA	ATCTATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Harpochloafalx670	-----TGTGA	ATCTATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Harpochloafalx5118	-----TGTGA	ATCTATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Rendliaaltera5133	-----TGTGA	ATCTATTCCC	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Leptochloafusca7622	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Enteropogonmacrostachyus9339	-----TGTGA	ATCTATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Dactylocteniumaegyptium9326	-----TGTGA	ATCCATTCCC	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Tragusberteronianus6620	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAATAATCAA	ATCCTTCAAT
Tragusracemosus7108	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAATAATCAA	ATCCTTCAAT
Traguskoelerioides6729	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAATAATCAA	ATCCTTCAAT
Polevansiarigida7621	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAATAATCAA	ATCCTTCAAT
Perotispatens9323	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eragrostiscurvula6529	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eragrostisobtusa6623	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eragrostisobtusa6732	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNCAAT
Eragrostisechinochloidea7485	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eragrostispseudobtusa7508	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eragrostislehmanniana20	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eragrostischloromelas6947	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eragrostischloromelas7474	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eragrostisracemosa6533	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eragrostisbiflora6705	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eragrostissuperba7470	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eragrostiscapensis6551	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eragrostiscapensis6532	-----TGTGA	ATCCATTCCC	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Eragrostislehmanniana7514	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Pogonarthriasquarrosa7483	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Pogonarthriasquarrosa7504	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Catalepisgracilis6962	-----TGTGA	ATCCATTCCA	CTCGAATATA	GAGTAATCAA	ATCCTTCAAT
Catalepisgracilis6653	-----TGTGA	ATCCATTCCA	CTCGAATATA	GAGTAATCAA	ATCCTTCAAT
Cladoraphiscyperoides5704	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Cladoraphisspinosa6335	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Cladoraphiscyperoides4894	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Fingerhuthiaafricana7507	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Entoplocamiaaristulata9353	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Enneapogonscoparius18	-----TGTGA	ATCCATTCCA	CTCAAATATT	GAGTAATCAG	ATCCTTCATT
Enneapogoncenchroides7506	-----TGTGA	ATCCATTCCA	CTCAAATATT	GAGTAATCAG	ATCCTTCATT
Enneapogonscoparius7486	-----TGTGA	ATCCATTCCA	CTCAAATATT	GAGTAATCAG	ATCCTTCATT
Schmidtiapappophoroides6334	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCATT
Schmidtiakalihariensis7490	-----TGGGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Schmidtiakalihariensis7487	-----TGGGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Sporobolusafricanus7476	-----TGGAA	TTCCATTCCA	CTCGAATATT	GAATAATCAA	ATCCTTCAAT
Sporobolusconsimilis9354	-----TGTGA	ATCCATTCCA	CTCGAAT---	-----	-----
Sporobolusfimbriatus7509	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Sporoboluspyramidalis6966	-----TGTGA	ATCCATTCCA	CTCGAATATT	GAGTAATCAA	ATCCTTCAAT
Sporobolusfimbriatus9328	-----TGTGA	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN



PHRAGMITESAUSTRALIS6557	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
ARISTIDAADSCENSIONIS6540	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
KARROOCHLOAPURPUREA6241	ACGAGAATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
CHAETOBROMUSINVOLUCRATES6284	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
SORGHUMHALEPENSE	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
IMPERATACYLINDRICA	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
PANICUMLANIPES	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
PANICUMSTAFFIANUM	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eleusinecoracana	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eleusineindica	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Cynodondactylon	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Cynodondactylon7480	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Chlorisvirgata6616	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Chlorisvirgata7498	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eustachyspaspaloides5597	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Harpochloafalx6530	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Harpochloafalx6670	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Harpochloafalx5118	-CGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Rendliaaltera5133	ACGAGGATAA	AGAGAGAGTC	CCATTTTACA	TGTCAATACT	GACAACAATG
Leptochloafusa7622	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostisogonummacrostachyus9339	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Dactylocteniumaegyptium9326	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Tragusberteronianus6620	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Tragusracemosus7108	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Traguskoelerioides6729	GCGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Polevansiarigida7621	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Perotispatens9323	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostiscurvula6529	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostisobtusa6623	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostisobtusa6732	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostisechinoides7485	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostispseudobtusa7508	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostislehmanniana20	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostischloromelas6947	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostischloromelas7474	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostisracemosa6533	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostisbiflora6705	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostissuperba7470	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostiscapensis6551	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostiscapensis6532	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Eragrostislehmanniana7514	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Pogonarthriasquarrosa7483	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Pogonarthriasquarrosa7504	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Catalepisgracilis6962	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Catalepisgracilis6653	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Cladoraphiscyperoides5704	ACGAGGATAA	AGAGAGAGTC	CCATTTTACA	TGTCAATACT	GACAACAATG
Cladoraphisspinosa6335	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Cladoraphiscyperoides4894	GCGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Fingerhuthiaafricana7507	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Entoplocamiaaristulata9353	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Enneapogonscoparius18	ACGAGGATAA	AGAGAGAGTC	CCATTTTACA	TGTCAATACT	GACAACAATG
Enneapogoncenchroides7506	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Enneapogonscoparius7486	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Schmidtiaapappophoroides6334	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Schmidtiaakalihariensis7490	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Schmidtiaakalihariensis7487	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Sporobolusafricanus7476	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Sporobolusconsimilis9354	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Sporobolusfimbriatus7509	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATACT	GACAACAATG
Sporoboluspyramidalis6966	ACGAGGATAA	AGAGAGAGTC	CCATTCTACA	TGTCAATGGT	GACAACAATG
Sporobolusfimbriatus9328	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNCAATG

PHRAGMITESAUSTRALIS6557  
 ARISTIDAADSCENSIONIS6540  
 KARROOCHLOAPURPUREA6241  
 CHAETOBROMUSINVOLUCRATES6284  
 SORGHUMHALEPENSE  
 IMPERATACYLINDRICA  
 PANICUMLANIPES  
 PANICUMSTAPFIANUM  
*Eleusinecoracana*  
*Eleusineindica*  
*Cynodondactylon*  
*Cynodondactylon7480*  
*Chlorisvirgata6616*  
*Chlorisvirgata7498*  
*Eustachyspaspaloides5597*  
*Harpochloafalx6530*  
*Harpochloafalx6670*  
*Harpochloafalx5118*  
*Rendliaaltera5133*  
*Leptochloafusca7622*  
*Enteropogonmacrostachyus9339*  
*Dactylocteniumaegyptium9326*  
*Tragusberteronianus6620*  
*Tragusracemosus7108*  
*Traguskoelerioides6729*  
*Polevansiarigida7621*  
*Perotispatens9323*  
*Eragrostiscurvula6529*  
*Eragrostisobtusa6623*  
*Eragrostisobtusa6732*  
*Eragrostispechinochloidea7485*  
*Eragrostispseudobtusa7508*  
*Eragrostislehmanniana20*  
*Eragrostischloromelas6947*  
*Eragrostischloromelas7474*  
*Eragrostisracemosa6533*  
*Eragrostisbiflora6705*  
*Eragrostissuperba7470*  
*Eragrostiscapensis6551*  
*Eragrostiscapensis6532*  
*Eragrostislehmanniana7514*  
*Pogonarthriasquarrosa7483*  
*Pogonarthriasquarrosa7504*  
*Catalepisgracilis6962*  
*Catalepisgracilis6653*  
*Cladoraphiscyperoides5704*  
*Cladoraphisspinosa6335*  
*Cladoraphiscyperoides4894*  
*Fingerhuthiaafricana7507*  
*Entolopocamiaaristolata9353*  
*Enneapogonscoparius18*  
*Enneapogoncenchroides7506*  
*Enneapogonscoparius7486*  
*Schmidtiaappophoroides6334*  
*Schmidtiaakalihariensis7490*  
*Schmidtiaakalihariensis7487*  
*Sporobolusafricanus7476*  
*Sporobolusconsimilis9354*  
*Sporobolusfimbriatus7509*  
*Sporoboluspyramidalis6966*  
*Sporobolusfimbriatus9328*

PHRAGMITESAUSTRALIS6557	AAGTCCCTCT	ATCCCCAAAC	CCTCTTTTAT	TTACTAACCC	TAGTATTTAT
ARISTIDAADSCENSIONIS6540	AAGTCCCTCT	ATCCCCAAAC	CCTCTTTTAT	TCCCTAACCA	TAGTATTTAT
KARROOCHLOAPURPUREA6241	AAGTCCCTCT	ATCCCCAAAC	CCTCTTTTAT	TCCCTAACCA	TAGTATTTAT
CHAETOBROMUSINVOLUCRATES6284	AAGTCCCTCT	ATCCCCAAAC	CCTCGTTTAT	TCCCTAACCA	TAGTATTTAT
SORGHUMHALEPENSE	AAGTCCCTCT	ATCCCCAAAC	CCTCTTTTAT	TCGCTAACCA	TAGTTGTTAT
IMPERATACYLINDRICA	AAGTCCCTCT	ATCCCCAAAC	CCTCTTTTAT	TCCCTAACCA	TAGTTGTTAT
PANICUMLANIPES	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMSTAFFIANUM	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon7480	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Chlorisvirgata6616	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Chlorisvirgata7498	AAGTCCCTTT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Eustachyspaspaloides5597	NNNNNNNNNT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Harpochloafalx6530	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Harpochloafalx6670	AAGTCCCTTT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Harpochloafalx5118	AAGTCCCTTT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Rendliaaltera5133	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTAAT
Leptochloafusca7622	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCG	TAGTATTTAT
Enteropogonmacrostachyus9339	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Dactylocteniumaegyptium9326	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Tragusberteronianus6620	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	-AC--TTTAT
Tragusracemosus7108	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	-AC--TTTAT
Traguskoelerioides6729	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	-AC--TTTAT
Polevansiarigida7621	AAGTCCCTTT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	-A-----
Perotispates9323	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAATCA	TAGTATTTAT
Eragrostiscurvula6529	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Eragrostisobtusata6623	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Eragrostisobtusata6732	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Eragrostislechmanniana7485	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Eragrostispseudobtusata7508	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Eragrostislehmanniana20	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Eragrostischloromelas6947	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Eragrostischloromelas7474	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Eragrostisracemosa6533	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Eragrostisbiflora6705	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TACTATTTAT
Eragrostissuperba7470	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TCGTATTTAT
Eragrostiscapensis6551	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Eragrostiscapensis6532	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Eragrostislehmanniana7514	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Pogonarthriasquarrosa7483	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Pogonarthriasquarrosa7504	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Catalepisgracilis6962	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNCC	TAGTATTTAT
Catalepisgracilis6653	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Cladoraphiscyperoides5704	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Cladoraphisspinosa6335	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Cladoraphiscyperoides4894	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Fingerhuthiaafricana7507	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Entollocamiae aristulata9353	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Enneapogonscoparius18	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTAAT
Enneapogoncenchroides7506	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTAAT
Enneapogonscoparius7486	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTAAT
Schmidtia pappophoroides6334	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTAAT
Schmidtia kalihariensis7490	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAAT
Schmidtia kalihariensis7487	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAAT
Sporobolus africanus7476	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Sporobolus consimilis9354	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Sporobolus fimbriatus7509	AAGTCCCTCT	ATCCCCAAAC	CCTCCTTTAT	TCCCTAACCA	TAGTATTTAT
Sporobolus pyramidalis6966	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Sporobolus fimbriatus9328	AAGTCCCTCT	ATCCCCAAAC	CCCCTTTAT	TCCCTAACCA	TAGTATTTAT

PHRAGMITESAUSTRALIS6557	-----CCTTT	TTTTCTTTT	AT-----CAA	TGGGTTT---	--AAGATTCA
ARISTIDAADSCENSIONIS6540	-----CCTCT	TTTTT-TTT-	ATGAATGCAA	TGGGTTT---	--AAGATTCA
KARROOCHLOAPURPUREA6241	-----CCTAT	TTTTT-TTT-	-----CAA	TAGGTTT---	--AAGATTCA
CHAETOBROMUSINVOLUCRATES6284	-----CCTAT	TTTTT-TTTT	-----CAA	TGGGTTT---	--AAGATTCA
SORGHUMHALEPENSE	-----CCT-T	TTTTT---TT	AT-----CAA	TGGGTTT---	--AAGATTCA
IMPERATACYLINDRICA	-----CCT-T	TTTTCTTTT	AT-----CAA	TGGGTTT---	--AAGATTCA
PANICUMLANIPES	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMSTAPFIANUM	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon7480	-----CCTCT	TTTCTCTTT	AT-----CTA	TGG-TTCAAG	ACAAGATTCA
Chlorisvirgata6616	-----CCTCT	TTTCTCTTT	AT-----CGA	TGGGTT---	-CAAGATTCA
Chlorisvirgata7498	-----CCTCT	TTTCTCTTT	AT-----CGA	TGGGTT---	-CAAGATTCA
Eustachyspaspaloides5597	-----CCTCT	TTTCTCTTT	AT-----CAA	TGGGTTCAAG	ACAAGATTCA
Harpochoafalx6530	-----CCTCT	TTTCTCTTT	AT-----CAA	TGGGTT---	-CAAGATTCA
Harpochoafalx6670	-----CCTCT	TTTCTCTTT	AT-----CAA	TGGGTT---	-CAAGATTCA
Harpochoafalx5118	-----CCTCT	TTTCTCTTT	AT-----CAA	TGGGTT---	-CAAGATTCA
Rendliaaltera5133	-----CCTCT	TTYTCTTTT	MT-----CAA	TGGGTTYAAG	ACAAGATTCA
Leptochloafusca7622	CTTATCCTCT	TTTCTCTTT	AT-----CAA	TGGGTT---	-CAAGATTCA
Enteropogonmacrostachyus9339	-----CCTCT	TTTCTCTTT	AT-----CAA	TGGGTT---	-CAAGATTCA
Dactylocteniumaegyptium9326	-----CCTCT	TTTCTCTTT	AT-----CAA	TGGGTT---	-CAAGATTCA
Tragusberteronianus6620	-----	-----TCCCTA	AC-----CAA	TGGGTT---	-CAAGATTCA
Tragusracemosus7108	-----	-----TCCCTA	AC-----CAA	TGGGTT---	-CAAGATTCA
Traguskoelerioides6729	-----	-----TCCCTA	AC-----CAA	TGGGTT---	-CAAGATTCA
Polevansiarigida7621	-----	-----	-----	TGGGTT---	-CAAGATTCA
Perotispatens9323	CTTATCCTCT	TTTCTCTTT	AT-----CAA	TGGGTT---	-CAAGATTCA
Eragrostiscurvula6529	-----CCTAT	TTTTCTTTT	AT-----CAA	TGGATTT---	-CAAGATTCA
Eragrostisobtusa6623	-----CCTAT	TTTTCTTTT	ATCAATGCAA	TGGGTTT---	-CAAGATTCA
Eragrostisobtusa6732	-----CCTAT	TTTTCTTTT	ATCAATGCAA	TGGGTTT---	-CAAGATTCA
Eragrostisechinochloidea7485	-----CCTAT	TTTATCTTT	ATCAATGCAA	TGGGTTT---	-CAAGATTCA
Eragrostispseudobtusa7508	-----CCTAT	TTTTCTTTT	ATCAATGCAA	TGGGTTT---	-CAAGATTCA
Eragrostislehmanniana20	-----CCTAT	TTTTCTTTT	AT-----CAA	TGGATTT---	-CAAGATTCA
Eragrostischloromelas6947	-----CCTAT	TTTTCTTTT	AT-----CAA	TGGATTT---	-CAAGATTCA
Eragrostischloromelas7474	-----CCTAT	TTTTCTTTT	AT-----CAA	CGGGTTT---	-CAAGATTCA
Eragrostisracemosa6533	-----CCTAT	TTTTCTTTT	AT-----CAA	TGGGTTT---	-CAAGATTCA
Eragrostisbiflora6705	-----CCTAT	TTTTCTTTT	AT-----CAA	TGGGTTT---	-CAAGATTCA
Eragrostissuperba7470	-----CCTCT	TTTTCTTTT	AG-----CAA	GGGGTTT---	-CAAGATTCA
Eragrostiscapensis6551	-----CCTAT	TTTTCTTTT	AT-----CAA	TGGGTTT---	-CAAGATTCA
Eragrostiscapensis6532	-----CCTAT	TTTTCTTTT	AT-----CAA	TGGGTTT---	-CAAGATTCA
Eragrostislehmanniana7514	-----CCTAT	TTTTCTTTT	AT-----CAA	TGGATTT---	-CAAGATTCA
Pogonarthriasquarrosa7483	-----CCTAT	TTTTCTTTT	AT-----CAA	TGGATTT---	-CAAGATTCA
Pogonarthriasquarrosa7504	-----CCTAT	TTTTCTTTT	AT-----CAA	TGGATTT---	-CAAGATTCA
Catalepisgracilis6962	-----CCTTT	TTTTCTTTT	AT-----CAA	TGGGTTT---	-CAAGATTCA
Catalepisgracilis6653	-----CCTTT	TTTTCTTTT	AT-----CAA	TGGGTTT---	-CAAGATTCA
Cladoraphiscyperoides5704	-----CCTCT	TTTTCTTTT	AT-----CAA	TGGGTTT---	-CAAGATTCA
Cladoraphisspinosa6335	-----CCTCT	TTTTCTTTT	AT-----CAA	TGGGTTT---	-CAAGATTCA
Cladoraphiscyperoides4894	-----CCTCT	TTTTCTTTT	AT-----CAA	TGGGTTT---	-CAAGATTCA
Fingerhuthiaafricana7507	-----CCTCT	TTTTCTTTT	AT-----CAA	TGGGTTT---	-CAAGATTCA
Entoplocamiaaristulata9353	-----CCTCT	TTTTCTTTT	AT-----CAA	TGGGTTT---	-CAAGATTCA
Enneapogonscoparius18	-----CCTCT	TTTTCTTTT	CT-----CAA	TGGGTTT---	-CAAGATTCA
Enneapogoncenchroides7506	-----CCTCT	TTTTCTTTT	CT-----CAA	TGGGTTT---	-CAAGATTCA
Enneapogonscoparius7486	-----CCTCT	TTTTCTTTT	CT-----CAA	TGGGTTT---	-CAAGATTCA
Schmidtiaappophoroides6334	-----CCTCT	TTTTCTTTT	CT-----CAA	TGGGTTT---	-CAAGATTCA
Schmidtiaalihariensis7490	-----CCTCT	TTTTCTTTT	AT-----CAA	TGGGTTT---	-CAAGATTCA
Schmidtiaalihariensis7487	-----CCTCT	TTTTCTTTT	AT-----CAA	TGGGTTT---	-CAAGATTCA
Sporobolusafricanus7476	CCTATCCTCT	TTTTCTTTT	AT-----TAA	-----	-----
Sporobolusconsimilis9354	-----CCTCT	TTTTCTTTT	AT-----CAA	TGGGTTCAA-	ACAAAATTCA
Sporobolusfimbriatus7509	-----CCTAT	TTTTCTTTT	AT-----CAA	TGGATTT---	-CAAGATTCA
Sporoboluspyramidalis6966	NNNNNNNNNN	NNNNNNNTTT	AT-----CAA	TGGATTT---	-CAAGATTCA
Sporobolusfimbriatus9328	-----CCTCT	TTTTCTTTT	AT-----CAA	TGGGTT---	-CGAGATTCA

PHRAGMITESAUSTRALIS6557	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCGAAGA
ARISTIDAADSCENSIONIS6540	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTCCGAAGA
KARROOCHLOAPURPUREA6241	TTAGCT----	TTCTCATTCT	ATT-----CT	TTCACAAAGG	ACTGCAAAGA
CHAETOBROMUSINVOLUCRATES6284	TTAGCT----	TTCTCATTCT	ATT-----CA	TTCACAAAGG	AATGCAAAGA
SORGHUMHALEPENSE	CTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCGACAA
IMPERATACYLINDRICA	CTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCGACAA
PANICUMLANIPES	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMSTAPFIANUM	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon7480	TTAGCTTGCT	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCAAAGA
Chlorisvirgata6616	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCAAAGA
Chlorisvirgata7498	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCAAAGA
Eustachyspaspaloides5597	TTAGTT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCAAAGA
Harpochloafalx6530	TTAGTT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCAAAGA
Harpochloafalx6670	TTAGTT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCAAAGA
Harpochloafalx5118	TTAGTT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCAAAGA
Rendliaaltera5133	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCAAAGA
Leptochloafusca7622	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCAAAGA
Enteropogonmacrostachyus9339	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCAAAGA
Dactylocteniumaegyptium9326	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	GGTGCGAAGA
Tragusberteronianus6620	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCGAAAA
Tragusracemosus7108	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCGAAAA
Traguskoelerioides6729	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCGAAAA
Polevansiarigida7621	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCGAAGA
Perotispatens9323	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCGAAGA
Eragrostiscurvula6529	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Eragrostisobtusa6623	TTAGCT----	TTCTCATTCT	ACTTTACTCT	TTCATAAAGG	AGTGCGAAGA
Eragrostisobtusa6732	TTAGCT----	TTCTCATTCT	ACTTTACTCT	TTCATAAAGG	AGTGCGAAGA
Eragrostisechinochloidea7485	TTAGCT----	TTCTCATTCT	ACTTTACTCT	TTCATAAAGG	AGTGCGAAGA
Eragrostispseudobtusa7508	TTAGCT----	TTCTCATTCT	ACTTTACTCT	TTCATAAAGG	AGTGCGAAGA
Eragrostislehmanniana20	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Eragrostischloromelas6947	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Eragrostischloromelas7474	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Eragrostisracemosa6533	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Eragrostisbiflora6705	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Eragrostissuperba7470	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Eragrostiscapensis6551	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Eragrostiscapensis6532	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Eragrostislehmanniana7514	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Pogonarthriasquarrosa7483	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Pogonarthriasquarrosa7504	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Catalepisgracilis6962	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Catalepisgracilis6653	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Cladoraphiscyperoides5704	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Cladoraphisspinosa6335	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Cladoraphiscyperoides4894	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Fingerhuthiaafricana7507	TTAGTT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGAGCGAAGA
Entolocamiaaristulata9353	TTAGTT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Enneapogonscoparius18	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCGAAGA
Enneapogoncenchroides7506	TTAGCT----	TTCTCATTCT	ATT-----CT	TTCACAAAGG	AGTGCGAAGA
Enneapogonscoparius7486	TTAGCT----	TTCTCATTCT	ACT-----TT	TTCACAAAGG	AGTGCGAAGA
Schmidtiaappophoroides6334	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCGAAGA
Schmidtiaalihariensis7490	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCGAAGA
Schmidtiaalihariensis7487	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCGAAGA
Sporobolusafricanus7476	-----	-----	-----	-----	-----
Sporobolusconsimilis9354	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCGAAGA
Sporobolusfimbriatus7509	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Sporoboluspyramidalis6966	TTAGTT----	TTCTCATTCT	ACT-----CT	TTCATAAAGG	AGTGCGAAGA
Sporobolusfimbriatus9328	TTAGCT----	TTCTCATTCT	ACT-----CT	TTCACAAAGG	AGTGCGAAGA



PHRAGMITESAUSTRALIS6557	GAACTCAATG	GATCTTATGC	TATTCATTAA	AATGGATCTT	ATGCTATTCA
ARISTIDAADSCENSIONIS6540	GAACTCAATG	GATCTTATGC	TATTCATTAA	A-TGGAT---	-----
KARROOCHLOAPURPUREA6241	GAACTCAATG	GATCTTATGC	TATTCATTAA	A-TGGAT---	-----
CHAETOBROMUSINVOLUCRATES6284	GAACTCAATG	GATCTTATGC	TATTCATTAA	A-TGGAT---	-----
SORGHUMHALEPENSE	GAACTCAATG	AATCTTATGC	TATTCATTAA	A-TAGAT---	-----
IMPERATACYLINDRICA	GAACTCAATG	AATCTTATGC	TATTCATTAA	A-TAGAT---	-----
PANICUMLANIPES	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMSTAPFIANUM	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon7480	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Chlorisvirgata6616	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Chlorisvirgata7498	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Eustachyspaspaloides5597	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Harpochloafalx6530	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Harpochloafalx6670	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Harpochloafalx5118	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Rendliaaltera5133	RAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Leptochloafusca7622	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Enteropogonmacrostachyus9339	GAACTCAATA	AATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Dactylocteniumaegyptium9326	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Tragusberberianus6620	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Tragusracemosus7108	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Traguskoelerioides6729	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Polevansiarigida7621	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Perotispatens9323	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Eragrostiscurvula6529	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Eragrostisobtusa6623	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Eragrostisobtusa6732	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Eragrostisechinochloidea7485	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Eragrostispseudobtusa7508	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Eragrostislehmanniana20	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Eragrostischloromelas6947	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Eragrostischloromelas7474	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Eragrostisracemosa6533	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Eragrostisbiflora6705	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Eragrostis superba7470	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Eragrostiscapensis6551	GAACTCAATA	GATCTTATGT	TATTCAGTAA	A-TAGAT---	-----
Eragrostiscapensis6532	GAACTCAATA	GATCTTATGT	TATTCAGTAA	A-TAGAT---	-----
Eragrostislehmanniana7514	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Pogonarthriasquarrosa7483	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Pogonarthriasquarrosa7504	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Catalepisgracilis6962	GAACTCAATA	GATCTTATGC	TATTCGTTAA	A-TAGAT---	-----
Catalepisgracilis6653	GAACTCAATA	GATCTTATGC	TATTCGTTAA	A-TAGAT---	-----
Cladoraphiscyperoides5704	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Cladoraphisspinosa6335	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Cladoraphiscyperoides4894	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Fingerhuthiaafricana7507	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Entoplocamiaaristulata9353	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Enneapogonscoparius18	AAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Enneapogoncenchroides7506	AAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Enneapogonscoparius7486	AAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Schmidtiaappophoroides6334	AAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Schmidtiaakalihariensis7490	AAACTCAATG	GATCTTATGC	TATTTATTAA	A-TAGAT---	-----
Schmidtiaakalihariensis7487	AAACTCAATG	GATCTTATGC	TATTTATTAA	A-TAGAT---	-----
Sporobolusafricanus7476	-----	-----TATGC	TATTCATTAA	A-TGGAT---	-----
Sporobolusconsimilis9354	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-CGGAT---	-----
Sporobolusfimbriatus7509	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Sporoboluspyramidalis6966	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TAGAT---	-----
Sporobolusfimbriatus9328	GAACTCAATA	GATCTTATGC	TATTCATTAA	A-TGGAT---	-----

<i>PHRAGMITESAUSTRALIS</i> 6557	TTAAATAGAT	TTCTTTTT--	---TATT---	-----	-----
<i>ARISTIDAADSCENSIONIS</i> 6540	-----	TTATTTTT--	---TATT---	-----	-----
<i>KARROOCHLOAPURPUREA</i> 6241	-----	TTCTTTTT--	---TATT---	-----	-----
<i>CHAETOBROMUSINVOLUCRATES</i> 6284	-----	TTCTTTTT--	---TATT---	-----	-----
<i>SORGHUMHALEPENSE</i>	-----GAT	TTCTTTTT--	---TATCTCT	TTTTTTTTTA	TTTATTAGAG
<i>IMPERATACYLINDRICA</i>	-----GAT	TTCTTTTT--	---TATTTCT	TTTTTTTT--	TTTATTT-AT
<i>PANICUMLANIPES</i>	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
<i>PANICUMSTAFIANUM</i>	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
<i>Eleusinecoracana</i>	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
<i>Eleusineindica</i>	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
<i>Cynodondactylon</i>	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
<i>Cynodondactylon</i> 7480	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Chlorisvirgata</i> 6616	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Chlorisvirgata</i> 7498	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Eustachyspaspaloides</i> 5597	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Harpochloafalx</i> 6530	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Harpochloafalx</i> 6670	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Harpochloafalx</i> 5118	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Rendliaaltera</i> 5133	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Leptochloafusca</i> 7622	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Enteropogonmacrostachyus</i> 9339	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Dactylocteniumaegyptium</i> 9326	-----	TGCTTTTT--	---TATT---	-----	-----
<i>Tragusberteronianus</i> 6620	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Tragusracemosus</i> 7108	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Traguskoelerioides</i> 6729	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Polevansiarigida</i> 7621	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Perotispatens</i> 9323	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Eragrostiscurvula</i> 6529	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Eragrostisobtusa</i> 6623	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Eragrostisobtusa</i> 6732	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Eragrostisechinochloidea</i> 7485	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Eragrostispseudobtusa</i> 7508	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Eragrostislehmanniana</i> 20	-----	TTCTTTTTT-	---TATT---	-----	-----
<i>Eragrostischloromelas</i> 6947	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Eragrostischloromelas</i> 7474	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Eragrostisracemosa</i> 6533	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Eragrostisbiflora</i> 6705	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Eragrostissuperba</i> 7470	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Eragrostiscapensis</i> 6551	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Eragrostiscapensis</i> 6532	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Eragrostislehmanniana</i> 7514	-----	TTCTTTTTT-	---TATT---	-----	-----
<i>Pogonarthriasquarrosa</i> 7483	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Pogonarthriasquarrosa</i> 7504	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Catalepisgracilis</i> 6962	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Catalepisgracilis</i> 6653	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Cladoraphiscyperoides</i> 5704	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Cladoraphisspinosa</i> 6335	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Cladoraphiscyperoides</i> 4894	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Fingerhuthiaafricana</i> 7507	-----	TACTTTTT--	---TATT---	-----	-----
<i>Entoplocamiaaristulata</i> 9353	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Enneapogonscoparius</i> 18	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Enneapogoncenchroides</i> 7506	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Enneapogonscoparius</i> 7486	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Schmidtiaappophoroides</i> 6334	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Schmidtiaakalihariensis</i> 7490	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Schmidtiaakalihariensis</i> 7487	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Sporobolusafricanus</i> 7476	-----	TTCTTTTTGA	TTTATT--	-----	-----
<i>Sporobolusconsimilis</i> 9354	-----	TTCTTTTTTA	TTTATT--	-----	-----
<i>Sporobolusfimbriatus</i> 7509	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Sporoboluspyramidalis</i> 6966	-----	TTCTTTTT--	---TATT---	-----	-----
<i>Sporobolusfimbriatus</i> 9328	-----	TACTTTTT--	---TATT---	-----	-----

PHRAGMITESAUSTRALIS6557	-AGAGTATCG GCAAGGA--- -----A	TCTCGATTAT	TAATTAGATT
ARISTIDAADSCENSIONIS6540	-AGAGTATCG GCAAGGA--- -----A	TCTCGATTAT	TAATTAGATT
KARROOCHLOAPURPUREA6241	-AGAGTATCA GAAAGAA--- -----A	TCCCCATTAG	TAATTCGATT
CHAETOBROMUSINVOLUCRATES6284	-AGAGTATCG GCAAGAA--- -----A	TCCCCATTAT	TAATTCGATT
SORGHUMHALEPENSE	TACAGTATCG GCAAGGA--- -----A	TCTCGATTAT	TAATTCGATT
IMPERATACYLINDRICA	TAGAGTATCG GCAAGGA--- -----A	TCTCGATTAT	TAATTCGATT
PANICUMLANIPES	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMSTAPFIANUM	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon7480	-AGAGTATCC GCAAAAA--- -----A	TCTCAATTAT	TAATTA AATT
Chlorisvirgata6616	-AGAGTATCC ATAAAAA--- -----A	TCTCAATTAT	TAATTA AATT
Chlorisvirgata7498	-AGAGTATCC ATAAAAA--- -----A	TCTCAATTAT	TAATTA AATT
Eustachyspaspaloides5597	-AGAGTATCC GCAAAAA--- -----A	TCTCAATTAT	TAATTA AATT
Harpochloafalx6530	-AGAGTATCC GCAAAAA--- -----A	TCTCAATTAT	TAATGA AATT
Harpochloafalx6670	-AGAGTATCC GCAAAAA--- -----A	TCTCAATTAT	TAATGA AATT
Harpochloafalx5118	-AGAGTATCC GCAAAAA--- -----A	TCTCAATTAT	TAATGA AATT
Rendliaaltera5133	-AGAGTATYS GCAARRA--- -----A	TCTCAATTAT	TAATTA AATT
Leptochloafusca7622	-AGAGTATCC GCAAAAA--- -----A	TCTCAATTAT	TATTTA AATT
Enteropogonmacrostachyus9339	-AGAGTATCC GCAAAAATAG AGTATCCGCA	TCTCAATTAT	TAATGA AATT
Dactylocteniumaegyptium9326	-AGAGTATCC ACAA AAAA--- -----A	TCTCAATTAT	TAATTTA AATT
Tragusberteronianus6620	-CGAGTATCC GCAAAGA--- -----A	TCTCAATTAT	TAATTA AATT
Tragusracemosus7108	-CGAGTATCC GCAAAGA--- -----A	TCTCAATTAT	TAATTA AATT
Traguskoelerioides6729	-CGAGTATCC TCAAAGA--- -----A	TCTCAATTAT	TAATTA AATT
Polevansiarigida7621	-TGAGTATCC GCAAAGA--- -----A	TCTCAATTAT	TAATTA AATT
Perotispatens9323	-AGAGTATCC GAAAAAA--- -----A	TCTCAATTAT	TAATGA AATT
Eragrostiscurvula6529	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Eragrostisobtusa6623	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Eragrostisobtusa6732	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Eragrostisechinochloidea7485	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TACTTA AATT
Eragrostispseudobtusa7508	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Eragrostislehmanniana20	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Eragrostischloromelas6947	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Eragrostischloromelas7474	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Eragrostisracemosa6533	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Eragrostisbiflora6705	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Eragrostisuperba7470	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Eragrostiscapensis6551	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Eragrostiscapensis6532	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Eragrostislehmanniana7514	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Pogonarthriasquarrosa7483	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Pogonarthriasquarrosa7504	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Catalepisgracilis6962	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Catalepisgracilis6653	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Cladoraphiscyperoides5704	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Cladoraphisspinosa6335	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Cladoraphiscyperoides4894	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Fingerhuthiaafricana7507	-AGAGTATCG GC--GGA--- -----A	TCTCAATTAT	TAATTA AATT
Entoplocamiaaristulata9353	-AGAGTATCG GC--GGA--- -----A	TCTCAATTAT	TAATTA AATT
Enneapogonscoparius18	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Enneapogoncenchroides7506	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Enneapogonscoparius7486	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Schmidtiaappophoroides6334	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Schmidtiaakaliariensis7490	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Schmidtiaakaliariensis7487	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Sporobolusafricanus7476	-AGAGTATCC GCAAAGA--- -----A	TCTTAATTAT	TAATTTA AATT
Sporobolusconsimilis9354	-AGAGTATCC GCAAAGA--- -----A	TCTCAATTAT	TAATTA AATT
Sporobolusfimbriatus7509	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Sporoboluspyramidalis6966	-AGAGTATCG GCAAGGA--- -----A	TCTCAATTAT	TAATTA AATT
Sporobolusfimbriatus9328	-AGAGTATCC ACAAAGA--- -----A	TTTA AATTAT	TAATTTA AATT

PHRAGMITESAUSTRALIS6557	TT-----	-----TAA	GTATTA----	-----TTA	
ARISTIDAADSCENSIONIS6540	TT-----	-----TAA	GTATTA----	-----TTA	
KARROOCHLOAPURPUREA6241	TT-----	-----TAA	GTATTTTTTT	ATTAATATTA	
CHAETOBROMUSINVOLUCRATES6284	CT-----	-----TAA	TTATTATTTT	ATTAA-----	
SORGHUMHALEPENSE	TTTT-----	-----TAA	GTATTA----	-----TTA	
IMPERATACYLINDRICA	TTTT-----	-----TAA	GTATTA----	-----TTA	
PANICUMLANIPES	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	
PANICUMSTAFFIANUM	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	
Eleusinecoracana	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	
Eleusineindica	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	
Cynodondactylon	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	
Cynodondactylon7480	TA-----	-----TAA	GTATTA----	-----TTA	
Chlorisvirgata6616	TC-----	-----TAA	GTATTA----	-----TTA	
Chlorisvirgata7498	TC-----	-----TAA	GTATTA----	-----TTA	
Eustachyaspaspaloides5597	TA-----	-----TAA	GTATTA----	-----TTA	
Harpochloafalx6530	TA-----	-----TAA	GTATTA----	-----TTA	
Harpochloafalx6670	TA-----	-----TAA	GTATTA----	-----TTA	
Harpochloafalx5118	TT-----	-----TAA	GTATTA----	-----TTA	
Rendliaaltera5133	TATAAGTWTT	AT-----	-----TAA	GTATTA----	-----TTA
Leptochloafusca7622	TA-----	-----TAA	GTATTA----	-----TTA	
Enteropogonmacrostachyus9339	TA-----	-----TAA	GTATTA----	-----TTA	
Dactylocteniumaegyptium9326	TA-----	-----TAA	GTATTA----	-----TTA	
Tragusberteronianus6620	AA-----	-----TAA	GTATTA----	-----TTA	
Tragusracemosus7108	AA-----	-----TAA	GTATTA----	-----TTA	
Traguskoelerioides6729	AA-----	-----TAA	GTATTA----	-----TTA	
Polevansiarigida7621	TATAAGAATT	TATTACAATT	TATAAGATAA	GTATTA----	-----TTA
Perotispatens9323	AA-----	-----TAA	GTATTA----	-----TTA	
Eragrostiscurvula6529	AT-----	-----TAA	GTATTA----	-----TTA	
Eragrostisobtusa6623	AT-----	-----TAA	GTATTA----	-----TTA	
Eragrostisobtusa6732	AT-----	-----TAA	GTATTA----	-----TTA	
Eragrostisechinochloidea7485	AT-----	-----TAA	GTATTA----	-----TTA	
Eragrostispseudobtusa7508	AT-----	-----TAA	GTATTA----	-----TTA	
Eragrostislehmanniana20	AT-----	-----TAA	GTATTA----	-----TTA	
Eragrostischloromelas6947	AT-----	-----TAA	GTATTA----	-----TTA	
Eragrostischloromelas7474	AT-----	-----TAA	TTATTA----	-----TTA	
Eragrostisracemosa6533	AT-----	-----TAA	GTATTA----	-----TTA	
Eragrostisbiflora6705	AT-----	-----TAA	GTATTA----	-----TTA	
Eragrostissuperba7470	AT-----	-----TAA	GTATTA----	-----TTA	
Eragrostiscapensis6551	AT-----	-----TAA	GTATTA----	-----TTA	
Eragrostiscapensis6532	AT-----	-----TAA	GTATTA----	-----TTA	
Eragrostislehmanniana7514	AT-----	-----TAA	GTATTA----	-----TTA	
Pogonarthriasquarrosa7483	AT-----	-----TAA	GTATTA----	-----TTA	
Pogonarthriasquarrosa7504	AT-----	-----TAA	GTATTA----	-----TTA	
Catalepisgracilis6962	AT-----	-----TAA	TTATTA----	-----TTA	
Catalepisgracilis6653	AT-----	-----TAA	TTATTA----	-----TTA	
Cladoraphiscyperoides5704	ATA-----	-----TAA	GTATTAT--	-----TTA	
Cladoraphis spinosa6335	ATA-----	-----TAA	GTATTAT--	-----TTA	
Cladoraphiscyperoides4894	ATA-----	-----TAA	GTATTAT--	-----TTA	
Fingerhuthiaafricana7507	TT-----	-----TAA	GTATTA----	-----TTA	
Entoplocamiaaristulata9353	TT-----	-----TAA	GTATTA----	-----TTA	
Enneapogonscoparius18	TTT-----	-----GAA	GTATTA----	-----TTAATATTA	
Enneapogoncenchroides7506	TTT-----	-----GAA	GTATTA----	-----TTAATATTA	
Enneapogonscoparius7486	TTT-----	-----GAA	GTATTA----	-----TTAATATTA	
Schmidtia pappophoroides6334	TT-----	-----GAA	GTATTA----	-----TTAATATTA	
Schmidtia kalihariensis7490	TT-----	-----GAA	GTATTA----	-----TTAATATTA	
Schmidtia kalihariensis7487	TT-----	-----GAA	GTATTA----	-----TTAATATTA	
Sporobolusafricanus7476	TT-----	-----TAA	GTATTA----	-----TTA	
Sporobolus consimilis9354	TT-----	-----TAA	GTATTA----	-----TTA	
Sporobolus fimbriatus7509	AT-----	-----TAA	GTATTA----	-----TTA	
Sporobolus pyramidalis6966	AT-----	-----TAA	GTATTA----	-----TTA	
Sporobolus fimbriatus9328	TG-----	-----GAA	TTATTA----	-----TTA	

PHRAGMITESAUSTRALIS6557	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
ARISTIDAADSCENSIONIS6540	AGTA-----	-----	-----	AGCC	ATCCATAATG	CATAGGAC--
KARROOCHLOAPURPUREA6241	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
CHAETOBROMUSINVOLUCRATES6284	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
SORGHUMHALEPENSE	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
IMPERATACYLINDRICA	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
PANICUMLANIPES	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMSTAFFIANUM	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon7480	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Chlorisvirgata6616	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Chlorisvirgata7498	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Eustachyspaspaloides5597	AGTA-----	-----	-----	AGCC	GTCCACAATG	CATAGGAC--
Harpochloafalx6530	AGTA-----	-----	-----	AGCC	ATCTACAATG	CATAGGAC--
Harpochloafalx6670	AGTA-----	-----	-----	AGCC	ATCTACAATG	CATAGGAC--
Harpochloafalx5118	AGTA-----	-----	-----	AGCC	ATCTACAATG	CATAGGAC--
Rendliaaltera5133	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Leptochloafusca7622	ACTA-----	-----	-----	AGCC	ATCCACAATG	TATAGGAC--
Enteropogonmacrostachyus9339	AGTA-----	-----	-----	AGCC	ATCCACAATG	TATAGGAC--
Dactylocteniumaegyptium9326	CGTA-----	-----	-----	AGTC	ATCCACAATG	CATAGGAC--
Tragusberteronianus6620	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Tragusracemosus7108	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Traguskoelerioides6729	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Polevansiarigida7621	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Perotispatens9323	AGTA-----	-----	-----	AGCC	GTCCACAATG	CATAGGAC--
Eragrostiscurvula6529	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Eragrostisobtusa6623	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Eragrostisobtusa6732	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATCGGAC--
Eragrostisechinochloidea7485	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Eragrostispseudobtusa7508	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATCGGAC--
Eragrostislehmanniana20	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Eragrostischloromelas6947	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Eragrostischloromelas7474	AATA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Eragrostisracemosa6533	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Eragrostisbiflora6705	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Eragrostissuperba7470	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Eragrostiscapensis6551	AGTA-----	-----	-----	AGCC	GTCCACAATG	CATAGGAC--
Eragrostiscapensis6532	AGTA-----	-----	-----	AGCC	GTCCACAATG	CATAGGAC--
Eragrostislehmanniana7514	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Pogonarthriasquarrosa7483	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Pogonarthriasquarrosa7504	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Catalepisgracilis6962	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Catalepisgracilis6653	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Cladoraphiscyperoides5704	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGTAC--
Cladoraphis spinosa6335	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Cladoraphiscyperoides4894	AGTATT--AT	T-----	TAAGTAAGCC	ATCCACAATG	CATAGGAC--	
Fingerhuthiaafricana7507	AGTA-----	-----	-----	ATCC	ATCCACAATG	CATAGGAC--
Entoplocamia aristulata9353	AGTAATCCAT	CCACAATGCA	TAAGTAATCC	ATCCACAATG	CATAGGAC--	
Enneapogon scoparius18	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Enneapogon cenchroides7506	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Enneapogon scoparius7486	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Schmidtia pappophoroides6334	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Schmidtia kalihariensis7490	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGACCA
Schmidtia kalihariensis7487	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGACCA
Sporobolus africanus7476	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Sporobolus consimilis9354	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAT--
Sporobolus fimbriatus7509	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Sporobolus pyramidalis6966	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--
Sporobolus fimbriatus9328	AGTA-----	-----	-----	AGCC	ATCCACAATG	CATAGGAC--

PHRAGMITESAUSTRALIS6557	-----TACC	CTCTCAC---	ATTCCAAAT	TTGGAATAGA	ATACTTT---
ARISTIDAADSCENSIONIS6540	-----TACC	CCTC-----	ATTCCAAAT	T-----TGGA	ATACTTT---
KARROOCHLOAPURPUREA6241	-----TACT	CCCC-----	ATTCCAAAAT	TAGGAATGGA	AAACTTT---
CHAETOBROMUSINVOLUCRATES6284	-----	-----	-----C	TAGGAATGGA	AAACTTG---
SORGHUMHALEPENSE	-----TACC	CCTCCCC---	ATTTCCTAAT	TTTGAATGGA	ATACTTT---
IMPERATACYLINDRICA	-----TACC	CCTCCCC---	ATTTCCTAAT	TTGGAATGGA	ATACTTT---
PANICUMLANIPES	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMSTAPFIANUM	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon7480	-----TATC	CCCCCCCC--	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Chlorisvirgata6616	-----TACC	CCCCCCCC--	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Chlorisvirgata7498	-----TACC	CCCCCCCC--	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Eustachyspaspaloides5597	-----TATC	CCCCCCC---	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Harpochloafalx6530	-----TACC	CCCCCCC---	ATTTCCAAAT	TAGGAATGGA	ATACTTTATT
Harpochloafalx6670	-----TACC	CCCCCCCC--	ATTTCCAAAT	TAGGAATGGA	ATACTTTATT
Harpochloafalx5118	-----TACC	CCCCCCC---	ATTTCCAAAT	TAGGAATGGA	ATACTTTATT
Rendliaaltera5133	-----TAGC	CCCCCTT---	-TTTCCAAAT	TAGGAATGGA	ATACTTT---
Leptochloafusca7622	-----TATC	CCCCTCCC--	ATTTCTAAA	TAGGAATGGA	ATACTTT---
Enteropogonmacrostachyus9339	-----TACT	CCCCCCC---	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Dactylocteniumaegyptium9326	-----TACC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Tragusberteronianus6620	-----TACC	CCCC-----	GTTTCCAAAT	TAGGAATGGA	ATACTTT---
Tragusracemosus7108	-----TACC	CCCC-----	GTTTCCAAAT	TAGGAATGGA	ATACTTT---
Traguskoelerioides6729	-----TACC	CCCC-----	GTTTCCAAAT	TAGGAATGGA	ATACTTT---
Polevansiarigida7621	-----TACC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Perotispatens9323	-----TATC	CCCCCCCC--	ATTTCCAAAT	TAGGATGGA	ATACTTT---
Eragrostiscurvula6529	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Eragrostisobtusa6623	-----TATC	CCCC-----	ATTTACAAAT	TAGGAATGGA	ATACTTT---
Eragrostisobtusa6732	-----TATC	CCCC-----	ATTTACAAAT	TAGGAATGGA	ATACTTT---
Eragrostisechinochloidea7485	-----TATC	CCCC-----	ATTTACAAAT	TAGGAATGGA	ATACTTT---
Eragrostispseudobtusa7508	-----TATC	CCCC-----	ATTTACAAAT	TAGGAATGGA	ATACTTT---
Eragrostislehmanniana20	-----TACC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Eragrostischloromelas6947	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Eragrostischloromelas7474	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Eragrostisracemosa6533	-----TATC	CCCCG-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Eragrostisbiflora6705	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Eragrostissuperba7470	-----TATC	CCCC-----	ATTTCTAAAT	TAGGAATGGA	ATACTTT---
Eragrostiscapensis6551	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Eragrostiscapensis6532	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Eragrostislehmanniana7514	-----TACC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Pogonarthriasquarrosa7483	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Pogonarthriasquarrosa7504	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Catalepisgracilis6962	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Catalepisgracilis6653	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Cladoraphiscyperoides5704	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTTATT
Cladoraphisspinosa6335	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTTATT
Cladoraphiscyperoides4894	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTTATT
Fingerhuthiaafricana7507	-----TATC	CCCC-----	ATTTCTAAAT	TAGGAATGGA	ATACTTT---
Entoplocamiaaristulata9353	-----TATC	CCCC-----	ATTTCTAAAT	TAGGAATGGA	ATACTTT---
Enneapogonscoparius18	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Enneapogoncnenchroides7506	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATAGA	ATACTTT---
Enneapogonscoparius7486	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Schmidtiaappophoroides6334	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Schmidtiaalihariensis7490	TAGGACTATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Schmidtiaalihariensis7487	TAGGACTATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Sporobolusafricanus7476	-----TATC	CCCCCCCC-T	AATTCAAAAT	TAGGAATGGA	ATACTTT---
Sporobolusconsimilis9354	-----TATC	C-----	-----AAAT	TAGGAATGGA	ATACTTT---
Sporobolusfimbriatus7509	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Sporoboluspyramidalis6966	-----TATC	CCCC-----	ATTTCCAAAT	TAGGAATGGA	ATACTTT---
Sporobolusfimbriatus9328	-----TACC	CCCCCCCC--	ATTTCCAAAT	TAGGAATGGA	ATACTTT---

PHRAGMITESAUSTRALIS6557	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
ARISTIDAADSCENSIONIS6540	-----	---ATT-TTT	TTTTACTCCC	TTTAATT---	----GACATA
KARROOCHLOAPURPUREA6241	-----	---AACTGATT	TTTTAATCCC	TTTAATTGAC	AATTGACATA
CHAETOBROMUSINVOLUCRATES6284	-----	---AATTGATT	TTTTAATCCC	TTTAATT---	----GACATA
SORGHUMHALEPENSE	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
IMPERATACYLINDRICA	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
PANICUMLANIPES	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMSTAPFIANUM	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon7480	-----	---ATTGATT	TTTTAGTCTC	TTTAATT---	----GACATA
Chlorisvirgata6616	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Chlorisvirgata7498	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eustachyaspaloides5597	-----	---ATTGATT	TTTTAGTCTC	TTTAATT---	----GACATA
Harpochloafalx6530	GATTGAATAC	TTTATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Harpochloafalx6670	GATTGAATAC	TTTATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Harpochloafalx5118	GATTGAATAC	TTTATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Rendliaaltera5133	-----	---ATTAATT	TTTTAGTCTC	TTTAATT---	----GACATA
Leptochloafusca7622	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Enteropogonmacrostachyus9339	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Dactylocteniumaegyptium9326	-----	---TTTGATT	TTTTAGTCCC	ATTAATT---	----GACATA
Tragusberteronianus6620	-----	---ATTGATT	TTT-AGTCTC	TTTAATT---	----GACATA
Tragusracemosus7108	-----	---ATTGATT	TTT-AGTCTC	TTTAATT---	----GACATA
Traguskoelerioides6729	-----	---ATTGATT	TTT-AGTCTC	TTTAATT---	----GACATA
Polevansiarigida7621	-----	---ATTGATT	TTT-AGTCTC	TTTAATT---	----GACATA
Perotispatens9323	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eragrostiscurvula6529	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eragrostisobtusa6623	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eragrostisobtusa6732	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eragrostisechinochloidea7485	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eragrostispseudobtusa7508	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eragrostislehmanniana20	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eragrostischloromelas6947	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eragrostischloromelas7474	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eragrostisracemosa6533	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eragrostisbiflora6705	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eragrostissuperba7470	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eragrostiscapensis6551	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eragrostiscapensis6532	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Eragrostislehmanniana7514	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Pogonarthriasquarrosa7483	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Pogonarthriasquarrosa7504	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Catalepisgracilis6962	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Catalepisgracilis6653	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Cladoraphiscyperoides5704	GATT-----	---TGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Cladoraphisspinosa6335	GATT-----	---TGATT	TTTT-----C	TTTAATT---	----GACATA
Cladoraphiscyperoides4894	GATT-----	---TGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Fingerhuthiaafricana7507	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Entoplocamiaaristulata9353	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Enneapogonscoparius18	-----	---ATTGATC	TTTTAGTCCC	TTTAATT---	----GACATA
Enneapogoncenchroides7506	-----	---ATTGAT-	TTTTAGTCCC	TTTAATT---	----GACATA
Enneapogonscoparius7486	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Schmidtiaappophoroides6334	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Schmidtiaalihariensis7490	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Schmidtiaalihariensis7487	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Sporobolusafricanus7476	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Sporobolusconsimilis9354	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Sporobolusfimbriatus7509	-----	---ATTGATT	TTTTGGTCCC	TTTAATT---	----GACATA
Sporoboluspyramidalis6966	-----	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA
Sporobolusfimbriatus9328	-----	---ATTTATT	TTTTAGTCCC	TTTAATT---	----GACATA





**APPENDIX H.** Indels coded by GapCoder for *trnL-F* data set. Where missing data was coded in the sequence data set (N), the corresponding indel was coded as ?. The indel characters are coded with 1 for present, 0 for missing and - for inapplicable. As the gap matrix was added directly to the sequencing matrix the numbering continues from the last character in the sequence data set.

	1192	1251
<i>PHRAGMITESAUSTRALIS6557</i>	?01000010000000000100101-0010-00001-0100101----	010000001---
<i>ARISTIDAADSCENSIONIS6540</i>	??0001-0100101----	000000001---
<i>KARROOCHLOAPURPUREA6241</i>	1000000101000000000000001-01-0-00001-0100101----	010100001---
<i>CHAETOBROMUSINVOLUCRATES6284</i>	1000000000010000000000001-0010-00001-0100101----	010100001---
<i>SORGHUMHALEPENSE</i>	????????????????000000000100-1-00001-0-1-----	0-----001---
<i>IMPERATACYLINDRICA</i>	????????????????00100000010010-10001-0-1-----	0-----001---
<i>PANICUMLANIPES</i>	0000000000000000000000001-0010-00001-1000101----	010000001---
<i>PANICUMSTAFFIANUM</i>	0000000000000000000000001-0010-00001-1000101----	010000001---
<i>Eleusinecoracana</i>	100000000000000000000000010-00001-00001001001010000001---	
<i>Eleusineindica</i>	100000000000000000000000010-00001-00001001001010000001---	
<i>Cynodondactylon</i>	10000000000000000000000001-01-0-00001-00001001001010000001---	
<i>Cynodondactylon7480</i>	?????010000000010000001-01-0-00001-00001001001010000001---	
<i>Chlorisvirgata6616</i>	1010000100010000000000001-01-0-00001-0000100-1--	010000001---
<i>Chlorisvirgata7498</i>	?????????010000000000001-01-0-00001-00001001001010000001---	
<i>Eustachyspaspaloides5597</i>	??0010010000010000001---	
<i>Harpochloafalx6530</i>	10000000001-0000000000001-01-0-00001-000010000001010000001---	
<i>Harpochloafalx6670</i>	100000000010000000000001-01-0-00001-00001000001010000001---	
<i>Harpochloafalx5118</i>	????????????????????00001-1-----00111-00001000001010000001---	
<i>Rendliaaltera5133</i>	????????????0000000000001-01-0-00001-00001001001010000001---	
<i>Leptochloafusca7622</i>	????????00100000000001--01-0-00001-00001001001010000001---	
<i>Enteropogonmacrostachyus9339</i>	????????001000000000001-01-0-00001-00001001001010000001---	
<i>Dactylocteniumaegyptium9326</i>	??00001000011010000001---	
<i>Tragusberteronianus6620</i>	????????????????000000000101-0-00001-00001001001010000001---	
<i>Tragusracemosus7108</i>	????????????????????000000101-0-00001-00001001001010000001---	
<i>Traguskoelerioides6729</i>	????????????????????0000000101-0-00001-0000100000010100000001---	
<i>Polevansiarigida7621</i>	????????000000000000000101-0-00001-00001001001010000001---	
<i>Perotispatens9323</i>	??01-0000100100101000000100	
<i>Eragrostiscurvula6529</i>	10000000000-1-00010000001-0010-00001-0000101----	010001001---
<i>Eragrostisobtusa6623</i>	100010000000000000000001-0010-00001-0000101----	010000001---
<i>Eragrostisobtusa6732</i>	??	
<i>Eragrostisechinochloidea7485</i>	1000100000000100001000001-0010-00001-000011-----	010000001---
<i>Eragrostispseudobtusa7508</i>	10000010000-1-0000000001-0010-00001-0000101----	010000001---
<i>Eragrostislehmanniana20</i>	?000000010000000000000001-0010-00001-0000101----	010000001---
<i>Eragrostischloromelas6947</i>	1000000000000000000000001-0010-00001-000011-----	010000001---
<i>Eragrostischloromelas7474</i>	1001000000000000000000001-0010-00001-0000101----	010000001---
<i>Eragrostisracemosa6533</i>	1000000000000000000000001-0010-00001-0000101----	1-----01---
<i>Eragrostisbiflora6705</i>	?????????000000000000001-0010-00001-0000101----	010000001---
<i>Eragrostisuperba7470</i>	10000000000-1-00000000001-0010-00001-0000101----	010010001---
<i>Eragrostiscapensis6551</i>	100100010001000010000101--0010-0100010001101---	010000001---
<i>Eragrostiscapensis6532</i>	??1001--0010-000000000101---	010000001---
<i>Eragrostislehmanniana7514</i>	1000000000000000000000001-0010-00001-0000101----	010000001---
<i>Pogonarthriasquarrosa7483</i>	????????????000100000001--0010-00001-0000101----	010000001---
<i>Pogonarthriasquarrosa7504</i>	100000000000000000000001--0010-00001-0000101----	010000001---
<i>Catalepisgracilis6962</i>	100000000000010010000001-0010-00001-0000101----	010000001---
<i>Catalepisgracilis6653</i>	????????00000010010000001-0010-00001-0000101----	010000001---
<i>Cladoraphiscyperoides5704</i>	??0001-0000101----	010000001---
<i>Cladoraphis spinosa6335</i>	????????0000000000000001-000000001-0000101----	010000001---
<i>Cladoraphiscyperoides4894</i>	??0001-0000101----	010000001---
<i>Fingerhuthiaafricana7507</i>	1001000100000100000001001-01-0-00001-0000001----	010000001---
<i>Entoplocamia aristulata9353</i>	????????????????????0000001-01-0-00001-0000101----	011000011---
<i>Enneapogon scoparius18</i>	????????????000000000001-0010-00001-0000101----	010000000001
<i>Enneapogon cenchroides7506</i>	????????????????0000000001-0010-00001-0000101----	010000000001
<i>Enneapogon scoparius7486</i>	10000000000010000000001-0010-00001-0000101----	010000000001
<i>Schmidtia pappophoroides6334</i>	????????????????0000000001-0010-00001-0000101----	010000100001
<i>Schmidtia kalihariensis7490</i>	?000-1-0000-1-0000000001-0000100001-0000101----	010000000000
<i>Schmidtia kalihariensis7487</i>	1000000000000000000000001-0000100001-0000101----	010000000000
<i>Sporobolus africanus7476</i>	-1100001000000000000000001-0010-00001-00001001001010000001---	
<i>Sporobolus consimilis9354</i>	1001000100000000000000001-0010-00001-00001001001010000001---	
<i>Sporobolus fimbriatus7509</i>	1010000000000000000000001-0010-00001-0000101----	010000001---
<i>Sporobolus pyramidalis6966</i>	????????????????????000001-0010-00001-0000101----	010000001---
<i>Sporobolus fimbriatus9328</i>	??0001-00001001001010000001---	

PHRAGMITESAUSTRALIS6557	000001010101100001-010100000100000010001--110001-1--011---1
ARISTIDAADSCENSIONIS6540	1000--011---110001-0101000001001010000001--1111-1-1--011---1
KARROOCHLOAPURPUREA6241	0100--000101100101-01110000010010-1--0001--1111-1-1--011---0
CHAETOBROMUSINVOLUCRATES6284	0-1---010101100101-01010000010010001-0001--1111-1-1--011---0
SORGHUMHALEPENSE	0-1---010101100001-010100000101-100010001--111011-00001001-1
IMPERATACYLINDRICA	0-1---010101100001-0101000001010000010001--111011-01101001-1
PANICUMLANIPES	0-01--010000100011-010100????????????????????????????????????
PANICUMSTAPFIANUM	0-01--010100100011-010100????????????????????????????????????
Eleusinecoracana	0-1---010101100001-010100????????????????????????????????????
Eleusineindica	0-1---010101100001-010100????????????????????????????????????
Cynodondactylon	0-1---010101100001-010100????????????????????????????????????
Cynodondactylon7480	0-1---010101100001-0101000001000000010100000111-1-1--011---1
Chlorisvirgata6616	0-1---01011-000001-1-01000001000000010010--1111-1-1--011---1
Chlorisvirgata7498	0-1---01011-000001-1-01000001000000010010--1111-1-1--011---1
Eustachyspaspaloides5597	0-1---01010110000????????0001000000010000001111-1-1--011---1
Harpochloafalx6530	0-1---010101100001-0101000001000000010010--1111-1-1--011---1
Harpochloafalx6670	0-1---010101100001-0101000001000000010010--1111-1-1--011---1
Harpochloafalx5118	0-1---010101100001-0101100001000000010010--1111-1-1--011---1
Rendliaaltera5133	0-1---010100100001-0101000001000000010000001111-1-1--0100011
Leptochloafusca7622	0-1---010101100001-0101000000000000010010--1111-1-1--011---1
Enteropogonmacrostachyus9339	0-1---010101100001-0101000001000000010010--1111-1-1--001---1
Dactylocteniumaegyptium9326	0-1---01010110000010101000001000000010010--1111-1-1--011---1
Tragusberteronianus6620	0-1---010101100001-010100101-1-0000010010--1111-1-1--011---1
Tragusracemosus7108	0-1---010101100001-010100101-1-0000010010--1111-1-1--011---1
Traguskoelerioides6729	0-1---010101100001-010100101-1-0000010010--1111-1-1--011---1
Polevansiarigida7621	0-1---010001100001-01010011-----0010--1111-1-1--0100001
Perotispates9323	0-1---010101100001-0101000000000000010010--1111-1-1--011---1
Eragrostiscurvula6529	0-1---010001100001-01010000010000000100001-1111-1-1--011---1
Eragrostisobtusa6623	0-1---010001100001-01010000010000000000001-1011-1-1--011---1
Eragrostisobtusa6732	????????????????001-01010000010000000000001-1011-1-1--011---1
Eragrostisechinocloidea7485	0-1---010001100001-01010000010000000000001-1011-1-1--011---1
Eragrostispseudobtusa7508	0-1---010001100001-01010000010000000000001-1011-1-1--011---1
Eragrostislehmanniana20	0-1---010001100001-01010000010000000100001-1111-011--011---1
Eragrostischloromelas6947	0-1---010001100001-01010000010000000100001-1111-1-1--011---1
Eragrostischloromelas7474	0-1---010001100001-01010100010000000100001-1111-1-1--011---1
Eragrostisracemosa6533	0-1---010001100001-01010000010000000100001-1111-1-1--011---1
Eragrostisbiflora6705	0-1---010001100001-01010000010000000100001-1111-1-1--011---1
Eragrostissuperba7470	0-1---01000110000001010000010000000100001-1111-1-1--011---1
Eragrostiscapensis6551	0-1---010000100001-01010000010000000100001-1111-1-1--011---1
Eragrostiscapensis6532	0-1---010000100001-01010000010000000100001-1111-1-1--011---1
Eragrostislehmanniana7514	0-1---010001100001-01010000010000000100001-1111-011--011---1
Pogonarthriasquarrosa7483	0-1---010001100001-01010000010000000100001-1111-1-1--011---1
Pogonarthriasquarrosa7504	0-1---010001100001-01010000010000000100001-1111-1-1--011---1
Catalepisgracilis6962	0-1---010001100001-01010000010000000100001-1111-1-1--011---1
Catalepisgracilis6653	0-1---010001100001-01010000010000000100001-1111-1-1--011---1
Cladoraphiscyperoides5704	0-1---010001100001-01010000010000000100001-1111-1-1--0101--0
Cladoraphiscyperoides6335	0-1---010001100001-01010000010000000100001-1111-1-1--0101--0
Cladoraphiscyperoides4894	0-1---010001100001-01010000010000000100001-1111-1-1--0101--0
Fingerhuthiaafricana7507	0-1---010001100001-01010000010000000100001-1111-1-1--111---1
Entoplocamiaaristulata9353	0-1---010001100001-01010000010000000100001-1111-1-1--111---1
Enneapogonscoparius18	00001-110001100001-00000000010000000100001-1111-1-1--0101--0
Enneapogoncnchroides7506	000000010001100001-00000000010000000100001-1111-1-1--011---0
Enneapogonscoparius7486	00001-110001100001-00000000010000000100001-1111-1-1--0101--0
Schmidtiaappophoroides6334	0-1---010001100001-01000000010000000100001-1111-1-1--011---0
Schmidtiaalihariensis7490	0-1---010001100001-01010000010000000100001-1111-1-1--011---0
Schmidtiaalihariensis7487	0-1---010001100001-01010000010000000100001-1111-1-1--011---0
Sporobolusafricanus7476	0-1---0101011000000101000000000000011-----11-001--011---1
Sporobolusconsimilis9354	0-1---010101101-----01000001000000010000011111-001--011---1
Sporobolusfimbriatus7509	0-1---010001100001-01010000010000000100001-1111-1-1--011---1
Sporoboluspyramidalis6966	0-1---010001100001-01010????????000100001-1111-1-1--011---1
Sporobolusfimbriatus9328	0-1---01011-1????????????00001000000010010--1111-1-1--011---1

PHRAGMITESAUSTRALIS6557	--01--1000010---01-00000101
ARISTIDAADSCENSIONIS6540	--01--101---0---11-01000101
KARROOCHLOAPURPUREA6241	0001--101---0---00100000001
CHAETOBROMUSINVOLUCRATES6284	001-----00100000101
SORGHUMHALEPENSE	--01--1000010---01-0000010?
IMPERATACYLINDRICA	--01--1000010---01-0000010?
PANICUMLANIPES	????????????????????????????
PANICUMSTAFFIANUM	????????????????????????????
Eleusinecoracana	????????????????????????????
Eleusineindica	????????????????????????????
Cynodondactylon	????????????????????????????
Cynodondactylon7480	--01--10000001--01-00000101
Chlorisvirgata6616	--01--100000000101-00000101
Chlorisvirgata7498	--01--100000000001-00000101
Eustachyspaspaloides5597	--01--1000010---01-00000101
Harpochoalofalx6530	--01--1000010---00000000100
Harpochoalofalx6670	--01--10000001--00000000100
Harpochoalofalx5118	--01--1000010---00000000100
Rendliaaltera5133	--01--10000-1---01-00000101
Leptochloafusca7622	--01--10000001--01-00000101
Enteropogonmacrostachyus9339	--01--1000010---01-00000101
Dactylocteniumaegyptium9326	--01--1001--0---01-00000111
Tragusberteronianus6620	--01--1001--0---01-00010101
Tragusracemosus7108	--01--1001--0---01-00010101
Traguskoelerioides6729	--01--1001--0---01-00010101
Polevansiarigida7621	--01--10001-0---01-00010101
Perotispatens9323	--01--100000000101-00000101
Eragrostiscurvula6529	--01--1001--0---01-00000101
Eragrostisobtusa6623	--01--1001--0---01-00000101
Eragrostisobtusa6732	--01--1001--0---01-00000101
Eragrostisechinochloidea7485	--01--1001--0---01-00000101
Eragrostis pseudobtusa7508	--01--1001--0---01-00000101
Eragrostislehmanniana20	--01--1001--0---01-00000101
Eragrostischloromelas6947	--01--1001--0---01-00000101
Eragrostischloromelas7474	--01--101--0---01-00000101
Eragrostisracemosa6533	--01--1001--0---01-00000101
Eragrostisbiflora6705	--01--1001--0---01-00000101
Eragrostis superba7470	--01--1001--0---01-00000101
Eragrostiscapensis6551	--01--1001--0---01-00000101
Eragrostiscapensis6532	--01--1001--0---01-00000101
Eragrostislehmanniana7514	--01--1001--0---01-00000101
Pogonarthriasquarrosa7483	--01--1001--0---01-00000101
Pogonarthriasquarrosa7504	--01--1001--0---01-00000101
Catalepisgracilis6962	--01--1001--0---01-00000101
Catalepisgracilis6653	--01--1001--0---01-00000101
Cladoraphiscyperoides5704	0101--1001--0---00010000101
Cladoraphis spinosa6335	0101--1001--0---00010001101
Cladoraphiscyperoides4894	0100111001--0---00010000101
Fingerhuthiaafricana7507	--01--1001--0---01-00000101
Entoplocamiaaristulata9353	--00001001--0---01-00000101
Enneapogon scoparius18	1001--1001--0---01-00000111
Enneapogon cenchroides7506	1001--1001--0---01-00100101
Enneapogon scoparius7486	1001--1001--0---01-00000101
Schmidtia pappophoroides6334	1001--1001--0---01-00000101
Schmidtia kalihariensis7490	1001--0001--0---01-00000101
Schmidtia kalihariensis7487	1001--0001--0---01-00000101
Sporobolusafricanus7476	--01--100000001001-00000101
Sporobolus consimilis9354	--01--11-----01-00000101
Sporobolus fimbriatus7509	--01--1001--0---01-00000101
Sporobolus pyramidalis6966	--01--1001--0---01-00000101
Sporobolus fimbriatus9328	--01--1000010---01-00000101

**APPENDIX I.** Table of correspondence between the indels and the regions they represent (Appendix H) generated by GapCoder for the *trnL-F* data set.

Indel Character	Sequence Region	Indel Character	Sequence Region	Indel Character	Sequence Region
1192	4-4	1241	278-283	1290	734-734
1193	4-5	1242	286-286	1291	737-741
1194	10-10	1243	290-290	1292	738-742
1195	14-14	1244	293-296	1293	738-741
1196	15-15	1245	297-297	1294	740-740
1197	15-16	1246	326-330	1295	757-760
1198	16-16	1247	348-348	1296	774-778
1199	17-17	1248	360-365	1297	832-832
1200	19-19	1249	360-360	1298	838-860
1201	22-22	1250	362-365	1299	838-857
1202	26-27	1251	364-365	1300	869-873
1203	27-27	1252	375-381	1301	870-873
1204	27-28	1253	380-382	1302	878-901
1205	28-28	1254	380-384	1303	889-890
1206	42-42	1255	380-383	1304	898-898
1207	52-52	1256	380-381	1305	913-914
1208	59-59	1257	381-381	1306	918-929
1209	60-60	1258	392-392	1307	953-977
1210	63-63	1259	432-432	1308	954-977
1211	70-70	1260	437-441	1309	955-977
1212	82-82	1261	438-438	1310	963-977
1213	85-85	1262	440-441	1311	987-997
1214	98-98	1263	441-441	1312	987-991
1215	104-109	1264	450-455	1313	988-997
1216	105-109	1265	461-465	1314	996-1079
1217	106-109	1266	478-529	1315	1005-1026
1218	115-123	1267	506-506	1316	1007-1008
1219	116-119	1268	511-511	1317	1012-1020
1220	117-119	1269	517-521	1318	1049-1056
1221	117-122	1270	518-521	1319	1062-1076
1222	118-119	1271	526-527	1320	1065-1070
1223	126-130	1272	527-527	1321	1066-1070
1224	134-134	1273	532-532	1322	1067-1070
1225	135-135	1274	538-542	1323	1068-1070
1226	144-146	1275	551-551	1324	1068-1071
1227	155-160	1276	607-607	1325	1069-1070
1228	155-155	1277	691-691	1326	1069-1069
1229	203-204	1278	693-730	1327	1070-1070
1230	205-206	1279	694-695	1328	1082-1086
1231	205-324	1280	701-705	1329	1098-1113
1232	211-211	1281	701-714	1330	1098-1112
1233	232-236	1282	709-709	1331	1105-1115
1234	244-265	1283	716-716	1332	1117-1117
1235	245-263	1284	716-718	1333	1120-1120
1236	245-249	1285	720-720	1334	1124-1124
1237	245-261	1286	720-727	1335	1125-1129
1238	252-253	1287	721-727	1336	1138-1144
1239	256-261	1288	723-727	1337	1170-1170
1240	272-338	1289	731-815	1338	1181-1184

**APPENDIX J.** Alignment of combined *ITS+trnL-F* sequencing data. Numbers indicate the consecutive positions of 1 to 1831 (5' to 3') from the beginning of the *ITS1* region to the end of the *trnL-F* region. Dashes denote gaps. Arrows indicate the beginning of the *ITS* and *trn* regions. N indicates large regions of missing data where sequencing reactions failed.

	↓	1		50
<i>IMPERATACYLINDRICA</i>		NNNNNNNNNN	NNNNNNNNNN	NNNCCGCGAA CGA-GTCTCT CGTGCCGCCG
<i>SORGHUMHALEPENSE</i>		TCGTGACC-T	TAAACAAAAC	AGACCGTGAA CAT-GTCTCT CATGTCGTCG
<i>PANICUMLANIPES</i>		TCGTGACCCT	TAAACAAAAC	AGACCGTGAA CGT-GTCATC CATGTCGTCCT
<i>PANICUMSTAPFIANUM</i>		TCGTGACCCT	TAAACAAAAC	AGACCGTGAA CAT-GTCATC CATGTCGTCG
<i>KAROOCHLOAPURPUREA6241</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>CHAETOBROMUSINVOLUCRATES6284</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>Cynodondactylon7480</i>		TCGTGACCCT	GACC-AAAAC	AAACCGTGAA CCT-GTCACC TATGCTGCCG
<i>Cynodondactylon</i>		TCGTGACCCT	GATG-AAAAC	AACCCGTGAA TTT-GTCACC TATGCTGCCG
<i>Eleusinecoracana</i>		TCGTGACCCT	AAACCAAAAAC	AAACCGTGAA CAT-GTCATC CATGCTGCCG
<i>Eleusineindica</i>		TCGTGACCCT	AAACCAAAAAC	AAACCGTGAA CAT-GTCATC CATGCTGCCG
<i>Chlorisvirgata6616</i>		TCGTGACCCT	GACC-AAAAC	AGACCGTGAA CAT-GTCATC TATGCTGCCG
<i>Chlorisvirgata7498</i>		TCGTGACCCT	GACC-AAAAC	AGACCGTGAA CAT-GTCATC TATGCTGCCG
<i>Eustachyspaspaloides5597</i>		TCGTTACCCT	GACCCAAAAT	AGACCGTGAA CATTGTGATC TAAGCTGCCG
<i>Harpochloafalx6530</i>		TCGTGACCCT	GACC-AAAAG	AGACTGTGAA CAT-GTCATC CATGCTGCTG
<i>Harpochloafalx5118</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>Harpochloafalx6670</i>		TAGTGATCCT	GACC-AAAAG	AGACTGTGAA CAT-GTCATC CATGCTGGTG
<i>Rendliaaltera5133</i>		TCGTGACACT	GACC-AAAAT	AGACTGTGAA CAT-GTCATC CATGCTGCCG
<i>Leptochloafusca7622</i>		TCGTGACCCT	GACC-AAAAC	AGACCACGAA CAT-GTCATC CATGCTGCCG
<i>Enteropogonmacrostachyus9339</i>		TCGTGACCCT	GACC-AAAAC	AGACCGTGAA CAT-GTCATC CA-GCGGCCG
<i>Dactylocteniumaegyptium9326</i>		TCGTGACCCT	GACC-AAAAY	AGACCGTGAA CGC-GTCATC CATGGTGCCG
<i>Tragusberteronianus6620</i>		TCGTGACCCT	GACC-AAAAC	AGACCGTGAA CGT-GTCATC CATGCTCCCG
<i>Tragusracemosus7108</i>		TCGTGACCCT	GACC-AAAAC	AGACCGTGAA CGT-GTCATC CATGCTCCCG
<i>Traguskoelerioides6729</i>		TCKTGACCCT	GACC-AAAAC	AGACCGTGAA CGT-GTCATC CATGCTCCCG
<i>Polevansiarigida7621</i>		TCGTGACCCT	GACC-AAAAC	ACACCGTGAA CAT-GTCATC CATGCTCCCG
<i>Perotispatens9323</i>		TCGTGACCCT	GACC-AAAAG	AGATCGTGAA CTT-GTCATC TATCTTACTA
<i>Eragrostiscurvula6529</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>Eragrostisobtusa6623</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>Eragrostisobtusa6732</i>		TCGTGACCCT	GACC-AAAAC	AGACCGTGAA CGT-GTCATC CATCCTGCCG
<i>Eragrostisechinochloidea7485</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>Eragrostispseudohloidea7508</i>		TCGTGACCCT	GACC-AAAAC	AGACCGTGAA CAT-GTCATC CATCTTGCCG
<i>Eragrostislehmanniana20</i>		TCGTGACCCT	GACC-AAAAC	AGACCGTGAA CGT-GTCATC CATCTTGCCG
<i>Eragrostischloromelas6947</i>		TCGTGACCCT	GACC-AAAAC	AGACCGTGAA CGT-GTCATC CATCCTGCCG
<i>Eragrostischloromelas7474</i>		TCGTGACCCT	TACC-AAAAC	AGACCGCGAR CAT-GTCATC CATGCTGCCG
<i>Eragrostisracemosa6533</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>Eragrostisbiflora6705</i>		TCGTGACCCT	TAAC-AAAAC	AGACCGCGAA CAT-GTCATC CATGCTGCCG
<i>Eragrostissuperba7470</i>		TCGTGACCCT	AACC-AAAAC	AGACCGCGAA CGT-GTCATC CATGCTGCCG
<i>Eragrostiscapensis6551</i>		TCGTGACCCT	GACC-AAAAC	AGACCGTGAA CGT-GTCATC CATGCCGCCG
<i>Eragrostiscapensis6532</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>Pogonarthriasquarrosa7483</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>Pogonarthriasquarrosa7504</i>		TCGTGACCCT	GAAC-AAAAC	AGACCGTGAA CGT-GTCATC CATCTTGCCG
<i>Catalepisgracilis6962</i>		TCGTGACCCT	TAAC-AAAAC	AGACCGCGAA CAT-GTCATC CATGCTGCCG
<i>Catalepisgracilis6653</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>Cladoraphiscyperoides5704</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>Cladoraphisspinosa6335</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>Cladoraphiscyperoides4894</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>Fingerhuthiaafricana7507</i>		TCGTGACCCT	TACG-AAAAC	AGACCGTGAA CAT-GTCACC CATGCCGTCG
<i>Entoplocamiaaristulata9353</i>		TCGTGACCCT	TACC-AAAAC	AGACCGTGAA CAT-GTCATC CATGCCGTCG
<i>Enneapogonscoparius18</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>Enneapogoncenchroides7506</i>		TCGTGATCCT	GACC-AAAAC	AGACCGCGAA CGT-GTTATC TAAGCCGCCG
<i>Enneapogonscoparius7487</i>		TCGTGACCCT	GACC-AAAAC	AGACCGTGAA CGT-GTTATC CAAGCCGCCG
<i>Schmidtiaappophoroides6334</i>		TCGTGACCCT	TAAG-AAAAC	AGACCCCGAA CGA-GTCATC CATCCCGCCG
<i>Schmidtiaakalihariensis7490</i>		TCGTGACCCT	GACC-AAAAC	AGACCGCGAA CGC-GTCATC CATGCCGCCG
<i>Schmidtiaakalihariensis7486</i>		NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
<i>Sporobolusafricanus7476</i>		TCGTGACCCT	AAAC-AAAAC	AGACTGTGAA TAT-GTCTTC CATGTGTCA
<i>Sporobolusconsimilis9354</i>		TCGTGACCCT	AACC-AAAAG	AGACTGTGAA TAT-GTCATC CATCTGTGCG
<i>Sporobolusfimbriatus9328</i>		TCGTGACCCT	AACC-AAAAT	AGACTGTGAA CAT-GTCATC AATTCACCG

IMPERATACYLINDRICA  
 SORGHUMHALEPENSE  
 PANICUMLANIPES  
 PANICUMSTAFFIANUM  
 KARROOCHLOAPURPUREA6241  
 CHAETOBROMUSINVOLUCRATES6284  
 Cynodondactylon7480  
 Cynodondactylon  
 Eleusinecoracana  
 Eleusineindica  
 Chlorisvirgata6616  
 Chlorisvirgata7498  
 Eustachyspaspaloides5597  
 Harpochloafalx6530  
 Harpochloafalx5118  
 Harpochloafalx6670  
 Rendliaaltera5133  
 Leptochloafusca7622  
 Enteropogonmacrostachyus9339  
 Dactylocteniumaegyptium9326  
 Tragusberteronianus6620  
 Tragusracemosus7108  
 Traguskoelerioides6729  
 Polevansiarigida7621  
 Perotispatens9323  
 Eragrostiscurvula6529  
 Eragrostisobtusa6623  
 Eragrostisobtusa6732  
 Eragrostisechinochloidea7485  
 Eragrostispseudobtusa7508  
 Eragrostislehmanniana20  
 Eragrostischloromelas6947  
 Eragrostischloromelas7474  
 Eragrostisracemosa6533  
 Eragrostisbiflora6705  
 Eragrostissuperba7470  
 Eragrostiscapensis6551  
 Eragrostiscapensis6532  
 Pogonarthriasquarrosa7483  
 Pogonarthriasquarrosa7504  
 Catalepisgracilis6962  
 Catalepisgracilis6653  
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 Cladoraphiscyperoides4894  
 Fingerhuthiaafricana7507  
 Entoplocamiaaristulata9353  
 Enneapogonscoparius18  
 Enneapogoncenchroides7506  
 Enneapogonscoparius7487  
 Schmidtiaappophoroides6334  
 Schmidtiakalihariensis7490  
 Schmidtiakalihariensis7486  
 Sporobolusafricanus7476  
 Sporobolusconsimilis9354  
 Sporobolusfimbriatus9328

51  
 GGC----- TCCGGCCCG CCAAGGCCCC CGAGCTCCGT  
 AGC----- TTTGGCTCG CCAAGGTCCC CTTGCTCCAA  
 GGC----- TACGGCCCG CCAAGGCCCC CAACCTTCGT  
 GGC----- TACGTTCCGA CCAAGGCCCC CCTTCACATT  
 NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN CGACCTCCGT  
 NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN  
 GTTGATGGG- CTT--GCACC TATCTCTCG TCTAGGCCAC CGACCTTCTT  
 GTTGATGGAG CTT--GCACC TATCTCTCG TCTAGGCCAC CGACCTTCTT  
 GGCGATGGGT CTT--GCACC CATCTCTGG AACAGGGCCG CCACCTTCTT  
 GGCGATGGGT CTT--GCACC CATCTCTGG AACAGGGCCG CCACCTTCTT  
 TTWGATGGGT CTT--GCACC TATCTCTCG TTTAGGCCCG CAACCTTCTT  
 TTTGATGGGT CTT--GCACC TATCTCTCG TTTAGGCCCG CAACCTTCTT  
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 GTTGATGGG- CTT--GCACC TATCTCCCG TCTTGGCCAC TGACCTTCTT  
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 GTTGATGGGG CTT--GCACC TATCTCTGG TCTAGGCCAC CAACCTTCTT  
 GGTGATGGGG CTT--GCACC CGTCTCCCG TCTTGGCCAC CGACCTTCTT  
 GG-GATGGGG CTT--GCACC TATCTGTCA CGGTATCCCC CGATCCTCTT  
 GTTGACGGGG CTC--GCACC CGTCTCTCG TAGGGCAGG- CGACCTTCTT  
 AGTGATGGGT CTT--GCACC CGTCTCTGG CCCTGGGCTC CGAACTTCGA  
 AGTGATGGGT CTT--GCACC CGTCTCTGG CCCTGGGCTC CGAACTTCGA  
 AGTGACGGGT CTT--GCACC CGTCTCTGG CCCTGGGCTC CGAACTTCGA  
 AGNGATGGGS CTT--GCACC CATCTCTCG TCCATGGCTC C-AACTTCTC  
 GGTGATGGGG -TA--GCACC TGTCTCTAG TATTGGTGAC TTACCTTCTT  
 NNNNNNNNNN NNNNNNNNTC AA-CTCCCG CTTAG-CCCC AGA-CTTCCG  
 NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNGCT  
 GGTGACGGGG CTTTTCCTC GC-CTCCCG CTTAGGCCCC AGA-CTTCCG  
 NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN  
 GGTGACGGGG CTTTTCCTC GC-CTCCCG CTTAGGCCCC AGA-CTTCCG  
 GGWGACGGGG CTT--GCCTC GG-CTCCCG CTTAGGCCCC AGA-CTTCCA  
 GGTGACGGGG CTT--GCCTC GR-CTCCCG CTTAGGCCCC AGA-CTTCCG  
 GGTGACGGGG CTT--GCCTC GC-CTCCCG CTTAGGTCCC AGA-CTTCCG  
 NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN  
 Eragrostisracemosa6533 NNNNNNNNNN NNNNNNNNNN A-A-CTTTCG  
 GGTGACGGGG CTT--GCCTC GC-CTCCCG CTTAGGTCCC AGA-CTTCTG  
 GGCGACGGGG CTC--ACCTC GC-CTCCCG TTTAGGCCCC AGA-CTTTCG  
 GCTGACGGGG CTC--GCCTC GC-CTCCCG CTTAGGCCCC TGT-CTTCTT  
 NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNCT  
 NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN  
 AGTTACGGGG CCT--GCCTC GT-CACTCG CTTAGGCCCC AGA-CTTCCG  
 GGTGATGGGG CTA--GCCTC GC-CTCCCG CTTAGGTCCC AGA-CTTCTT  
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 NNNNNNNNNN NNNNNNNNTC GC-CTCTCG CTTAGGCCCC AGA-CTTTCG  
 NNNNNNNNNN NNNNNNNNCT GC-CTCTCG CTTAGGCCCC AGA-CTTTCG  
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 GGCGACGGGG CTT--GCTCC GC-CTCCCG CATAGGCCCC AGA-CTTCTT  
 GGCGATAGGG CTT--GCCTT GT-CTCCCG CTTAGGCCCC CGA-TCTCCG  
 NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNCGA TGT-CAGCCT  
 GAAGACGGGG CTC--RCCTC GC-CTCCCG CATCGGCCGA TGT-CATCCT  
 GAAGACGGGG CTC--GCCCC GC-CTCCCG CCTCGGCCGA TGT-CGTCTT  
 GGAGACGGGG CTT--GCCTC GC-CTCACGG CATCGGCTGA TGA-CCTCCT  
 GGAGACGGGG CTC--GCCTC GC-CTCACGG CATCGGCCGA TGA-CCTCCT  
 NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNCGA TGA-CCTCCT  
 GGTGATGGTC ATT--GCACT GT-CTCTGG CTTGACCCT TGT-CGTCTT  
 GGCGACGGTT CTT--GCACC GT-CTCTGG CATAGACCCT TAT-CTTCTT  
 GGTGATGGCC ATA--RTGTC GT-CTCTGG CATGGGCCCT TAT-CCTTCT

IMPERATACYLINDRICA  
 SORGHUMHALEPENSE  
 PANICUMLANIPES  
 PANICUMSTAPFIANUM  
 KARROOCHLOAPURPUREA6241  
 CHAETOBROMUSINVOLUCRATES6284  
 Cynodondactylon7480  
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 Eleusinecoracana  
 Eleusineindica  
 Chlorisvirgata6616  
 Chlorisvirgata7498  
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 Harpochloafalx6530  
 Harpochloafalx5118  
 Harpochloafalx6670  
 Rendliaaltera5133  
 Leptochloafusca7622  
 Enteropogonmacrostachyus9339  
 Dactylocteniumaegyptium9326  
 Tragusberteronianus6620  
 Tragusracemosus7108  
 Traguskoelerioides6729  
 Polevansiarigida7621  
 Perotispatens9323  
 Eragrostiscurvula6529  
 Eragrostisobtusa6623  
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 Eragrostisechinochloidea7485  
 Eragrostispseudobtusa7508  
 Eragrostislehmanniana20  
 Eragrostischloromelas6947  
 Eragrostischloromelas7474  
 Eragrostisracemosa6533  
 Eragrostisbiflora6705  
 Eragrostissuperba7470  
 Eragrostiscapensis6551  
 Eragrostiscapensis6532  
 Pogonarthriasquarrosa7483  
 Pogonarthriasquarrosa7504  
 Catalepisgracilis6962  
 Catalepisgracilis6653  
 Cladoraphiscyperoides5704  
 Cladoraphis spinosa6335  
 Cladoraphiscyperoides4894  
 Fingerhuthiaafricana7507  
 Entoplocamia aristulata9353  
 Enneapogon scoparius18  
 Enneapogon cenchroides7506  
 Enneapogon scoparius7487  
 Schmidtiapappophoroides6334  
 Schmidtiakalihariensis7490  
 Schmidtiakalihariensis7486  
 Sporobolusafricanus7476  
 Sporobolus consimilis9354  
 Sporobolus fimbriatus9328

101

CCC---GGGG CGGAG---GG GCCG-CAACA -GAACCCACG GCGCCTAG-G  
 CCT---GGGG CAGAG---GG GCCA-CAAAA -GAACCCACG GCGCCTTA-G  
 TTCGTTGGAT G-GAG----- GCCG-C AAAA -GAACCCACG GCGC AAAA-G  
 TTTGTTGGAG G-GAG----- GCCG-C AAAA -GAACCCACG GCGCCGAA-G  
 CA---GGAG GAGAG---TG GCCG-CAAAA -GAACCAACG GCGCCGAAACG  
 NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN  
 C---AGAA GGGCG---GG GTGC-CAAAA -GAACCCACG GCGCCGTATG  
 C---AGAA GGGCG---GG GTGC-CAAAA -GAACCCACG GCGCCGTATG  
 TG---TAGA AGGTG---GT GGTC-CAAAA -GAACCTACG GCGCCGCATG  
 TG---TAGA AGGTG---GT GGTC-CAAAA -GAACCTACG GCGCCGCATG  
 CC---AGAG GGATG---CG TGGC-CAAAA -GAACCCACG GCGTCGTATG  
 CC---AGAG GGATG---CG TGGC-CAAAA -GAACCCACG GCGTCGTATG  
 AC---GGAG GGATG---GT GTG--CAAAA AGAACCCACG GCGCTGTATG  
 TT---GGAG TGATT---GT GTGC-CAAAG -GAACCCACG GCGCTGTATG  
 TT---GGAG TGATT---GT GTGC-CAAAG -GAACCCACG GCGCTGTATG  
 TT---GGAG GGACG---GG GTGC-CAAAA -GAACCTATG GCGCTGTATA  
 CG---MGAG GKACG---GG GACC-CAAAA -GAACCCACG GCGCCGTTTG  
 CC---TGGG GGACG---GG GGCC-CAAAA -GAACCCACG GCGCCGTATG  
 TC---GGAG GGGAGGAGGG GCCC-CAAAA -GAACCCACG GCACCGTATG  
 T---AGAG GGA---GG GGCC-CAAAA -GAACCCACG GCGCCGTATG  
 TC---AGAG GGA---GG GGCC-CAAAA -GAACCCACG GCGCCGTATG  
 TC---AGAG GGA---GG GGCC-CAAAA -GAACCCACG GCGCCGTATG  
 ATT---ASAG GC AA---GG GGCC-CAAAA -GAACCCACG GCGCCGTATG  
 T---AGGA TGGTG---AGT GGCCACAAA -GAACCC- CG GCGCCGTGTG  
 TA---TGGAG GGCTG---GA GCCG-CAAAA -GAACCCACG GCGCCATCTG  
 A---TGGAG GGCTG---GA GCCG-CAAAA -GAACCCACG GCGCCG-ATG  
 TA---TGGAG GGCTG---GA GCCG-CAAAA -GAACCCACG GCGCCG-ATG  
 TA---TGGAG GGCTG---GA GCCG-CAAAA -GAACCCACG GCGCCG-ATG  
 TA---TGGAG GGCTG---GA GCCG-CAAAA -GAACCCACG GCGCCA-TCG  
 TA---TGGAG GGCTG---GA GCCG-CAAAA -GAACCCACG GCGCCA-TCG  
 TA---TGGAG GGCTG---GG GCCG-CAAAA -GAACCCACG GCGCCA-TGG  
 TA---CSGAG GGATG---GG GCCG-CAAAA -GAACCCACG GCGCCA-TCG  
 TA---TGGAG GTTTG---GG GCCG-CAAAA -GAACCCACG GCGCCA-TCG  
 TA---TGGAG GGATG---GG GCCA-CAACA -GAACCCACG GCGCCG-ATG  
 T---TGGAG GGCTA---GG GCCG-CAAAA -GAACCCACG GCGCCG-ATG  
 T---TGGAG GGGTA---GG GCCG-CAAAA -GAACCCACG GCGCCG-ATG  
 NNNNNNNNNN NNNNNNNNNN NCCA-CAAAA -GAACCCACA GCGCCG-ATG  
 TA---TGGAG GGCTG---GA GCCG-CAAAA -GAACCCACG GCGCCG-ATG  
 TA---TGGAG GGATG---GG GCCG-CAAAA -GAACCCACG GCGCCA-TCG  
 TA---TGGAG GGATG---GG GCCG-CAAAA -GAACCCACG GCGCCA-TCG  
 TA---TGGAT GGCTG---GG GCCG-CAACA -AAACCCACG GCGCCG-ATG  
 TA---TGGAT GGCTG---GG GCCG-CAACA -AAACCCACG GCGCCG-ATG  
 NA---TGGAT GGCTG---GG GCCG-CAACA -AAACCCACG GCGCCG-ATG  
 TA---TGGAG GGATG---GG GCCG-CAACA -GAACCCACG GCGCCGTATG  
 CT---TGGTG GTTTG---GG GCCG-CAACA -GAACCCACG GCGCCGTATG  
 CA---CGGAG GGCTT---CG GACA-CAAAA -GAACCCACG GCGCCGGTTG  
 CA---CGGAG GGCTT---CG GCCA-CAAAA -GAACCCACG G-----GCTG  
 CA---CGGAG GGCTT---CG GCCA-CAAAA -GAACCCACG GCGCCGGTTG  
 AA---CGGAG GGCTT---CG GCCA-CAAAA AGAACACACG GCGCCGGTTG  
 CA---CGGAG GGCTT---CG GCCA-CAAAA -GAACCCACG GCGCCGGTTG  
 CA---CGGAG GGCTT---CG GCCA-CAAAA -GAACCCACG GCGCCGGTTG  
 CA---TGAAG GGGAA---GG GTCT-CAAAC AGAACCCACG GCGCTGGATG  
 TG---TGGAG GGGAA---GG GTCCCAACA -GAACCCACG GCGCCGGATG  
 TA---T-GAC GGGTG---TG GTCCACAAA -GAACCCACG ACGTCCGATG

150

	151	200
IMPERATACYLINDRICA	G-CGTCAAGG -AACACTT-- --CTATTGCC --TTGC-TCG	GCGGAGCGGT
SORGHUMHALEPENSE	G-CGTCAAGG -AACACTC-- --ATGTTGCC --TTGC-ACA	GCGGAGTGGT
PANICUMLANIPES	G-CGTCAAGG -AACACTG-- --ATATTGCC --TTGC-TTG	GGGTCTTGTT
PANICUMSTAPFIANUM	G-CGTCAAGG -AACACTG-- --ATATTGCC --TTGC-TTG	GGGTCTTGTT
KARROOCHLOAPURPUREA6241	G-CGTCAAGG -AACACTT-- --ATATTGCC --TTGC-GCG	GCGCGGTGGC
CHAETOBROMUSINVOLUCRATES6284	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN	NNNNNNNNNN
Cynodondactylon7480	G-CGTCAAGG -AGAACTA-- --ATGTTGCC --TTGC-TTG	GGGCTCGGC
Cynodondactylon	G-CGTCAAGG -AGAACTA-- --ATGTTGCC --TTGC-TTG	GGGCTCGGC
Eleusinecoracana	G-CGTCAAGG -AAAACAT-- --ATATTGCT --TTGC-CGG	TGGTC-CATT
Eleusineindica	G-CGTCAAGG -AAAACAT-- --ATATTGCT --TTGC-CGG	TGGTC-CATT
Chlorisvirgata6616	G-CGTCAAGG -AAAACAT-- --ATGTTGCC --TTGC-CTG	GGGCTGTGAT
Chlorisvirgata7498	G-CGTCAAGG -AAAACAT-- --ATGTTGCC --TTGC-CTG	GGGCTGTGAT
Eustachyspaspaloides5597	G-CGTCAAGG -ATGACTT-- --ATATTGCC --TTGC-TTG	GGGCCACGAT
Harpochloafalx6530	G-CGTCAAGG -AAAAGTA-- --ATATTGAC --TTGC-CTG	GGGCCACAAC
Harpochloafalx5118	G-CGTCAAGG -AAAAGTA-- --ATATTGAC --TTGC-CTG	GGGCCACAAC
Harpochloafalx6670	G-CGTCAAGG -AAAAGTA-- --ATATTGAC --TTGC-CTG	GGGCCACAAC
Rendliaaltera5133	G-CGTCAAGG -AAAACAT-- --ATATTGCC --TTGC-ATG	GGGCCATGGC
Leptochloafusca7622	G-CGTCAAGG -AAAACAT-- --ATATTGCC --TTGC-CTG	GGGCCACGTC
Enteropogonmacrostachyus9339	G-CGTCAAGG -AAAACAT-- --ATATTGCC --TTGC-TTG	GGGCCATGAC
Dactylocteniumaegyptium9326	G-TGTCAAGG AGACCAAT-- --ATTTGCCA ATTTGC-TCG	GGTCCA-GAC
Tragusberteronianus6620	G-CGTCAAGG -AACACTT-- --GTTTTGCA --GTAC-TCG	GGGAAACGAT
Tragusracemosus7108	G-CGTCAAGG -AACACTT-- --GTTTTGCA --GTAC-TCG	GGGAAACGAT
Traguskoelerioides6729	G-CGTCAAGG -AACACTT-- --GTTTTGCA --GTAC-TCA	GAGAAACGAT
Polevansiarigida7621	G-CGTCAAGG -AACACTT-- --GTTTTGCA --TTGC-TTG	GGGAAAGCTAT
Perotispatens9323	G-CGTCAAGG -AACACAA-- --ATGTTGCC --TTGC-TTG	GTGTAATGTC
Eragrostiscurvula6529	G-CGTCAAGG -AACACTA-- --ATATTGCC --TTGCTCAA	GGATTGTGAC
Eragrostisobtusa6623	G-CGTCAAGG -AACACTA-- --ATATTGCC --TTGCCCAA	GGTC-GCGAC
Eragrostisobtusa6732	G-CGTCAAGG -AACACTA-- --ATATTGCC --TTGCCCAA	GGTC-GCGAC
Eragrostisechinochloidea7485	G-CGTCAAGG -AACACTA-- --ATATTGCC --TTGCCCAA	GGTC-GCGAC
Eragrostispseudobtusa7508	G-CGTCAAGG -AACACTA-- --ATATTGCC --TTGCCCAA	GGTC-GCGAC
Eragrostislehmanniana20	G-CGTCAAGG -AACACTA-- --ATATTGCC --TTGCTCAA	GGTTTGTGAC
Eragrostischloromelas6947	G-CGTCAAGG -AACACTA-- --ATATTGCC --TTGCTCAA	GGATTGTGAC
Eragrostischloromelas7474	G-CGTCAAGG -AATACTA-- --ATATTGCC --TTGCCCAA	G-TTCACGAT
Eragrostisracemosa6533	G-CGTCAAGG -AACACTA-- --ATGTTGCC --TTGCCCAA	G-TTCACGAT
Eragrostisbiflora6705	G-CGTCAAGG -AACACTA-- --ATATTGCC --TTGCCCAA	G-TTCACGAT
Eragrostissuperba7470	G-CGTCAAGG -AACACTA-- --CTATTGCC --TTGCCCAA	GG-TCGCGAT
Eragrostiscapensis6551	G-CGTCAAGG -AACACTA-- --TTGTTGCA --TTGCCCAA	GG-TCGCGTT
Eragrostiscapensis6532	G-CGTCAAGG -AACACTA-- --TTGTTGCA --TTGCCCAA	GG-TCGCGTT
Pogonarthriasquarrosa7483	G-CGTCAAGG -AACACTA-- --ATATTGCC --TTGCCCAA	GG-TCACGAA
Pogonarthriasquarrosa7504	G-CGTCAAGG -AACACTA-- --ATATTGCC --TTGCCCAA	GG-TCACGAT
Catalepisgracilis6962	G-CGTCAAGG -AATACCA-- --ATATTGCC --TTGCCCAA	G-TTTATGAT
Catalepisgracilis6653	G-CGTCAAGG -AATACCA-- --ATATTGCC --TTGCCCAA	G-TTTATGAT
Cladoraphiscyperoides5704	G-CGTCAAGG -AACACTA-- --ATATTGCC --TTGCCCAA	GG-CCGTGTT
Cladoraphis spinosa6335	G-CGTCAAGG -AACACTA-- --ATATTGCC --TTGCCCAA	GG-CCGTGTT
Cladoraphiscyperoides4894	G-CGTCAAGG -AACACTA-- --ATATTGCC --TTGCCCAA	GG-CCGTGTT
Fingerhuthiaafricana7507	G-CGTCAAGG -AACACTT-- --CTATTGCC --ATGCCCAA	G-ATTGTGAT
Entoplocamiaaristulata9353	G-CGTCAAGG -AACACTA-- --TTATTGCT --TTGCCCAA	G-ACCGTGAT
Enneapogon scoparius18	G-CGTCAAGG -AACACTT-- --ATATTGCC --TTGCCCAA	GG-TTGCGGT
Enneapogon cenchroides7506	G-CGTCAAGG -AACACTT-- --ATATTGCC --TTGCCCAA	GG-CTGCGGT
Enneapogon scoparius7487	G-CGTCAAGG -AACACTT-- --ATATTGCC --TTGCCCAA	GG-TTGCGGT
Schmidtia pappophoroides6334	G-CGTCAAGG -AACACTA-- --ATGTTGCC --TTGCCCAA	AG-TCGCGGT
Schmidtia kalihariensis7490	G-CGTCAAGG -AACACTG-- --ATATTGCC --TTGCCCAA	GG-CTGCGGC
Schmidtia kalihariensis7486	G-CGTCAAGG -AACACTG-- --ATATTGCC --TTGCCCAA	GG-CTGCGGC
Sporobolus africanus7476	GGCGTCAAGG -AACACTG-- --ATGTTGC --TTGCTTGG	GG-TTTTGGC
Sporobolus consimilis9354	G-CGTCAAGG -AATACTA-- --ATGTTGC --TTGCTCGG	GG-TTACGGC
Sporobolus fimbriatus9328	A-CGTCAAGG -AACACTTTT ATTTATTGC --TTGCACAA	TG-GTGTGAA



IMPERATACYLINDRICA  
 SORGHUMHALEPENSE  
 PANICUMLANIPES  
 PANICUMSTAFFIANUM  
 KARROOCHLOAPURPUREA6241  
 CHAETOBROMUSINVOLUCRATES6284  
 Cynodondactylon7480  
 Cynodondactylon  
 Eleusinecoracana  
 Eleusineindica  
 Chlorisvirgata6616  
 Chlorisvirgata7498  
 Eustachyspaspaloides5597  
 Harpochloafalx6530  
 Harpochloafalx5118  
 Harpochloafalx6670  
 Rendliaaltera5133  
 Leptochloafusca7622  
 Enteropogonmacrostachyus9339  
 Dactylocteniumaegyptium9326  
 Tragusberteronianus6620  
 Tragusracemosus7108  
 Traguskoelerioides6729  
 Polevansiarigida7621  
 Perotispatens9323  
 Eragrostiscurvula6529  
 Eragrostisobtusa6623  
 Eragrostisobtusa6732  
 Eragrostisechinochloidea7485  
 Eragrostispseudobtusa7508  
 Eragrostislehmanniana20  
 Eragrostischloromelas6947  
 Eragrostischloromelas7474  
 Eragrostisracemosa6533  
 Eragrostisbiflora6705  
 Eragrostissuperba7470  
 Eragrostiscapensis6551  
 Eragrostiscapensis6532  
 Pogonarthriasquarrosa7483  
 Pogonarthriasquarrosa7504  
 Catalepisgracilis6962  
 Catalepisgracilis6653  
 Cladoraphiscyperoides5704  
 Cladoraphisspinosa6335  
 Cladoraphiscyperoides4894  
 Fingerhuthiaafricana7507  
 Entoplocamiaaristulata9353  
 Enneapogonscoparius18  
 Enneapogoncnenchroides7506  
 Enneapogonscoparius7487  
 Schmidtiaappophoroides6334  
 Schmidtiakalihariensis7490  
 Schmidtiakalihariensis7486  
 Sporobolusafricanus7476  
 Sporobolusconsimilis9354  
 Sporobolusfimbriatus9328

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CGGCCTGCCT TC--CGCTCC CC-GCGCAGC GAT-GATA-- TCTTAATCCA  
 CGGCATGCCT TC--CGCTCC CT-GAGCAGC GAT-GATA-- TCTTAATCCA  
 CGGCCTGCCG CG--CAAGCC CC-GTGAAT GTT-GCTA-- TCTTAATCAA  
 CGGCCTGCCG CG--CATGTC CC-ATGAAT GTT-GCTA-- TCCAAATCCA  
 CGGCCTGCCG GA--CGCTCC GT-GCGCAGC GAT-TGTA-- TACTAATCCA  
 NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN  
 CGGCTTGCCG GA--TGCACC CC-TTGCAGC GAT-GCTA-- TGGAAATCA  
 CGGCTTGCCG GA--TGCACC CC-TTGCAGC GAT-GCTA-- TGGAAATCA  
 TGGCATGCC TA--TGTACC CC-GTGCAGT GAT-GCAT-- TGTTAAGTCA  
 TGGCATGCC TA--TGTACC CC-GTGCAGT GAT-GCAT-- TGTTAAGTCA  
 CGGCTTGCCG GA--CACACC TC-TGCAGC GAT-GCTA-- TGGAAATCCA  
 CGGCTTGCCG GA--CACACC TC-TGCAGC GAT-GCTA-- TGGAAATCCA  
 CGGCTTGCCG GA--CGTACT CC-TAGCAGC GAT-GCKT-- TGGAAATCCA  
 TGGCTTGCTG GA--CGCACC CG-GTGCAGT GAT-GCTA-- TAAAAATTTA  
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 CGGCTTGCCG GACGTGCACC CG--TGCAGA GATTGATA-- TGGAAATCCA  
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 CGGCTTGCCG AA--CGAACC TC-GGGTTGC ACT-GATA-- --TTAATCAA  
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 CGGCTTGCCG GA--CGTGTC TT-GAGCGGC GATACCTA-- TTGTAATCCA  
 CGGCTTGCCG GA--CGCGAC TT-GAGCGGC GATACCTA-- TCTTAATCCA  
 CGGCCTGCCG AG--CGCGTC TT-GAGCAGC GATACCTA-- TTGTAATCCA  
 TGGCTTGCCG GA--CACGTC TT-GAGCAGC GATACCTA-- CCATAATCCA  
 TGGCTTGCCG GA--CACGTC TT-GAGCAGC GATACCTA-- CCATAATCCA  
 CGGCTTGCCG GA--CGTGTC TT-GAGCAGC GATACATA-- TCATAATCCA  
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 CGGCTTGCCG GA--CAAGTC TT-GAGCAGC GATACCTA-- TCTTAATCCA  
 CGGCTTGCCG GA--CAAGTC TT-GAGCAGC GATACCTA-- TCTTAATCCA  
 TGGCCTGCCA AA--CGCGTC TT-GGGCAGC GATACCAA-- TTATAATCCA  
 TGGCCTGCCG GA--CGCGTC TT-GGGCAGC GATACCCA-- TTATAATCCA  
 TGGCCTGCCA AA--CGCGTC TT-GGGCAGC GATACCAA-- TTATAATCCA  
 CGGCCTGCCG GA--CGCGTC TT-GGGCAGC GAT-GATA-- TCTTAATCCA  
 CGGCTTGCCG GA--CGCGTC TT-GAGCAGC GAT-GCTA-- TCTTAATCCA  
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 CGGCATGCCG TT--CGCGTC TT-GAGCAGC GAT-GCTA-- TCTCAATCCA  
 CAGCCTGCTG GC--AGATCC CT-GAGCTGC GAT-GGTA-- CCTTAATCCA  
 CGGCCTGCTG GA--CGAGCC CT-GTGCAGC GAT-GCTATA CCTGAATCCA  
 CTCTCTGTTT GA--TACACC TT-GTGCAGT GAT-GCTA-- CCTTAATCCA

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IMPERATACYLINDRICA	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
SORGHUMHALEPENSE	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
PANICUMLANIPES	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
PANICUMSTAFFIANUM	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
KARROOCHLOAPURPUREA6241	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
CHAETOBROMUSINVOLUCRATES6284	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Cynodondactylon7480	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Cynodondactylon	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eleusinecoracana	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eleusineindica	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Chlorisvirgata6616	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Chlorisvirgata7498	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eustachyspaspaloides5597	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Harpochloafalx6530	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Harpochloafalx5118	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Harpochloafalx6670	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Rendliaaltera5133	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Leptochloafusca7622	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eragrostipogonmacrostachyus9339	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Dactylocteniumaegyptium9326	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Tragusberteronianus6620	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Tragusracemosus7108	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Traguskoelerioides6729	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Polevansiarigida7621	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Perotispatens9323	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eragrostiscurvula6529	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eragrostisobtusa6623	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eragrostisobtusa6732	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eragrostisechinoclodea7485	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eragrostispseudobtusa7508	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eragrostislehmanniana20	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eragrostischloromelas6947	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eragrostischloromelas7474	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eragrostisracemosa6533	NCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eragrostisbiflora6705	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eragrostissuperba7470	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eragrostiscapensis6551	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Eragrostiscapensis6532	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Pogonarthriasquarrosa7483	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Pogonarthriasquarrosa7504	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Catalepisgracilis6962	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Catalepisgracilis6653	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Cladoraphiscyperoides5704	NCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Cladoraphis spinosa6335	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Cladoraphiscyperoides4894	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Fingerhuthiaafricana7507	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Entoplocamia aristulata9353	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Enneapogonscoparius18	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Enneapogonscenchroides7506	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Enneapogonscoparius7487	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Schmidtia pappophoroides6334	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Schmidtia kalihariensis7490	NNNNNNNNNN	NNNNTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Schmidtia kalihariensis7486	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Sporobolusafricanus7476	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT
Sporobolus consimilis9354	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NATCCCAGCA	ACCATCGAGT
Sporobolus fimbriatus9328	GCAAAATGCG	ATACCTGGTG	TGAATTGCAG	AATCCCAGCA	ACCATCGAGT

IMPERATACYLINDRICA	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGTCGAGGG	CACGTCTGCC
SORGHUMHALEPENSE	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	CGGCCGAGGG	CACGTCTGCC
PANICUMLANIPES	TTTTGAACGC	AAGTTGCGCC	CGAGACCTTC	TGGTTGAGGG	CACGCCTGCC
PANICUMSTAFFIANUM	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTT	TGGTTGAGGG	CACGTCTGCC
KARROOCHLOAPURPUREA6241	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGTCGAGGG	CACCTCTGCC
CHAETOBROMUSINVOLUCRATES6284	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGCCTGCC
Cynodondactylon7480	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTG	TGGCCGAGGG	CACGTCTGCC
Cynodondactylon	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTG	TGGCCGAGGG	CACGTCTGCC
Eleusinecoracana	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTT	TGGTCGAGGG	CACGTCTGCC
Eleusineindica	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTT	TGGTCGAGGG	CACGTCTGCC
Chlorisvirgata6616	TTTTGAACGC	AAGTTGCGCC	TAAGGCCTTT	TGGCCAAGGG	CACGTCTGCC
Chlorisvirgata7498	TTTTGAACGC	AAGTTGCGCC	TAAGGCCTTT	TGGCCAAGGG	CACGTCTGCC
Eustachyspaspaloides5597	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTT	TGGTCGAGGG	CACGTCTGCC
Harpochloafalx6530	TTTTGAATGC	AAGTTGCGCC	CGAGGCCTTT	TGGCTAAGGG	TACGTTTGCT
Harpochloafalx5118	TTTTGAATGC	AAGTTGCGCC	CGAGGCCTTT	TGGCTAAGGG	TACGTTTGCT
Harpochloafalx6670	TTTTGAATGC	AAGTTGCGCC	CGAGGCCTTT	TGGCTAAGGG	TACGTTTGCT
Rendliaaltera5133	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTT	GGGTTGAGGG	CACGTCTGCC
Leptochloafusca7622	TTTTGAACGC	AAGTTGCGCC	TGAGGCCTTC	TGGCTGAGGG	CACGTCTGCC
Enteropogonmacrostachyus9339	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTT	CGGCCGAGGG	CACGTCTGCC
Dactylocteniumaegyptium9326	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGACCGAGGG	CACGTCTGCC
Tragusberteronianus6620	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Tragusracemosus7108	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Traguskoelerioides6729	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Polevansiarigida7621	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Perotispatens9323	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTT	TGGTTGAGGG	CACGTCTGCC
Eragrostiscurvula6529	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Eragrostisobtusa6623	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGTCGAGGG	CACGTCTGCC
Eragrostisobtusa6732	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGTCGAGGG	CACGTCTGCC
Eragrostisechinochloidea7485	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Eragrostispseudobtusa7508	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGTCGAGGG	CACGTCTGCC
Eragrostislehmanniana20	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Eragrostischloromelas6947	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Eragrostischloromelas7474	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Eragrostisracemosa6533	TTTTGAACGC	AAGTTGCGCC	CGAG - CCTTC	TGGCCGAGG -	CACGTCTGCC
Eragrostisbiflora6705	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGTCGAGGG	CACGTCTGCC
Eragrostisissuperba7470	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Eragrostiscapensis6551	TTTTGAACGC	AAGTTGCGCC	CGAGACCTTC	TGGTTGAGGG	CACGTCTGCC
Eragrostiscapensis6532	TTTTGAACGC	AAGTTGCGCC	CGAGACCTTC	TGGTTGAGGG	CACGTCTGCC
Pogonarthriasquarrosa7483	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Pogonarthriasquarrosa7504	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Catalepisgracilis6962	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Catalepisgracilis6653	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Cladoraphiscyperoides5704	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Cladoraphisspinosa6335	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCAAGGG	CACGTCTGCC
Cladoraphiscyperoides4894	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Fingerhuthiaafricana7507	TTTTGAACGC	AAGTTGCGCC	CGAGACCTTC	TGGTCGAGGG	CACGTCTGCC
Entoplocamiaaristulata9353	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGCCTGCC
Enneapogonscoparius18	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Enneapogoncnenchroides7506	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Enneapogonscoparius7487	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Schmidtiaappophoroides6334	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Schmidtiaalihariensis7490	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Schmidtiaalihariensis7486	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTTC	TGGCCGAGGG	CACGTCTGCC
Sporobolusafricanus7476	TTTTGAACGC	AAGTTGCGCC	TGAGGCCTCC	TGGTTGAGGG	CACGTCTGCC
Sporobolusconsimilis9354	TTTTGAACGC	AAGTTGCGCC	CGAGGCCTCC	TGGTTGAGGG	CACGTCTGCC
Sporobolusfimbriatus9328	TTTTGAACGC	AAGTTGCGCC	TGAGGCCTACT	TGGTCGAGGG	CACGTCTGCC

IMPERATACYLINDRICA	TGGGCGTCAC	GCCAAAAGAC	A-CTCCAAC	CCACCCTC-G	GG-----
SORGHUMHALEPENSE	TGGGCGTCAC	GCCAACAGAC	A-CTCCAAC	CCACCCTC-G	GG-----
PANICUMLANIPES	TGGGCGTCAC	GCCAAAAGAC	A-CTCCAAC	CCATCCTT-G	GG-----
PANICUMSTAFFIANUM	TGGGCGTCAC	GCCAAAAGAC	A-CTCTACC	CCATACTATG	GG-----
KARROOCHLOAPURPUREA6241	TGGGCGTCAC	GCCAAAAGAC	G-CTCCRC	CTACCTTT-G	GG-----
CHAETOBROMUSINVOLUCRATES6284	TGGGCGTCAC	GCCAAAAGAC	A-CTCCGAAC	CCCCCAA--C	GCAAGTTGTA
Cynodondactylon7180	TGGGCGTCAC	GCCAAAAGAC	A-CT-CCACC	GCAATCCC-G	GT-----
Cynodondactylon	TGGGCGTCAC	GCCAAAAGAC	A-CTTCCACC	GCAATCCC-G	GT-----
Eleusinecoracana	TGGGCGTCAC	GTCAAAAGAC	A-CTCCCTAC	CATTCTTT-G	GT-----
Eleusineindica	TGGGCGTCAC	GTCAAAAGAC	A-CTCCCTAC	CATTCTTT-G	GT-----
Chlorisvirgata6616	TGGGCGTCAC	GCCAAAAGAC	A-CTCCACAC	CAACTCT--G	GT-----
Chlorisvirgata7498	TGGGCGTCAC	GCCAAAAGAC	A-CTCCACAC	CAACTCT--G	GT-----
Eustachyspaspaloides5597	TGGGCGTCAC	GCCAAAAGAC	A-CTCCATAC	CCAACCC--G	GT-----
Harpochofalx6530	TGGGCGTCAC	ACCAAAAGAC	A-CTCCACAC	CTGTCTT--G	GT-----
Harpochofalx5118	TGGGCGTCAC	ACCAAAAGAC	A-CTCCACAC	CTGTCTT--G	GT-----
Harpochofalx6670	TGGGCGTCAC	ACCAAAAGAC	A-CTCCACAC	CTGTCTT--G	GT-----
Rendliaaltera5133	TGGGCGTCAT	GTGAAAAGAC	A-CTCCACAC	CAATCC--A	GT-----
Leptochloafusca7622	TGGGCGTCAC	GCCAAAAGAC	A-CTCTGCAC	CTACCCT--G	GT-----
Enteropogonmacrostachyus9339	TGGGCGTCAC	ACCAAAAGAC	A-CTCCAAAC	CAATCCC--G	GT-----
Dactylocteniumaegyptium9326	TGGGCGTCAC	GCCAGTATAC	AACTCCCCAC	CCATCCC--G	GT-----
Tragusberteronianus6620	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CAATCTT--G	GT-----
Tragusracemosus7108	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CAATCTT--G	GT-----
Traguskoelerioides6729	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CAATCTT--G	GT-----
Polevansiarigida7621	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CAAATTT--G	GT-----
Perotispatens9323	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCTAC	CCAATTT--G	GT-----
Eragrostiscurvula6529	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATCAT--G	GT-----
Eragrostisobtusata6623	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CAATCAT--G	GT-----
Eragrostisobtusata6732	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CAATCAT--G	GT-----
Eragrostisechinochloidea7485	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CAATCAT--G	GT-----
Eragrostispseudobtusata7508	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CAATCAT--G	GT-----
Eragrostislehmanniana20	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATCAT--G	GT-----
Eragrostischloromelas6947	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATCAT--G	GT-----
Eragrostischloromelas7474	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATCAT--G	GT-----
Eragrostisracemosa6533	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATCAT--G	GT-----
Eragrostisbiflora6705	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATCAT--G	GT-----
Eragrostissuperba7470	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCTAC	CCATCGT--G	GT-----
Eragrostiscapensis6551	TGGGCGTCAC	GCCAAAAGAC	A-CTCCACAC	CCATCGT--G	GT-----
Eragrostiscapensis6532	TGGGCGTCAC	GCCAAAAGAC	A-CTCCACAC	CCATCGT--G	GT-----
Pogonarthriasquarrosa7483	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATCAT--G	GT-----
Pogonarthriasquarrosa7504	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATCAT--G	GT-----
Catalepisgracilis6962	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATCAT--G	GT-----
Catalepisgracilis6653	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATCAT--G	GT-----
Cladoraphiscyperoides5704	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCAATCT--G	GT-----
Cladoraphisspinosa6335	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCAATCT--G	GT-----
Cladoraphiscyperoides4894	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCAATCT--G	GT-----
Fingerhuthiaafricana7507	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATCGC--G	GT-----
Entoplocamiaaristulata9353	TGGGCGTCAC	GCCAAAAGAC	A-CTCCAAAC	CCATCCC--G	GT-----
Enneapogonscoparius18	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATACA--G	GT-----
Enneapogoncenchroides7506	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATACC--G	GT-----
Enneapogonscoparius7487	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATACA--G	GT-----
Schmidtiaappophoroides6334	TGGGCGTCAC	GGCAAAAGAC	A-CTCCCGAC	CCAACCC--G	GT-----
Schmidtialihariensis7490	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATCTC--G	GT-----
Schmidtialihariensis7486	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCAC	CCATCTC--G	GT-----
Sporobolusafricanus7476	TGGGCGTCAC	GCCAAAGTAC	A-CTTACATA	CCATTCT--G	GT-----
Sporobolusconsimilis9354	TGGGCGTCAC	GCCAAAAGAC	A-CTCCCCTG	CCATCCT--G	GT-----
Sporobolusfimbriatus9328	TGGGCGTCAC	GCGAAAAGAC	A-CTCCCCA	CCCTTCA-TG	GA-----

IMPERATACYLINDRICA	-----GAGGG	ACGTGGTGTT	TGGACCCCG	CG----	CCG-	CAGGGCGCGG
SORGHUMHALEPENSE	-----GAGGG	ATGTGGTTTT	TGGCTCCCC	TG----	CCG-	CATGGCGCGG
PANICUMLANIPES	-----TACCG	ACGTGGTGTT	TGGCTCCCC	TG----	CCT-	CATGGTGTTG
PANICUMSTAFFIANUM	-----GTACCG	ATGTGGTGTT	TGGCTCCCC	TG----	CCT-	TGCGGCATGG
KARROOCHLOAPURPUREA6241	-----TGAGG	ATGCGGTGTA	TGGCTCCTCG	TG----	CCG-	-TGAGGCGCG
CHAETOBROMUSINVOLUCRATES6284	ACGGGAACGG	ACGCGGCATA	TGGCCCCCG	TG----	TCCG	CAAGGCGCGG
Cynodondactylon7480	-----CTGG	ACGTGGTGTT	TGGCCCCCA	TT----	CCAT	AGTGT-ATGG
Cynodondactylon	-----CTGG	ACGTGGTGTT	TGGCCCCCA	TT----	CCAT	AGTGT-ATGG
Eleusinecoracana	-----GTGG	ACGTGGA-TT	TGGCTCCTCA	TG----	CCTT	AGGGCGTGGT
Eleusineindica	-----GTGG	ACGTGGA-TT	TGGCTCCTCA	TG----	CCTT	CGGGCGTGGT
Chlorisvirgata6616	-----ATGG	ACGTGGTGTT	TGGCCCCCA	TA----	CCAC	ATGGTTATGA
Chlorisvirgata7498	-----ATGG	AYGTGGTGTT	TGGCCCCCA	TA----	CCAC	ATGGTTATGA
Eustachyaspaloides5597	-----ATGG	ATGTGGTGTT	TGGCCCCCA	TC----	CCAT	AGGGT-ATGA
Harpochloafalx6530	-----TTGG	ATGTGGTGCA	TGGCCCCCA	TA----	CCAT	CGGGT-TGA
Harpochloafalx5118	-----TTGG	ATGTGGTGCA	TGGCCCCCA	TA----	CCAT	CGGGT-TGA
Harpochloafalx6670	-----TTGG	ATGTGGTGCA	TGGCCCCCA	TA----	CCAT	CGGGT-TGA
Rendliaaltera5133	-----CTGG	ACGTGGTATT	TGGCCCCCA	TC----	CTAT	AAGGT-ATGA
Leptochloafusca7622	-----GTGG	ACGTGGTGTT	TGGCCCCCA	TG----	CCG	AGGGT-GTGG
Enteropogonmacrostachyus9339	-----CTGG	ACGTGGTGTT	TGGCCCCCA	TG----	CCCC	AGGGT-GTGG
Dactylocteniumaegyptium9326	-----GCGG	ACGTGGTGTC	CGGCCCCTC	TG----	CCTC	CGGGT-GCGG
Tragusberteronianus6620	-----GTGG	ATGTGGTATT	TGGCCCCTC	TT----	CCTT	ATGGC-ACGG
Tragusracemosus7108	-----GTGG	ATGTGGTATT	TGGCCCCTC	TT----	CCTT	ATGGC-ACGG
Traguskoelerioides6729	-----GTGG	ATGTGGTATT	TGGCCCCTC	TT----	CCTT	ATGGC-ACGG
Polevansiarigida7621	-----GAGG	ACGTGGTATT	TGGCCCCTC	TT----	CCGT	ATGGC-RCGG
Perotispatens9323	-----GGTGG	ATGTGGTGTT	TGGCTCCTCG	TG----	CCGT	C-GGT-GCGG
Eragrostiscurvula6529	-----GTGG	ACGTGGCATT	TGGCTCCCC	TG----	CCAC	ATGGT-GCGG
Eragrostisobtusa6623	-----GCGG	ATGTGGCATT	TGGCTCTCC	TG----	CCAT	ACGGC-GCGG
Eragrostisobtusa6732	-----GCGG	ATGTGGCATT	TGGCTCTCC	TG----	CCAT	ACGGC-GCGG
Eragrostisechinochloidea7485	-----GCGG	ATGTGGCATT	TGGCTCCCC	TG----	CCAC	ACGGC-GCGG
Eragrostispseudobtusa7508	-----GCGG	ATGTGGCATT	TGGCTCTCC	TG----	CCAT	ACGGC-GCGG
Eragrostislehmanna20	-----GTGG	ACGTGGCATT	TGGCTCCCC	TG----	CCAC	ATGGT-GCGG
Eragrostischloromelas6947	-----GTGG	ACGTGGCATT	TGGCTCCCC	TG----	CCAC	ATGGT-GCGG
Eragrostischloromelas7474	-----GTGG	ACGTGGCATT	TGGCTCCCC	TG----	CCTC	ACGGT-GCGG
Eragrostisracemosa6533	-----GTGG	ACGTGGCATT	TGGCTCCCC	TG----	CCAC	ACGGG-GCGG
Eragrostisbiflora6705	-----GTGG	ATGTGGCATT	TGGCTCCCC	TG----	CCTC	ACGGT-GCGG
Eragrostissuperba7470	-----GTGG	ATGTGGCATT	TGGCTCCCC	TG----	CCAC	ATGGT-GCGG
Eragrostiscapensis6551	-----GCGG	ATGTGGCATT	TGGCTCCCC	TG----	CYGC	ATGGT-GCGG
Eragrostiscapensis6532	-----GCGG	ATGTGGCATT	TGGCTCCCC	TG----	CTGC	ATGGT-GCGG
Pogonarthriasquarrosa7483	-----GCGG	ACGTGGCATT	TGGCTCCCC	TG----	CCTC	ACGGT-GCGG
Pogonarthriasquarrosa7504	-----GCGG	ACGTGGCATT	TGGCTCCCC	TG----	CCTC	ACGGT-GCGG
Catalepisgracilis6962	-----GTGG	ACGTGGCATT	TGGCTCCCC	TG----	CCTC	ACGGT-GCGG
Catalepisgracilis6653	-----GTGG	ACGTGGCATT	TGGCTCCCC	TG----	CCTC	ACGGT-GCGG
Cladoraphiscyperoides5704	-----GTGG	ATGTGGCATT	TGGCTCCCC	TG----	CCGC	ACGGT-GCGG
Cladoraphisspinosa6335	-----GCGG	ATGTGGCATT	TGGCTCCCC	TG----	CCGC	ACGGT-GCGG
Cladoraphiscyperoides4894	-----GTGG	ATGTGGCATT	TGGCTCCCC	TG----	CCGC	ACGGT-GCGG
Fingerhuthiaafricana7507	-----GCGG	ACGTGGCATT	TGGCTCCCC	TG----	CCAA	ACGGC-GCGG
Entoplocamiaaristulata9353	-----GCGG	ACGTGGCATT	TGGCTCCCC	TG----	CCAA	ACGGC-ACGG
Enneapogonscoparius18	-----GTGG	ACGTGGTGTT	TGGCTCCCC	TG----	CCAC	AAGGC-GCGG
Enneapogoncenchroides7506	-----GTGG	ACGTGGTGTT	TGGCTCCCC	TG----	CCAC	GAGGC-GCGG
Enneapogonscoparius7487	-----GTGG	ACGTGGTGTT	TGGCTCCCC	TG----	CCAC	AAGGC-GCGG
Schmidtiapappophoroides6334	-----GAGG	ACGTGGCATT	TGGCTCCCC	TG----	CCAC	ATGGC-ATGG
Schmidtiakalihariensis7490	-----GCGG	ACGTGGCATT	TGGCTCCCC	TG----	CCAC	ATGGT-GCGG
Schmidtiakalihariensis7486	-----GCGG	ACGTGGCATT	TGGCTCCCC	TG----	CCAC	ATGGT-GCGG
Sporobolusafricanus7476	-----ATTGG	ATGTGGATTC	TGACCTTCCA	TGCC--	TGAA	AGGGC-GTGG
Sporobolusconsimilis9354	-----GCTGG	ATGTGGCGTC	TGGCTTCCA	TGCCACTAGA	AAGGA-	GTGG
Sporobolusfimbriatus9328	-----GCGGG	ATGTGGTGTT	TGGCTTCCG	TGTCATGTGA	ATGGT-	TCCG

IMPERATACYLINDRICA	TGGGCCGAA- GTT-GGGGCT GCCGGCGAAC CGCGCCGGGC AC-AGCACGT
SORGHUMHALEPENSE	TGGGCCGAA- GTT-GGGGCT GCCGGCGATT CGTGTCCGGC AC-AGCACGT
PANICUMLANIPES	TGGGCTGAA- GTT-GGGGCT GCCGGCATAAC CGTGCCGGGC ACCTGCATGT
PANICUMSTAPFIANUM	TGGGCTAAA- GTT-GTGGCT GCCGGCATAAC CGTGTCCGGC ACCTGCATGT
KARROOCHLOAPURPUREA6241	TGGGCCGAA- GTT-GCGGCT GCCGGCGTAC CGTGCCGGGC AC-AGCACAT
CHAETOBROMUSINVOLUCRATES6284	TGGGCCGAA- GTT-GGGGCT GCCGGCGCAC CCGGCTGGGC AC-AGCACAA
Cynodondactylon7480	TGGGCCAAA- GAG-GCGGCT GCCGGCG --- -GTGCCGGTC AC-AGCAGAT
Cynodondactylon	TGGGCCAAA- GAG-GAGGCT GCCGGCG --- -GTGCCGGTC AC-AGCAGAT
Eleusinecoracana	GGACCAAAG- ATG-GAG-CT GCCGGTT --- -GTGCCGATC GC-AGCACAA
Eleusineindica	GGACCAAAG- ATG-GAG-CT GCCGGTT --- -GTGCCGATC GC-AGCACAA
Chlorisvirgata6616	TGGGCTTAA- GAT-GTGGCT ATCGGCA --- GGTGCCGATC AG-AGCACAA
Chlorisvirgata7498	TGGGCTTAA- GAT-GTGGCT ATCGGCA --- GGTGCCGATC AG-AGCACAA
Eustachyaspaloides5597	TGGGCCAAA- GAT-GCGGTT GCCGGT --- GGTGCTGATC AC-AGCACAT
Harpochloafalx6530	TGGGCCAAA- GTT-GCGGAT GCGGC --- GGTGCTGAAC AC-AGCACAT
Harpochloafalx5118	TGGGCCAAA- GTT-GCGGAT GCGGC --- GGTGCTGAAC AC-AGCACAT
Harpochloafalx6670	TGGGCCAAA- GTT-GCGGAT GCGGC --- GGTGCTGAAC AC-AGCACAT
Rendliaaltera5133	TGGGCCAAA- AAT-GTGGCT GCCGGT --- GGTGCCGATC AC-AGCACTT
Leptochloafusca7622	TGGGCCAAA- TTT-GGGGCT GCCGGC --- GGTGCCGATC AC-AGCACAA
Enteropogonmacrostachyus9339	TAGGCCAAA- GAT-GGGGCT GCCGGC --- GGTGCCGATC AC-AGCACAA
Dactylocteniumaegyptium9326	TGGGCCAAA- GTT-GGGGCT GCCGGC --- CGTGCCGATC AC-AGCACGA
Tragusberteronianus6620	TGGGCCGAA- GTT-AGGGCT GCCGGC --- GGTGCCGATC AC-AGCACAT
Tragusracemosus7108	TGGGCCGAA- GTT-ACGGCT GCCGGC --- GGTGCCGATC AC-AGCACAT
Traguskoelerioides6729	TGGGCCGAA- GTT-AGGGCT GCCGGC --- GGTGCCGATC AC-AGCACAT
Polevansiarigida7621	TGGGCCGAA- GTT-AGGGCT GCCGGC --- GGTGCCGATC AC-AGCACAT
Perotispatens9323	TGGGCCAAA- GTA-GGGGAT ACCGGC --- AGTGATCTC AC-AGCACAA
Eragrostiscurvula6529	TGGGCCGAA- GTT-GGGGCT GCCGGTTTAC GATACCGGTC AC-AGCACAA
Eragrostisobtusa6623	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAA GGTACCGGTC AC-AGCACAA
Eragrostisobtusa6732	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAA GGTACCGGTC AC-AGCACAA
Eragrostisechinochloidea7485	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAA GSTACCGGTC AC-AGCACAA
Eragrostispseudobtusa7508	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAA GGTACCGGTC AC-AGCACAA
Eragrostislehmanniana20	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAC GATACCGGTC AC-AGCACAA
Eragrostischloromelas6947	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAC GATACCGGTC AC-AGCACAA
Eragrostischloromelas7474	TGGGCCGAA- GAT-GGGGCT GCCGGCTTAT GGTGCCGATC AC-AGCACAA
Eragrostisracemosa6533	TGGG-CCAA- ATT-GGGCTG CCCGGCTTAC GGGGCCGGTC AC-ATCACAA
Eragrostisbiflora6705	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAC GGTGTCCGGTC AC-AGCACAA
Eragrostissuperba7470	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAT GGTACCGGTC AC-AGCACAA
Eragrostiscapensis6551	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAT GGTACCGGTC AC-AGCACAA
Eragrostiscapensis6532	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAT GGTACCG-TC AC-AGCACTT
Pogonarthriasquarrosa7483	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAC GGTACCGGTC AC-AGCACAA
Pogonarthriasquarrosa7504	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAC GGTACCGGTC AC-AGCACAA
Catalepisgracilis6962	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAA GGTGCCGGTC AC-AGCACAA
Catalepisgracilis6653	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAA GGTGCCGGTC AC-AGCACAA
Cladoraphiscyperoides5704	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAC GGTACCGGTC AC-AGCACAA
Cladoraphisspinosa6335	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAC GGTACCGGTC AC-AGCACAA
Cladoraphiscyperoides4894	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAC GGTACCGGTC AC-AGCACAA
Fingerhuthiaafricana7507	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAT GGTACCGGTC AC-AGCACAA
Entoplocamiaaristulata9353	TGGGCCGAA- GTT-GGGGCT GCCGGCTTAT GGTACCGGTC AC-AGCACAA
Enneapogonscoparius18	TGGGCCGAA- GTT-GGGGCT GCCGGTGAAA GGTACCGGTC AC-AGCGCAA
Enneapogoncenchroides7506	TGGGCCGAA- GTT-GGGGCT GCCGGTGAAA GGTACCGGTC AC-AGCGCAA
Enneapogonscoparius7487	TGGGCCGAA- GTT-GGGGCT GCCGGTGAAA GGTACCGGTC AC-AGCGCAA
Schmidtiaappophoroides6334	TGGGCCGAA- GTT-GGGGCT GCCGGTGAAA GGTACCGGTC AC-AGCACAA
Schmidtiaalihariensis7490	TGGGCCGAA- GTT-GGGGCT GCCGGTGAAA GGTACCGGTC AC-AGCACAA
Schmidtiaalihariensis7486	TGGGCCGAA- GTT-GGGGCT GCCGGTGAAA GGTACCGGTC AC-AGCACAA
Sporobolusafricanus7476	TAGGCTAAA- GTAAGAGGCT GCT-GC-GAT GGTGCTAGTC AC-AGCACTT
Sporobolusconsimilis9354	TAGGCTAAA- GTAAGAGGCT GCTGGC-TAT GGTGCCGGTC AC-AGCACTT
Sporobolusfimbriatus9328	TGGACTAAA- GTAT-AGGCT GCTAGC-TAT GGGGCTGGTC AT-AGCACTT

IMPERATACYLINDRICA	GGTGGGCGAC	A-TCAGTTGT	TCTCGGTGCA	--GCGCCCN	NNNNNNNNNN
SORGHUMHALEPENSE	GGTGGGCGAC	ACTTAGTTGT	TCTCGGTGCA	--GCGCCCN	NNNNNNNNNN
PANICUMLANIPES	GGTGGGCGAC	A-TAAGTTGT	TCTCGGTGCA	--GTGTCC--	-TGTCACG--
PANICUMSTAPPIANUM	GGTGGGCGAC	A-TTAGTTGT	TCTCGGTGCA	--GTGTCC--	-TGACACG--
KARROOCHLOAPURPUREA6241	GGTGGTCGAC	CAAAAGTTGT	CA-CGGTGT	--GTGCCTCG	-GTGCGTA-C
CHAETOBROMUSINVOLUCRATES6284	GGTGGGCGTA	ACA--TCCCT	TCAACGTGTM	--GTGCCTCG	-GCCGGTAGC
Cynodondactylon7480	GGTGGATGGC	ACAA-GTTAT	TCTCGGTGTT	--GTGATCCT	-GACAACCTC
Cynodondactylon	GGTGGATGGC	ACAA-GTTAT	TCTCGGTGTT	--GTGATCCT	-GACAACCTC
Eleusinecoracana	GGTGGATGAC	GAAT-GTTGT	TCTCACTGCT	--TTGATCGA	-AACAGCTCC
Eleusineindica	GGTGGATGAC	GAAT-GTTGT	TCTCACTGCT	--TTGATCGA	-AACAGCTCC
Chlorisvirgata6616	GGTGGATGAC	ACAT-GTTAT	TCTCGGTGTT	--ATGATCTG	-GAGAACTCC
Chlorisvirgata7498	GGTGGATGAC	ACAT-GTTAT	TCTCGGTGTT	--ATGATCTG	-SAGAACTCC
Eustachyspaspaloides5597	GGTGGATAAC	ACAA-GTTAT	TCTCGGTGTT	TTATTGAAT-	-CATGACACT
Harpochoafalx6530	GGTGGATGAC	ACAT-GTTAT	TCTCGGTGTT	TT----CGTT	--CGGACACA
Harpochoafalx5118	GGTGGATGAC	ACAT-GTTAT	TCTCGGTGTT	TT----CGTT	--CGGACACA
Harpochoafalx6670	GGTGGATGAC	ACAT-GTTAT	TCTCGGTGTT	TT----CGTT	--CGGACACA
Rendliaaltera5133	GGTGGATGAC	ACAA-GTTAT	TCTCTGTGCT	--ATTATCTG	-GACAACCTC
Leptochloafusca7622	GGTGGATGAC	GCAA-GTTGT	TCTCGGTGTT	--ATGATCCG	-GACCCTCC
Enteropogonmacrostachyus9339	GGTGGATGAC	GCAA-GTTGT	TCTCGGTGTT	--AGGATCCG	-GTGACTAA
Dactylocteniumaegyptium9326	GGTGGATGAC	GCAT-GTTGT	TCTCACTGCT	--AAAATCCG	-TACAGCTT-
Tragusberteronianus6620	GGTGGATAAC	GCGA-GTTGT	TCTCGGTGCT	--ATGATTCG	-GACTCCTTT
Tragusracemosus7108	GGNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Traguskoelerioides6729	GGTGGATAAC	GCGA-GTTGT	TCTCGGTGCT	--ATGATTCG	-GACTCCTTT
Polevansiarigida7621	GGTG-ATGAC	TTTT-GTTGT	TCTCGGTGCT	--ATGTTTCG	-GATGCCTTT
Perotispatens9323	GGTGGATGGC	ACAG-GTTAT	TCTCGGTGCT	--GTGATGCT	GTATTTGATT
Eragrostiscurvula6529	GGTGGGTGAC	ACAT-AGTGT	TCTCGCTGCT	--GTGGCCTG	-GATCATTGC
Eragrostisobtusa6623	GGTGGGTGAC	ACAT-AGTGT	TCTCGCTGTT	--GTGGCCTG	-GACTATTGC
Eragrostisobtusa6732	GGTGGGTGAC	ACAT-AGTGT	TCTCGCTGTT	--GTGGCCTG	-GACTATTGC
Eragrostisechinochloidea7485	GGTGGGTGAC	ACAT-AGTGT	TCTCGCTGTT	--GTGGCCTG	-GACTATTGC
Eragrostispseudobtusa7508	GGTGGGTGAC	ACAT-AGTGT	TCTCGCTGTT	--GTGGCCTG	-GACTATTGC
Eragrostislehmanniana20	GGTGGGTGAC	ACAT-AGTGT	TCTCGCTGTT	--GTGGCCTG	-GACTATTGC
Eragrostischloromelas6947	GGTGGGTGAC	ACAT-AGTGT	TCTCGCTGTT	--GTGGCCTG	-GACTATTGC
Eragrostischloromelas7474	GGTGGGTGAC	ACTC-GGTGT	TCTCGCTGCT	--GTGACTCT	-AACCCT-GC
Eragrostisracemosa6533	GGGGGGTGAC	ACAC-GGTGT	TCTCGCTGCT	--GTGACTTG	-AATCGTTGC
Eragrostisbiflora6705	GGTGGGTGAC	ACTT-GGTGT	TCTCGCTGCT	--GTGACTTG	-AACCCT-GC
Eragrostissuperba7470	GGTGGGTGAC	ACAT-AGTGT	TCTCGCTGCT	--GTGGCCTG	-GATCAATGC
Eragrostiscapensis6551	GGTGGGTGAC	ACAC-GGTGT	TCTCGCTGTT	--GTGACTTC	-TACCGTGC
Eragrostiscapensis6532	GGTGGACAAC	ATGC-GTTGT	TCTCGGAGTT	--GTGACTTG	-TACTGCTTC
Pogonarthriasquarrosa7483	GGTGGGTGAC	ACAT-GGTGT	TCTCGCTGCT	--GTGACATG	-GACCCTGC
Pogonarthriasquarrosa7504	GGTGGGTGAC	ACAT-GGTGT	TCTCGCTGCT	--GTGACATG	-GACCCTGC
Catalepisgracilis6962	GGTGGGTGAC	ACTC-GGTGT	TCTCGCTGCT	--GTGACTTG	-AACTGT-GC
Catalepisgracilis6653	GGTGGGTGAC	ACTC-GGTGT	TCTCGCTGCT	--GTGACTTG	-AACTGT-GC
Cladoraphiscyperoides5704	GGTGGGTGAC	ACAT-GTTGT	TCTCGCTGCT	--GTGACATG	-GCCGCT-GC
Cladoraphisspinosa6335	GGTGGGTGAC	ACAT-GTTGT	TCTCGCTGCT	--GTGACATG	-GCCGCT-GC
Cladoraphiscyperoides4894	GGTGGGTGAC	ACAT-GTTGT	TCTCGCTGCT	--GTGACATG	-GCCGCT-GC
Fingerhuthiaafricana7507	GGTGGGTGAC	ACTT-GWTGT	TCTCGCTGCT	--GCGACACG	-GACCCTT-C
Entoplocamiaaristulata9353	GGTGGGTGAC	ACTT-AGTGT	TCTCGCTGCT	--GCGACCTG	-GACCCTT-G
Enneapogonscoparius18	GGTGGGTGAC	ACAA-GGTGT	TCTCGCCGCA	--GTGGCTCG	-GATCACATC
Enneapogoncenchroides7506	GGTGGGTGAC	ACAC-GGTGT	TCTCGCCGCA	--GTGGCTCG	-GATCACATC
Enneapogonscoparius7487	GGTGGGTGAC	ACAA-GGTGT	TCTCGCCGCA	--GTGGCTCG	-GATCACATC
Schmidtiapappohoroides6334	GGTGGGTGAC	ACCG-GGTGT	TCTCGCTGCT	--CTGACTTG	-GATCACTGC
Schmidtiakalihariensis7490	GGTGGGTGAC	ACAA-GGTGT	TCTCGCTGCT	--GTGACTCG	-GATCCATGA
Schmidtiakalihariensis7486	GGTGGGTGAC	ACAA-GGTGT	TCTCGCTGCC	--GTGACTCG	-GATCCATGA
Sporobolusafricanus7476	GGTGGACAAC	ATGC-GTTGT	TCTCGGAGTN	NNNNNNNNNN	NNNNNNNNNN
Sporobolusconsimilis9354	GGTGGACAAC	GTGC-GTTGT	TCTCGGAGTT	--GTGACTTG	-TACTGGTTT
Sporobolusfimbriatus9328	GGTGGATGAC	AAAT-GTTGT	TCTCGGAGTT	--GTGACATG	-TATTGCTTC



IMPERATACYLINDRICA  
 SORGHUMHALEPENSE  
 PANICUMLANIPES  
 PANICUMSTAFFIANUM  
 KARROOCHLOAPURPUREA6241  
 CHAETOBROMUSINVOLUCRATES6284  
 Cynodondactylon7480  
 Cynodondactylon  
 Eleusinecoracana  
 Eleusineindica  
 Chlorisvirgata6616  
 Chlorisvirgata7498  
 Eustachyspaspaloides5597  
 Harpochloafalx6530  
 Harpochloafalx5118  
 Harpochloafalx6670  
 Rendliaaltera5133  
 Leptochloafusca7622  
 Enteropogonmacrostachyus9339  
 Dactylocteniumaegyptium9326  
 Tragusberteronianus6620  
 Tragusracemosus7108  
 Traguskoelerioides6729  
 Polevansiarigida7621  
 Perotispatens9323  
 Eragrostiscurvula6529  
 Eragrostisobtus6623  
 Eragrostisobtus6732  
 Eragrostisechinochloidea7485  
 Eragrostispseudobtus6708  
 Eragrostislehmannaiana20  
 Eragrostischloromelas6947  
 Eragrostischloromelas7474  
 Eragrostisracemosa6533  
 Eragrostisbiflora6705  
 Eragrostissuperba7470  
 Eragrostiscapensis6551  
 Eragrostiscapensis6532  
 Pogonarthriasquarrosa7483  
 Pogonarthriasquarrosa7504  
 Catalepisgracilis6962  
 Catalepisgracilis6653  
 Cladoraphiscyperoides5704  
 Cladoraphisspinosa6335  
 Cladoraphiscyperoides4894  
 Fingerhuthiaafricana7507  
 Entoplocamiaaristulata9353  
 Enneapogonscoparius18  
 Enneapogoncenchroides7506  
 Enneapogonscoparius7487  
 Schmidtiapappophoroides6334  
 Schmidtiakalihariensis7490  
 Schmidtiakalihariensis7486  
 Sporobolusafricanus7476  
 Sporobolusconsimilis9354  
 Sporobolusfimbriatus9328

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 --TGGTACAC AAT-GGCCCT CAT--GACCC AT--CAACC GAAGCGCAC-  
 C---TATGC AAT-AGTCT CGC--GACCC AT--CAACC GAAGCGCAT-  
 C---GGCG-AC ATT-GGCACT TT--GGACC CAT--GAATT G-AGCAAAT-  
 C----ATGC AAG-GGCCCT TGT--GACCC AT--CAACC -AAGCGCAT-  
 C----ATGC AAG-GGCCCT TGT--GACCC AT--CAACC -AAGCGCAT-  
 A--GGCATGC AAT-GGCCCT CAT--GACCC AT--GAACC AAAGAGCAT-  
 A--GGCATGC AAT-GGCCCT CAT--GACCC AT--GAACC AAAGAGCAT-  
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 C---TATGC AAT-GGCCCT CAT--GACCC AT--CAACC GATGCGCAC-  
 C---TATGC AAT-GGCCCT CAT--GACCC AT--CAACC GATGCGCAC-  
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 C---AAGGC AAC-GACCCT CAT--GACCC AT--CGACC GAAGCGCAC-  
 CA-GGCAAGC A-T-GGCCCT CAA--GACCC AT--CGACC GTAGCGCAC-  
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 CA-GGCAAGC A-T-GGCCCT CAA--GACCC AT--CGACC GTAGCGCAC-  
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 CT-GGCAAGC AAT-GGCCCT CAT--GACCC TT--CGACC GTAGCGCAC-  
 CT-GGCAAGC AAT-GGCCCT CAT--GACCC TT--CGACC GTAGCGCAC-  
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 TAGCGAT-AG TAT-GGCTGT -TA-TGACCC AT--GCATC GGAGCTTTG-

IMPERATACYLINDRICA	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
SORGHUMHALEPENSE	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMLANIPES	GGTG-----	CTCGGACGAT	ATGTATTGAG	CCTTGGTATG	GAAACCTGCT
PANICUMSTAPFIANUM	GGTG-----	CTCGGACGAT	ATGTATTGAG	CCTTGGTATG	GAAACCTGCT
KARROOCHLOAPURPUREA6241	GACG-----	CTCGGACNNT	-TGTATTGAG	CCT-GGTA-G	GAAACCTGCT
CHAETOBROMUSINVOLUCRATES6284	GTCACCTCGGA	TCGCGACAAT	-TGTATTGAG	CCTTGGTATG	GAA-CCTGCT
Cynodondactylon7480	GTAG-----	CTCGGACNNN	NNNNNNNNNN	NNT-GGTATG	GAAACCTGCT
Cynodondactylon	GTAG-----	CTCGGACGAT	-TGTATTGAG	CCTTGGTATG	GAAACCTGCT
Eleusinecoracana	GTTT-----	ATCAAACGAT	-TGTATTGAG	CCTTGGTATG	GAAACCTGCT
Eleusineindica	GTTT-----	ATCAAACGAT	-TGTATTGAG	CCTTGGTATG	GAAACCTGCT
Chlorisvirgata6616	GCCG-----	CTCGGTCGAT	-TGTAT-GAG	CCT-GGTATG	GAA-CCTGCT
Chlorisvirgata7498	GCCG-----	CTCGGTCNNN	NNNNNNNNNN	NNNNNNNNNG	AAA-CCTGCT
Eustachyspaspaloides5597	GTTG-----	CTTGACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Harpochloafalx6530	GCTT-----	CTTGACGAT	-TGTATTGAG	CCTTGGTATG	GA--CCTGCT
Harpochloafalx5118	GCTT-----	CTTGACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Harpochloafalx6670	GCTT-----	CTTGACGAT	-TGTATTGAG	CCTTGGTATG	GAA-CCTGCT
Rendliaaltera5133	GTTG-----	CTCGGACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNCCTGCT
Leptochloafusca7622	GTTG-----	CTCGGACNNN	NNNNNNNNNN	NNNNGGTATG	GAA-CCTGCT
Enteropogonmacrostachyus9339	GTTG-----	CTCGTACNNN	NNNNNNNNNN	NNNNNNNATG	GAA-CCTGCT
Dactylocteniummaegyptium9326	GTTA-----	CTCGGACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Tragusberteronianus6620	GTTG-----	CTCGGACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Tragusracemosus7108	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Traguskoelerioides6729	GTTG-----	CTCGGACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Polevansiarigida7621	GTTG-----	CTCGGACNNN	NNNNNNNNNN	NNNNNNNNNG	GAAACCTGCT
Perotispatens9323	GTCG-----	CTCGAACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostiscurvula6529	GTCG-----	CTCGGACGAT	-TGTATTGAG	CCTTGGTATG	GAA--CTGCT
Eragrostisobtusa6623	G-----	CTCGGACGAT	-TGTATTGAG	C-TTGGTATG	GAAACCTGGT
Eragrostisobtusa6732	G-----	CTCGGACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostisechinocloidea7485	G-----	CTCGGACGAT	-TGTATTGAG	C-TTGGTATG	GAAA-CTGCT
Eragrostispseudobtusa7508	G-----	CTCGGACGAT	-TGTATTGAG	CC-TGGTATG	GAA--CTGCT
Eragrostislehmanniana20	GTCG-----	CTCGGACNNN	NNGTATTGAG	CCTTG-TATG	GAAACCTGCT
Eragrostischloromelas6947	GTCG-----	CTCGGACGAT	-TGTATTGAG	CCTTGGTATG	GAAACCTGCT
Eragrostischloromelas7474	GTCG-----	CTCGGACGAT	-TGTATTGAG	CCTTGGTATG	GAAACCTGCT
Eragrostisracemosa6533	NNNNNNNNNN	NNNNNNNGAT	-TGTATTGAG	CCTCGGTATG	GAAACCTGCT
Eragrostisbiflora6705	GTCG-----	CTCGGACNNN	NNNNNNNNNN	NNNNNNNNNG	GAAACCTGCT
Eragrostissuperba7470	GTCG-----	CTCGGACGAT	-TGTATTGAG	CCTTGGTATG	GAA--CTGCT
Eragrostiscapensis6551	GTCG-----	CTCGGACGAT	-TGTATTGAG	-CT-GGTATG	GAA-CCTGCT
Eragrostiscapensis6532	GTTG-----	CTCGAACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Pogonarthriasquarrosa7483	G-----	CTCGGACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNCCTGCT
Pogonarthriasquarrosa7504	G-----	CTCGGACGAT	-TGTATTGAG	CCTTGGTATG	GAAACCTGCT
Catalepisgracilis6962	GTTG-----	CTTGACGAT	-TGTATTGAG	CCTTGGTATG	GAAACCTGCT
Catalepisgracilis6653	GTTG-----	CTTGACNNN	NNNNNNNNNN	NNNNGGTATG	GAAACCTGCT
Cladoraphiscyperoides5704	GTCG-----	CTCGGACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Cladoraphisspinosa6335	GTCG-----	CTCGGACNNN	NNNNNNNNNN	NNNNGGTATG	GAAACCTGCT
Cladoraphiscyperoides4894	GTCG-----	CTCGGACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Fingerhuthiaafricana7507	GTCG-----	CTCGGACGAT	-TGTATTGAG	-CT-GGTATG	GAAA-CTGCT
Entoplocamiae aristulata9353	GTCG-----	CTCGGACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Enneapogonscoparius18	GTCG-----	CTCGGACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNCCTGCT
Enneapogoncenchroides7506	GTCG-----	CTCGGACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Enneapogonscoparius7487	GTCG-----	CTCGGACGAT	-TGTATTGAG	CCTTGGTATG	GAAA-CTGCT
Schmidtiaappophoroides6334	GTCG-----	CTCGGATNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Schmidtiaalihariensis7490	GCCG-----	CTCGGACNNN	NTGTATTGAG	C--TGATG	GAA--CTGCT
Schmidtiaalihariensis7486	GCCG-----	CTCGGACGAT	-TGTATTGAG	CCTTGGTATG	GAAACCTGCT
Sporobolusafricanus7476	NNNNNNNNNN	NNNNNNNGAT	--GTAT-GAG	CCT-GGTATG	GAAACCTGCT
Sporobolusconsimilis9354	ATTG-----	CTCGACGAT	-TGTATTGAG	-CT-GGTATG	GAAACCTGCT
Sporobolusfimbriatus9328	ATTG-----	CTTCAACNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN



IMPERATACYLINDRICA	CCTGAGCCAA	ATCCACTTTT	TT----	GAAA	AAA---	CAAG	TG----	CAA
SORGHUMHALEPENSE	CCTGAGCCAA	ATCCACTTTT	TT----	CAA	AAA-----	G	TGGTTCTCAA	
PANICUMLANIPES	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
PANICUMSTAFFIANUM	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
KARROOCHLOAPURPUREA6241	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AA----	CACG	TGGTCTCAA	
CHAETOBROMUSINVOLUCRATES6284	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	CGGTCTCAA	
Cynodondactylon7480	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Cynodondactylon	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AA----	CAAG	TGGTTCTCAA	
Eleusinecoracana	CCTGAGCCAA	ATCCCTTTTT	TTCATTTTAA	AAA---	CAAG		TGGTTCTCAA	
Eleusineindica	CCTGAGCCAA	ATCCCTTTTT	TTCATTTTAA	AAA---	CAAG		TGGTTCTCAA	
Chlorisvirgata6616	CCTGAGCCAA	ATCCCTTTTT	T-----	TAAA	AA----	CAAG	TGGTTCTCAA	
Chlorisvirgata7498	CCTGAGCCAA	ATCCCTTTTT	T-----	TAAA	AA----	CAAG	TGGTTCTCAA	
Eustachyspaspaloides5597	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	
Harpochloafalx6530	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AA----	CAAG	TGGTTCTCAA	
Harpochloafalx5118	CCTGAGCCAA	GTCCCTTTTT	T-----	GAAA	A-----		TGGTTCTCAA	
Harpochloafalx6670	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AA----	CAAG	TGGTTCTCAA	
Rendliaaltera5133	CCTGAGCCAA	ATCCCTGTTT	T-----	GAAA	AA----	CAAG	TGGTTCTCAA	
Leptochloafusca7622	CCTGAGCCAA	ATCCCTTTTT	-----	GAAA	AA----	CAGG	TGGTTCTCAA	
Enteropogonmacrostachyus9339	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AA----	CAAG	TGGTTCTCAA	
Dactylocteniumaegyptium9326	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	
Tragusberteronius6620	CCTGAGCCAA	ATCCCTTTTT	TG----	GAAA	AA----	CAAG	TGGTTCTCAA	
Traguracemosus7108	CCTGAGCCAA	ATCCCTTTTT	TG----	GAAA	AA----	CAAG	TGGTTCTCAA	
Traguskoelerioides6729	CCTGAGCCAA	ATCCCTTTTT	TG----	GAAA	AA----	CAAG	TGGTTCTCAA	
Polevansiarigida7621	CCTGAGCCAA	ATCCCTTTTT	TT----	GAAA	AA----	CAAG	TGGTTCTCAA	
Perotispatens9323	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	
Eragrostiscurvula6529	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAR---	CAAG	TGGTTCTCAA	
Eragrostisobtusa6623	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Eragrostisobtusa6732	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	
Eragrostisechinochloidea7485	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Eragrostispseudobtusa7508	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Eragrostislehmanniana20	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Eragrostischloromelas6947	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Eragrostischloromelas7474	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Eragrostisracemosa6533	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Eragrostisbiflora6705	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Eragrostiscurvula7470	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAG---	CAAG	TGGTTCTCAA	
Eragrostiscapensis6551	C-TGAGCCAA	ATCCCTTTTA	-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Eragrostiscapensis6532	CCTGAGCCAA	ATCCCTTTTA	-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Pogonarthriasquarrosa7483	CCTGAGCCAA	ATCCCTTTTA	-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Pogonarthriasquarrosa7504	CCTGAGCCAA	ATCCCTTTTA	-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Catalepisgracilis6962	CCTGAGCCAA	ATCCCTGTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Catalepisgracilis6653	CCTGAGCCAA	ATCCCTGTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Cladoraphiscyperoides5704	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	TGGTTCTCAA	
Cladoraphisspinosa6335	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAAAACAAG		TGGTTCTCAA	
Cladoraphiscyperoides4894	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNA		
Fingerhuthiaafricana7507	C-TGAGCCAA	ATCCCTTTTT	T-----	GAAA	AA----	CAAG	TGGTTCTCAA	
Entoplocamiaaristulata9353	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AA----	CAAG	TGGTTCTCAA	
Enneapogonscoparius18	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Enneapogoncenchroides7506	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Enneapogonscoparius7487	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Schmidtiaappophoroides6334	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Schmidtialihariensis7490	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAAA--	CAA	CGTTCTCAA	
Schmidtialihariensis7486	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAAA--	CAAG	CGTTCTCAA	
Sporobolusafricanus7476	CCTGAGCCAA	ATCCCTTTTT	T-----	TAAA	AAA---	CAAG	TGGTTCTCAA	
Sporobolusconsimilis9354	CCTGAGCCAA	ATCCCTTTTT	T-----	GAAA	AAA---	CAAG	TGGTTCTCAA	
Sporobolusfimbriatus9328	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNG	TGGTTCTCAA	

IMPERATACYLINDRICA	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
SORGHUMHALEPENSE	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
PANICUMLANIPES	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
PANICUMSTAFFIANUM	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
KARROOCHLOAPURPUREA6241	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
CHAETOBROMUSINVOLUCRATES6284	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Cynodondactylon7480	ACCAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Cynodondactylon	ACCAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Eleusinecoracana	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Eleusineindica	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Chlorisvirgata6616	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Chlorisvirgata7498	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Eustachyspaspaloides5597	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Harpochoalfalx6530	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Harpochoalfalx5118	A-TATAACCC	---GGAAAAG	G-----ATC	TGTGCCGACA	CTCAATGGAA
Harpochoalfalx6670	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Rendliaaltera5133	ACAAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Leptochloafusca7622	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Enteropogonmacrostachyus9339	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Dactylocteniumaegyptium9326	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNCAATGGAA
Tragusberteronianus6620	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Tragusracemosus7108	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Traguskoelerioides6729	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Polevansiarigida7621	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Perotispatens9323	NNNNNAACCC	AAAGCAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Eragrostiscurvula6529	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Eragrostisobtusa6623	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Eragrostisobtusa6732	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostisechinochloidea7485	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Eragrostispseudobtusa7508	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Eragrostislehmanniana20	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Eragrostischloromelas6947	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Eragrostischloromelas7474	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Eragrostisracemosa6533	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Eragrostisbiflora6705	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Eragrostisuperba7470	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Eragrostiscapensis6551	-CTAGAACCC	AAATGAAAAG	G-ATAGGATA	GGTGCAGAGA	CTCAATGGAA
Eragrostiscapensis6532	ACTAGAACCC	AAATGAAAAG	GGATAGGATA	GGTGCAGAGG	CTCAATGGAA
Pogonarthriasquarrosa7483	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Pogonarthriasquarrosa7504	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Catalepisgracilis6962	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Catalepisgracilis6653	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Cladoraphiscyperoides5704	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Cladoraphisspinosa6335	ACTAGAATCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Cladoraphiscyperoides4894	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Fingerhuthiaafricana7507	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Entoplocamiaaristulata9353	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Enneapogonscoparius18	ACTAGAATCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Enneapogoncenchroides7506	ACTAGAATCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Enneapogonscoparius7487	ACTAGAATCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Schmidtiaappophoroides6334	ACTAGAATCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Schmidtiaalihariensis7490	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Schmidtiaalihariensis7486	ACTAGAACCC	AAATGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Sporobolusafricanus7476	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Sporobolusconsimilis9354	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA
Sporobolusfimbriatus9328	ACTAGAACCC	AAAGGAAAAG	G-----ATA	GGTGCAGAGA	CTCAATGGAA

IMPERATACYLINDRICA	GCTGTTCTAA	CGAATCGAAG	T-----	-----	-----
SORGHUMHALEPENSE	GCTGTTCTAA	CGAATCGAAG	T-----	-----	-----
PANICUMLANIPES	GCTGTTCTAA	CGAATCGAA-	-GTAATTACG	TTGTGTTGGT	AGTGGAAC--
PANICUMSTAPFIANUM	GCTGTTCTAA	CGAATCGAA-	-GTAATTACG	TTGTGTTGGT	AGTGGAAC--
KARROOCHLOAPURPUREA6241	GCTGTTCTAA	CGAATCGAGG	T--AATTACG	TTGTGTTGGT	AGTGTAAC--
CHAETOBROMUSINVOLUCRATES6284	GCTGTTCTAA	CGAATCGAGG	T--AATTGCG	TTGTGCTGGT	AGTGTAAT--
Cynodondactylon7480	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Cynodondactylon	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Eleusinecoracana	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Eleusineindica	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Chlorisvirgata6616	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGCGAAAC--
Chlorisvirgata7498	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGCGAAAC--
Eustachyaspaloides5597	NNNNNNNNNN	NNNNNNNNNN	NNNNNTACG	TTGTGTTGGT	AGTGAAAC--
Harpochloafalx6530	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Harpochloafalx5118	GCTGTTCTAA	CGAATCGAGG	TGTAATTACT	TTGTGTTGGT	AGTGAAAC--
Harpochloafalx6670	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Rendliaaltera5133	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Leptochloafusca7622	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Enteropogonmacrostachyus9339	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Dactylocteniumaegyptium9326	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Tragusberteronianus6620	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGCAAC--
Tragusracemosus7108	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGCAAC--
Traguskoelerioides6729	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGCAAC--
Polevansiarigida7621	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Perotispatens9323	GCTGTTCTAA	CGAATCGAGG	TATAATTACG	TTGTGTTGGT	AGTGAACC--
Eragrostiscurvula6529	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	ARTGAAAS--
Eragrostisobtusa6623	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AATGAAAC--
Eragrostisobtusa6732	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostisechinocloidea7485	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AATGAAAC--
Eragrostispseudobtusa7508	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AATGAAAC--
Eragrostislehmanniana20	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AATGAAAC--
Eragrostischloromelas6947	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AATGAAAC--
Eragrostischloromelas7474	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AATGAAAC--
Eragrostisracemosa6533	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AATGAAAC--
Eragrostisbiflora6705	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AATGAAAC--
Eragrostissuperba7470	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAG--
Eragrostiscapensis6551	GCTATTCTAA	CGAATCGAGG	TGTAATA-CG	TTGTGTTGGT	AGTGAAAC--
Eragrostiscapensis6532	GCTATTCTAA	CGAATCGAGG	TGTAATATCG	TTGTGTTGGT	AGTGAATC--
Pogonarthriasquarrosa7483	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AATGAAAC--
Pogonarthriasquarrosa7504	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AATGAAAC--
Catalepisgracilis6962	GCTATTCTAA	CGAATCGAAG	TGTAATTACG	TTGTGTTGGT	AATGAAAC--
Catalepisgracilis6653	GCTATTCTAA	CGAATCGAAG	TGTAATTACG	TTGTGTTGGT	AATGAAAC--
Cladoraphiscyperoides5704	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Cladoraphisspinosa6335	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Cladoraphiscyperoides4894	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Fingerhuthiaafricana7507	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAACTA
Entoplocamiaristulata9353	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Enneapogonscoparius18	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Enneapogoncenchroides7506	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Enneapogonscoparius7487	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Schmidtiaappophoroides6334	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Schmidtialihariensis7490	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Schmidtialihariensis7486	GCTATTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Sporobolusafricanus7476	GCTGTTCTAA	CGAATCAAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Sporobolusconsimilis9354	GCTGTTCTAA	CGAATCGAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--
Sporobolusfimbriatus9328	GCTGTTCTAA	CGAATCCAGG	TGTAATTACG	TTGTGTTGGT	AGTGAAAC--

IMPERATACYLINDRICA	-----	-----	-----	-----
SORGHUMHALEPENSE	-----	-----	-----	-----
PANICUMLANIPES	---TCCCTCG A-----	-----	-----	AATTATAGAA AGAAGGGCTT
PANICUMSTAPFIANUM	---TCCCTCG A-----	-----	-----	AATTATAGAA AGAAGGGCTT
KARROOCHLOAPURPUREA6241	---GCCCTCT A-----	-----	-----	AATTAGAGAA AGAAGGGCTT
CHAETOBROMUSINVOLUCRATES6284	---TCCTTCT A-----	-----	-----	AATTAGAGAA AGAGGGGCTT
Cynodondactylon7480	---TCCCTCT A-----	AATT AC-----	TA	AATTAGAGAA AGAAGGGCTT
Cynodondactylon	---TCCCTCT A-----	AATT AC-----	TA	AATTAGAGAA AGAAGGGCTT
Eleusinecoracana	---TCCCTCT A-----	AATT AC-----	TA	AATTAGAGAA AGAAGAGCTT
Eleusineindica	---TCCCTCT A-----	AATT AC-----	TA	AATTAGAGAA AGAAGAGCTT
Chlorisvirgata6616	---TCCCTCT C-----	-----	TA	AATTAGAGAA AGAAGGGCTT
Chlorisvirgata7498	---TCCCTCT A-----	AATT AC-----	TA	AATTAGAGAA AGAAGGGCTT
Eustachyspaspaloides5597	---TCCCTCT A-----	AATT ACTAAAACTA	AATTAGAGAA	AGAAGGGCTT
Harpochloafalx6530	---TCCCTTT AATTTAAATT	AC-----	TA	AATTAGAGAA AGAAGGGCTT
Harpochloafalx5118	---TCCCTTT AATTTAAATT	AC-----	TA	AATTAGAGAA AGAAGGGCTT
Harpochloafalx6670	---TCCCTTT AATTTAAATT	AC-----	TA	AATTAGAGAA AGAAGGGCTT
Rendlialaltera5133	---TCCCTCT A-----	AATT AT-----	TA	AATTAGAGAA AGAAGGGCTT
Leptochloafusca7622	---TCCCTCT A-----	AATT AC-----	TA	AATTAGAGAA AGAAGAGCTT
Enteropogonmacrostachyus9339	---TCCCTCT A-----	AATT AC-----	TA	AATTAGAGAA AGAAGGGCTT
Dactylocteniumaegyptium9326	---TCCCTCT ACTTTAAA--	AC-----	GA	AATTAGAGAA AGAAGGGCTT
Tragusberteronianus6620	---TCCCTCT A-----	AATT AC-----	TA	AATTAGAGAA AGAAGGGCTT
Tragusracemosus7108	---TCCCTCT A-----	AATT AC-----	TA	AATTAGAGAA AGAAGGGCTT
Traguskoelerioides6729	---TCCCTCT A-----	AATT AC-----	TA	AATTAGAGAA AGAAGGGCTT
Polevansiarigida7621	---TCCCTCT A-----	AATT AC-----	TA	AATTAGAGAA AGAAGGGCTT
Perotispatens9323	---TCCCTCT A-----	AATT AC-----	TA	AATTAGAGAA AGAAGGGCTT
Eragrostiscurvula6529	---TCCCTCT A-----	-----	-----	AATTTGAGAA AGAAGGGCTT
Eragrostisobtusa6623	---TCCCTCT A-----	-----	-----	AATTTGAGAA AGAAGGGCTT
Eragrostisobtusa6732	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eragrostisechinochloidea7485	---TCCCTCT	-----	---	TTGAGAA AGAAGGGCTT
Eragrostispseudobtusa7508	---TCCCTCT A-----	-----	-----	AATTTGAGAA AGAAGGGCTT
Eragrostislehmanniana20	---TCCCTCT A-----	-----	-----	AATTTGAGAA AGAAGGGCTT
Eragrostischloromelas6947	---TCCCTCT	-----	---	TTGAGAA AGAAGGGCTT
Eragrostischloromelas7474	---TCCCTCT A-----	-----	-----	AATTTGAGAA AGAAGGGCTT
Eragrostisracemosa6533	---TCCCTCT A-----	-----	-----	AATTTGAG --
Eragrostisbiflora6705	---TCCCTCT A-----	-----	-----	AATTTGAGAA AGAAGGGCTT
Eragrostissuperba7470	---TCCCTCT A-----	-----	-----	AATTTGAGAA AGAAGGGCTT
Eragrostiscapensis6551	---TCCCTCT A-----	-----	-----	AATTTGAGAA AGAAGGGCTT
Eragrostiscapensis6532	---TCCCTCT A-----	-----	-----	AATTTGAGAA AGAAGGGCTT
Pogonarthriasquarrosa7483	---TCCCTCT A-----	-----	-----	AATTTGAGAA AGAAGGGCTT
Pogonarthriasquarrosa7504	---TCCCTCT A-----	-----	-----	AATTTGAGAA AGAAGGGCTT
Catalepisgracilis6962	---TCCCTCT A-----	-----	-----	AATTTGAGAA AGAAGGGCTT
Catalepisgracilis6653	---TCCCTCT A-----	-----	-----	AATTTGAGAA AGAAGGGCTT
Cladoraphiscyperoides5704	---TCCCTCT A-----	-----	-----	AATTAGAGAA AGAAGGGCTT
Cladoraphisspinosa6335	---TCCCTCT A-----	-----	-----	AATTAGAGAA AGAAGGGCTT
Cladoraphiscyperoides4894	---TNCCTCT A-----	-----	-----	AATTAGAGAA AGAAGGGCTT
Fingerhuthiaafricana7507	AACTCCCTCT A-----	-----	-----	AATTAGAGAA AGAAGGGCTT
Entoplocamiaaristulata9353	---TCCCTCT A-----	-----	-----	AATTAGAGAA AGAAGC-CTT
Enneapogonscoparius18	---TCCCTCT A-----	-----	-----	AATTAGAGAA AGAAGGGCTT
Enneapogoncenchroides7506	---TCCCTCT A-----	-----	-----	AATTAGAGAA AGAAGGGCTT
Enneapogonscoparius7487	---TCCCTCT A-----	-----	-----	AATTAGAGAA AGAAGGGCTT
Schmidtiaappophoroides6334	---TCCCTCT A-----	-----	-----	AATTCGAGAA AGAAGGGCTG
Schmidtiaalihariensis7490	---TCCCTCT A-----	-----	-----	AATTAGAGAA AGAAGGGCTT
Schmidtiaalihariensis7486	---TCCCTCT A-----	-----	-----	AATTAGAGAA AGAAGGGCTT
Sporobolusafricanus7476	---TCCCTCT A-----	AATT AC-----	TA	AATTAGAGAA AGAAGAAGCTT
Sporobolusconsimilis9354	---TCCCTCT A-----	AATT AC-----	TA	AATTAGAGAA AGAAGAGCTT
Sporobolusfimbriatus9328	---TCCCTCA A-----	AATT AC-----	TA	AATTAAGAA AGAAGAGCTT





	1001		1050
IMPERATACYLINDRICA	CAGAACCCAT	ATTATAATAT	-----AGGT TCTTTATTTT -----ATTTT
SORGHUMHALEPENSE	CAGAACCCAT	ATTATAATAT	-----AGGT TCTTTATTTT -----ATTTT
PANICUMLANIPES	TAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTT -----ATTTT
PANICUMSTAFFIANUM	TAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTT -----ATTTT
KARROOCHLOAPURPUREA6241	CCGAACCTAT	ATCATAATAT	-----AGGT TCTTTATTTT ---TTTTTT
CHAETOBROMUSINVOLUCRATES6284	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTT -----TTTTT
Cynodondactylon7480	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Cynodondactylon	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Eleusinecoracana	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Eleusineindica	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Chlorisvirgata6616	CAGAACCCAT	ACGATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Chlorisvirgata7498	CAGAACCCAT	ACGATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Eustachyspaspaloides5597	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Harpochloafalx6530	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Harpochloafalx5118	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Harpochloafalx6670	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Rendliaaltera5133	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Leptochloafusca7622	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Enteropogonmacrostachyus9339	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Dactylocteniumaegyptium9326	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTYATTTA -----TTTTT
Tragusberteronianus6620	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTT -----TTTTT
Tragusracemosus7108	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTT -----TTTTT
Traguskoelerioides6729	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Polevansiarigida7621	CAGAACCCAT	ACCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Perotispatens9323	CAGAACCCAT	ACCATAATAT	AGTATAGGT TCCTTATTTA -----TTTTT
Eragrostiscurvula6529	CAGAACCCMT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Eragrostisobtusa6623	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Eragrostisobtusa6732	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN NNNNNNNNNN NNNNNNNNNN
Eragrostisechinochloidea7485	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Eragrostispseudobtusa7508	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Eragrostislehmanniana20	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Eragrostischloromelas6947	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Eragrostischloromelas7474	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Eragrostisracemosa6533	CAGAACCCGT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Eragrostisbiflora6705	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Eragrostissuperba7470	CAGAACCCCT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Eragrostiscapensis6551	TAGAACCCAT	ATCATAATAT	-----AGGT TCCTTATTTA -----TTTTT
Eragrostiscapensis6532	TAGAACCCAT	ATCATAATAT	-----AGGT TCCTTATTTA -----TTTTT
Pogonarthriasquarrosa7483	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Pogonarthriasquarrosa7504	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Catalepisgracilis6962	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Catalepisgracilis6653	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Cladoraphiscyperoides5704	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Cladoraphisspinosa6335	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Cladoraphiscyperoides4894	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----ATTTT
Fingerhuthiaafricana7507	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Entoplocamiaaristulata9353	CAGAACCC-T	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Enneapogonscoparius18	CAGAACCCAT	ATCATAATAT	ATAT--AGGT TCTTTATTTA --TTTTTTTT
Enneapogoncenchroides7506	CAGAACCCAT	ATCATAATAT	AT---AGGT TCTTTATTTA TTTTTTTTTT
Enneapogonscoparius7487	CAGAACCCAT	ATCATAATAT	ATAT--AGGT TCTTTATTTA --TTTTTTTT
Schmidtiaappophoroides6334	CAGAACCCAT	ATCATAATAT	ATAT--AGGT TCTTTATTTT -----TTTTT
Schmidtiaalihariensis7490	CAGAACCCAT	ATCATAATAT	ATATATAGGT TCTTTATTTA -----TTTTT
Schmidtiaalihariensis7486	CAGAACCCAT	ATCATAATAT	ATATATAGGT TCTTTATTTA -----TTTTT
Sporobolusafricanus7476	CAGAACCCGT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Sporobolusconsimilis9354	CAGAACCCAT	ATCATAATAT	-----AGGT TCTTTATTTA -----TTTTT
Sporobolusfimbriatus9328	CAGAACCCAT	ATCATAATAT	-----AGGT TTTTTATTTA -----TTTTT





IMPERATACYLINDRICA	AATCCTTCAA	TTCATTGTTT	TCGAGAT---	--CTTTT-AA	AAAGTGGA--
SORGHUMHALEPENSE	AATCCTTCAA	TTCATTGTTT	TCGAGAT---	--CTTTT-AA	AAAGTGGA--
PANICUMLANIPES	AATCCTTCAA	TTCATTGTTT	T-GAGAT---	--CTTCA-AA	AAAGTGGA--
PANICUMSTAFFIANUM	AATCCTTCAA	TTCATTGTTT	T-GAGAT---	--CTTCA-AA	AAAGTGGA--
KARROOCHLOAPURPUREA6241	AATCCTTCAA	TTCATT-TTT	TTGAGAT---	--CCTTT-AA	AA-GTGA--
CHAETOBROMUSINVOLUCRATES6284	AATCCTTCAA	TTCATT-TTT	TTGAGAT---	--CTTTT-AA	AAAGCGGA--
Cynodondactylon7480	AACCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAATAGGA--
Cynodondactylon	AACCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAATAGGA--
Eleusinecoracana	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-TA	AAATAGGA--
Eleusineindica	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-TA	AAATAGGA--
Chlorisvirgata6616	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--ATTT--AA	AAATAGGA--
Chlorisvirgata7498	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--ATTT--AA	AAATAGGA--
Eustachyspaspaloides5597	AACCCTTCAA	TTCATTGTTT	TTGAGATNNN	NNNNNNNNNN	NNNNNNNNNN
Harpochloafalx6530	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--ATTTT-TA	AAATAGGA--
Harpochloafalx5118	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--ATTTT-AA	AAATAGGA--
Harpochloafalx6670	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--ATTTT-AA	AAATAGGA--
Rendliaaltera5133	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAATAAGA--
Leptochloafusca7622	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--ATTTA-AA	AAATAGGA--
Enteropogonmacrostachyus9339	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTA-AA	AAATAGGA--
Dactylocteniumaegyptium9326	AATCCTTCAA	TTCATTGTTT	TTGAGATT--	--AAAAA-AA	AAATAGGA--
Tragusberteronianus6620	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAATAAGA--
Tragusracemosus7108	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAATAAGA--
Traguskoelerioides6729	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAATAAGA--
Polevansiarigida7621	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAATAAGA--
Perotispatens9323	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--ATTTT-TA	AAATAGGA--
Eragrostiscurvula6529	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Eragrostisobtusa6623	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Eragrostisobtusa6732	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Eragrostisechinochloidea7485	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Eragrostispseudobtusa7508	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Eragrostislehmanniana20	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Eragrostischloromelas6947	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Eragrostischloromelas7474	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Eragrostisracemosa6533	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Eragrostisbiflora6705	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Eragrostissuperba7470	AATCCTTCAA	TTCATTGTTT	TTGAGATCCT	TTATTTT-TA	AAAGAGGA--
Eragrostiscapensis6551	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CCTTT-AA	AAAGAGGA--
Eragrostiscapensis6532	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CCTTT-AA	AAAGAGGA--
Pogonarthriasquarrosa7483	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Pogonarthriasquarrosa7504	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Catalepisgracilis6962	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Catalepisgracilis6653	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Cladoraphiscyperoides5704	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Cladoraphisspinosa6335	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Cladoraphiscyperoides4894	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Fingerhuthiaafricana7507	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Entoplocamiae aristulata9353	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Enneapogonscoparius18	GATCCTTCAT	TTCATTGTTT	TTGAGAT---	--CTTTTAA	AAAGAGGATT
Enneapogoncenchroides7506	GATCCTTCAT	TTCATTGTTT	TTGAGAT---	--CTTTTAA	AAAGAGGATT
Enneapogonscoparius7487	GATCCTTCAT	TTCATTGTTT	TTGAGAT---	--CTTTTAA	AAAGAGGATT
Schmidtia pappophoroides6334	AATCCTTCAT	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGATT
Schmidtia kalihariensis7490	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Schmidtia kalihariensis7486	AATCCTTCAA	TTCATTGTTT	TTGAGAT---	--CTTTT-AA	AAAGAGGA--
Sporobolusafricanus7476	AATCCTTCAA	TTCATTGTTT	TTGAGATTTT	TTATTTT-AA	AAAGAAGA--
Sporobolus consimilis9354	-----	-----	-----	-----	AAAGAAGA--
Sporobolus fimbriatus9328	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN



	1251			1300	
IMPERATACYLINDRICA	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
SORGHUMHALEPENSE	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
PANICUMLANIPES	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
PANICUMSTAFFIANUM	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
KARROOCHLOAPURPUREA6241	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
CHAETOBROMUSINVOLUCRATES6284	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Cynodondactylon7480	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Cynodondactylon	TGACAACAAT	GAAATTTCTA	GTA AAAAGGNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	TGACAACAAT	GAAATTTCTA	GTA AAAAGGNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	TGACAACAAT	GAAATTTCTA	GTA AAAAGGNN	NNNNNNNNNN	NNNNNNNNNN
Chlorisvirgata6616	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Chlorisvirgata7498	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Eustachyspaspaloides5597	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Harpochloafalx6530	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTGGA	CTTTTATAAGT
Harpochloafalx5118	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTTATAAGT
Harpochloafalx6670	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTTATAAGT
Rendliaaltera5133	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGG	CTTTATAAGT
Leptochloafusca7622	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Enteropogonmacrostachyus9339	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTCTAAGT
Dactylocteniumaegyptium9326	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Tragusberteronianus6620	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Tragusracemosus7108	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Traguskoelerioides6729	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Polevansiarigida7621	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Perotispatens9323	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGC	CTTTATAAGT
Eragrostiscurvula6529	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Eragrostisobtusa6623	TGACAACAAT	GAAATTTCTA	GTA AAAAGGNN	NNNNNGTCGA	CTTTATAAGT
Eragrostisobtusa6732	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Eragrostisechinochloidea7485	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTATAAAGT
Eragrostispseudobtusa7508	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Eragrostislehmanniana20	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Eragrostischloromelas6947	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Eragrostischloromelas7474	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Eragrostisracemosa6533	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Eragrostisbiflora6705	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Eragrostissuaveolens7470	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Eragrostiscapensis6551	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Eragrostiscapensis6532	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Pogonarthriasquarrosa7483	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Pogonarthriasquarrosa7504	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Catalepisgracilis6962	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Catalepisgracilis6653	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Cladoraphiscyperoides5704	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Cladoraphisspinosa6335	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Cladoraphiscyperoides4894	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Fingerhuthiaafricana7507	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Entoplocamiaaristulata9353	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Enneapogonscoparius18	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Enneapogoncenchroides7506	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Enneapogonscoparius7487	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Schmidtiapappophoroides6334	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Schmidtiakalihariensis7490	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Schmidtiakalihariensis7486	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Sporobolusafricanus7476	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Sporobolusconsimilis9354	TGACAACAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT
Sporobolusfimbriatus9328	NNNNNNCAAT	GAAATTTCTA	GTA AAAAGGAA	AATCCGTCGA	CTTTATAAGT

IMPERATACYLINDRICA	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCTTTTA	TTCCTAACC
SORGHUMHALEPENSE	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCTTTTA	TTCCTAACC
PANICUMLANIPES	CGTGAGNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMSTAPFIANUM	CGTGAGNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
KARROOCHLOAPURPUREA6241	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCTTTTA	TTCCTAACC
CHAETOBROMUSINVOLUCRATES6284	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCGTTTA	TTCCTAACC
Cynodondactylon7480	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Cynodondactylon	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Chlorisvirgata6616	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Chlorisvirgata7498	CGTGAGGGTT	CAAGTCCCTT	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Eustachyspaspaloides5597	NNNNNNNNNN	NNNNNNNNNN	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Harpochloafalx6530	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Harpochloafalx5118	CGTGAGGGTT	CAAGTCCCTT	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Harpochloafalx6670	CGGGAGGGTT	CAAGTCCCTT	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Rendliaaltera5133	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Leptochloafusca7622	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Enteropogonmacrostachyus9339	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Dactylocteniumaegyptium9326	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Tragusberteronianus6620	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Tragusracemosus7108	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Traguskoelerioides6729	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Polevansiarigida7621	CGTGAGGGTT	CAAGTCCCTT	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Perotispatens9323	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Eragrostiscurvula6529	TGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Eragrostisobtusa6623	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Eragrostisobtusa6732	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Eragrostisechinochloidea7485	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Eragrostispseudobtusa7508	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Eragrostislehmanniana20	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Eragrostischloromelas6947	TGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Eragrostischloromelas7474	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Eragrostisracemosa6533	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Eragrostisbiflora6705	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Eragrostissuperba7470	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Eragrostiscapensis6551	CATGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Eragrostiscapensis6532	CATGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Pogonarthriasquarrosa7483	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Pogonarthriasquarrosa7504	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Catalepisgracilis6962	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Catalepisgracilis6653	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Cladoraphiscyperoides5704	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Cladoraphis spinosa6335	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Cladoraphiscyperoides4894	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Fingerhuthiaafricana7507	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Entoplocamia aristulata9353	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Enneapogon scoparius18	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Enneapogon cenchroides7506	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Enneapogon scoparius7487	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Schmidtia papporoides6334	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Schmidtia kalihariensis7490	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Schmidtia kalihariensis7486	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Sporobolus africanus7476	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Sporobolus consimilis9354	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCTCCTTTA	TTCCTAACC
Sporobolus fimbriatus9328	CGTGAGGGTT	CAAGTCCCTC	TATCCCCAAA	CCCCCTTTA	TTCCTAACC





IMPERATACYLINDRICA	---AAGATTC	ACTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
SORGHUMHALEPENSE	---AAGATTC	ACTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
PANICUMLANIPES	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMSTAFFIANUM	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
KARROOCHLOAPURPUREA6241	---AAGATTC	ATTAGCT---	-TTCTCATT	TATT-----	C	TTTCACAAAG
CHAETOBROMUSINVOLUCRATES6284	---AAGATTC	ATTAGCT---	-TTCTCATT	TATT-----	C	TTTCACAAAG
Cynodondactylon7480	GACAAGATTC	ATTAGCTTGC	TTTCTCATT	TACT-----	C	TTTCACAAAG
Cynodondactylon	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Chlorisvirgata6616	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Chlorisvirgata7498	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Eustachyspaspaloides5597	GACAAGATTC	ATTAGTT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Harpochloafalx6530	--CAAGATTC	ATTAGTT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Harpochloafalx5118	--CAAGATTC	ATTAGTT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Harpochloafalx6670	--CAAGATTC	ATTAGTT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Rendliaaltera5133	GACAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Leptochloafusca7622	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Enteropogonmacrostachyus9339	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Dactylocteniumaegyptium9326	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Tragusberteronianus6620	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Tragusracemosus7108	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Traguskoelerioides6729	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Polevansiarigida7621	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Perotispatens9323	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Eragrostiscurvula6529	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Eragrostisobtusa6623	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACTTTACTC		TTTCATAAAG
Eragrostisobtusa6732	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACTTTACTC		TTTCATAAAG
Eragrostisechinochloidea7485	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACTCTACTC		TTTCATAAAG
Eragrostispseudobtusa7508	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACTTTACTC		TTTCATAAAG
Eragrostislehmanniana20	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Eragrostischloromelas6947	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Eragrostischloromelas7474	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Eragrostisracemosa6533	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Eragrostisbiflora6705	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Eragrostisuperba7470	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Eragrostiscapensis6551	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Eragrostiscapensis6532	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Pogonarthriasquarrosa7483	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Pogonarthriasquarrosa7504	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Catalepisgracilis6962	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Catalepisgracilis6653	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Cladoraphiscyperoides5704	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Cladoraphis spinosa6335	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Cladoraphiscyperoides4894	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Fingerhuthiaafricana7507	--CAAGATTC	ATTAGTT---	-TTCTCATT	TACT-----	C	TTTCATAAAA
Entoplocamiaaristulata9353	--CAAGATTC	ATTAGTT---	-TTCTCATT	TACT-----	C	TTTCATAAAG
Enneapogon scoparius18	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Enneapogon cenchroides7506	--CAAGATTC	ATTAGCT---	-TTCTCATT	TATT-----	C	TTTCACAAAG
Enneapogon scoparius7487	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	T	TTTCACAAAG
Schmidtiapappophoroides6334	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAT
Schmidtiakalihariensis7490	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Schmidtiakalihariensis7486	--CAAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Sporobolusafricanus7476	-----	-----	-----	-----	-----	-----
Sporobolus consimilis9354	-ACAAAATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG
Sporobolus fimbriatus9328	--CGAGATTC	ATTAGCT---	-TTCTCATT	TACT-----	C	TTTCACAAAG





IMPERATACYLINDRICA	GCAAGGA---A	TCTCGATTAT	TAATTCGATT	TTTT-----
SORGHUMHALEPENSE	GCAAGGA---A	TCTCGATTAT	TAATTCGATT	TTTT-----
PANICUMLANIPES	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMSTAFFIANUM	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
KARROOCHLOAPURPUREA6241	GAAAGAA---A	TCCCCATTAG	TAATTCGATT	TT-----
CHAETOBROMUSINVOLUCRATES6284	GCAAGAA---A	TCCCCATTAT	TAATTCGATT	CT-----
Cynodondactylon7480	GCAAAAA---A	TCTCAATTAT	TAATTA AATT	TA-----
Cynodondactylon	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Chlorisvirgata6616	ATAAAAA---A	TCTCAATTAT	TAATTA AATT	TC-----
Chlorisvirgata7498	ATAAAAA---A	TCTCAATTAT	TAATTA AATT	TC-----
Eustachyspaspaloides5597	GCAAAAA---A	TCTCAATTAT	TAATTA AATT	TA-----
Harpochloafalx6530	GCAAAAA---A	TCTCAATTAT	TAATGAAATT	TA-----
Harpochloafalx5118	GCAAAAA---A	TCTCAATTAT	TAATGAAATT	TT-----
Harpochloafalx6670	GCAAAAA---A	TCTCAATTAT	TAATGAAATT	TA-----
Rendliaaltera5133	GCAARRA---A	TCTCAATTAT	TAATTA AATT	TATAAGTWTT
Leptochloafusca7622	GCAAAAA---A	TCTCAATTAT	TATTTA AATT	TA-----
Enteropogonmacrostachyus9339	GCAAAAATAG	AGTATCCGCA	TCTCAATTAT	TAATGAAATT
Dactylocteniumaegyptium9326	ACAAAAA---A	TCTCAATTAT	TAATTA AATT	TA-----
Tragusberteronianus6620	GCAAAGA---A	TCTCAATTAT	TAATTA AATT	AA-----
Tragusracemosus7108	GCAAAGA---A	TCTCAATTAT	TAATTA AATT	AA-----
Traguskoelerioides6729	TCAAAGA---A	TCTCAATTAT	TAATTA AATT	AA-----
Polevansiarigida7621	GCAAAGA---A	TCTCAATTAT	TAATTA AATT	TATAAGAATT
Perotispatens9323	GAAAAAA---A	TCTCAATTAT	TAATGAAATT	AA-----
Eragrostiscurvula6529	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Eragrostisobtusa6623	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Eragrostisobtusa6732	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Eragrostisechinochloidea7485	GCAAGGA---A	TCTCAATTAT	TACTTA AATT	AT-----
Eragrostispseudobtusa7508	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Eragrostislehmanniana20	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Eragrostischloromelas6947	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Eragrostischloromelas7474	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Eragrostisracemosa6533	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Eragrostisbiflora6705	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Eragrostis superba7470	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Eragrostiscapensis6551	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Eragrostiscapensis6532	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Pogonarthriasquarrosa7483	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Pogonarthriasquarrosa7504	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Catalepisgracilis6962	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Catalepisgracilis6653	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	AT-----
Cladoraphiscyperoides5704	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	ATA-----
Cladoraphis spinosa6335	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	ATA-----
Cladoraphiscyperoides4894	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	ATA-----
Fingerhuthiaafricana7507	GC--GGA---A	TCTCAATTAT	TAATTA AATT	TT-----
Entoplocamiaaristulata9353	GC--GGA---A	TCTCAATTAT	TAATTA AATT	TT-----
Enneapogonscoparius18	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	TTT-----
Enneapogoncenchroides7506	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	TT-----
Enneapogonscoparius7487	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	TTT-----
Schmidtia pappophoroides6334	ACAAGGA---A	TCTCAATTAT	TAATTA AATT	TT-----
Schmidtia kalihariensis7490	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	TT-----
Schmidtia kalihariensis7486	GCAAGGA---A	TCTCAATTAT	TAATTA AATT	TT-----
Sporobolusafricanus7476	GCAAAGA---A	TCTTAATTAT	TAATTA AATT	TT-----
Sporobolus consimilis9354	GCAAAGA---A	TCTCAATTAT	TAATTA AATT	TT-----
Sporobolus fimbriatus9328	ACAAAGA---A	TTTAAATTAT	TAATTA AATT	TG-----

IMPERATACYLINDRICA	-----TAA	GTATTA----	-----TTA	AGTA-----
SORGHUMHALEPENSE	-----TAA	GTATTA----	-----TTA	AGTA-----
PANICUMLANIPES	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMSTAPFIANUM	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
KARROOCHLOAPURPUREA6241	-----TAA	GTATTTTTTT	ATTAATATTA	AGTA-----
CHAETOBROMUSINVOLUCRATES6284	-----TAA	TTATTTATTT	ATTAA-----	-----
Cynodondactylon7480	-----TAA	GTATTA----	-----TTA	AGTA-----
Cynodondactylon	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Chlorisvirgata6616	-----TAA	GTATTA----	-----TTA	AGTA-----
Chlorisvirgata7498	-----TAA	GTATTA----	-----TTA	AGTA-----
Eustachyspaspaloides5597	-----TAA	GTATTA----	-----TTA	AGTA-----
Harpochloafalx6530	-----TAA	GTATTA----	-----TTA	AGTA-----
Harpochloafalx5118	-----TAA	GTATTA----	-----TTA	AGTA-----
Harpochloafalx6670	-----TAA	GTATTA----	-----TTA	AGTA-----
Rendliaaltera5133	AT-----	-----TAA	GTATTA----	-----TTA
Leptochloafusca7622	-----TAA	GTATTA----	-----TTA	AGTA-----
Enteropogonmacrostachyus9339	-----TAA	GTATTA----	-----TTA	AGTA-----
Dactylocteniumaegyptium9326	-----TAA	GTATTA----	-----TTA	AGTA-----
Tragusberteronianus6620	-----TAA	GTATTA----	-----TTA	AGTA-----
Tragusracemosus7108	-----TAA	GTATTA----	-----TTA	AGTA-----
Traguskoelerioides6729	-----TAA	GTATTA----	-----TTA	AGTA-----
Polevansiarigida7621	TATTACAATT	TATAAGATAA	GTATTA----	-----TTA
Perotispatens9323	-----TAA	GTATTA----	-----TTA	AGTA-----
Eragrostiscurvula6529	-----TAA	GTATTA----	-----TTA	AGTA-----
Eragrostisobtusa6623	-----TAA	GTATTA----	-----TTA	AGTA-----
Eragrostisobtusa6732	-----TAA	GTATTA----	-----TTA	AGTA-----
Eragrostisechinochloidea7485	-----TAA	GTATTA----	-----TTA	AGTA-----
Eragrostispseudobtusa7508	-----TAA	GTATTA----	-----TTA	AGTA-----
Eragrostislehmanniana20	-----TAA	GTATTA----	-----TTA	AGTA-----
Eragrostischloromelas6947	-----TAA	GTATTA----	-----TTA	AGTA-----
Eragrostischloromelas7474	-----TAA	TTATTA----	-----TTA	AGTA-----
Eragrostisracemosa6533	-----TAA	GTATTA----	-----TTA	AGTA-----
Eragrostisbiflora6705	-----TAA	GTATTA----	-----TTA	AGTA-----
Eragrostisuperba7470	-----TAA	GTATTA----	-----TTA	AGTA-----
Eragrostiscapensis6551	-----TAA	GTATTA----	-----TTA	AGTA-----
Eragrostiscapensis6532	-----TAA	GTATTA----	-----TTA	AGTA-----
Pogonarthriasquarrosa7483	-----TAA	GTATTA----	-----TTA	AGTA-----
Pogonarthriasquarrosa7504	-----TAA	GTATTA----	-----TTA	AGTA-----
Catalepisgracilis6962	-----TAA	TTATTA----	-----TTA	AGTA-----
Catalepisgracilis6653	-----TAA	TTATTA----	-----TTA	AGTA-----
Cladoraphiscyperoides5704	-----TAA	GTATTAT----	-----TTA	AGTA-----
Cladoraphisspinosa6335	-----TAA	GTATTAT----	-----TTA	AGTA-----
Cladoraphiscyperoides4894	-----TAA	GTATTAT----	-----TTA	AGTATT--AT
Fingerhuthiaafricana7507	-----TAA	GTATTA----	-----TTA	AGTA-----
Entoplocamiaaristulata9353	-----TAA	GTATTA----	-----TTA	AGTAATCCAT
Enneapogonscoparius18	-----GAA	GTATTA----	TTAATATTA	AGTA-----
Enneapogoncenchroides7506	-----GAA	GTATTA----	TTAATATTA	AGTA-----
Enneapogonscoparius7487	-----GAA	GTATTA----	TTAATATTA	AGTA-----
Schmidtiaappophoroides6334	-----GAA	GTATTA----	TTAATATTA	AGTA-----
Schmidtiaalihariensis7490	-----GAA	GTATTA----	TTAATATTA	AGTA-----
Schmidtiaalihariensis7486	-----GAA	GTATTA----	TTAATATTA	AGTA-----
Sporobolusafricanus7476	-----TAA	GTATTA----	-----TTA	AGTA-----
Sporobolusconsimilis9354	-----TAA	GTATTA----	-----TTA	AGTA-----
Sporobolusfimbriatus9328	-----GAA	TTATTA----	-----TTA	AGTA-----

IMPERATACYLINDRICA	-----AGCC	ATCCACAATG	CATAGGAC--	-----TACC
SORGHUMHALEPENSE	-----AGCC	ATCCACAATG	CATAGGAC--	-----TACC
PANICUMLANIPES	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMSTAPFIANUM	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
KARROOCHLOAPURPUREA6241	-----AGCC	ATCCACAATG	CATAGGAC--	-----TACT
CHAETOBROMUSINVOLUCRATES6284	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Cynodondactylon7480	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Cynodondactylon	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Chlorisvirgata6616	-----AGCC	ATCCACAATG	CATAGGAC--	-----TACC
Chlorisvirgata7498	-----AGCC	ATCCACAATG	CATAGGAC--	-----TACC
Eustachyaspaspaloides5597	-----AGCC	GTCCACAATG	CATAGGAC--	-----TATC
Harpochloafalx6530	-----AGCC	ATCTACAATG	CATAGGAC--	-----TACC
Harpochloafalx5118	-----AGCC	ATCTACAATG	CATAGGAC--	-----TACC
Harpochloafalx6670	-----AGCC	ATCTACAATG	CATAGGAC--	-----TACC
Rendliaaltera5133	-----AGCC	ATCCACAATG	CATAGGAC--	-----TAGC
Leptochloafusca7622	-----AGCC	ATCCACAATG	TATAGGAC--	-----TATC
Enteropogonmacrostachyus9339	-----AGCC	ATCCACAATG	TATAGGAC--	-----TACT
Dactylocteniumaegyptium9326	-----AGTC	ATCCACAATG	CATAGGAC--	-----TACC
Tragusberteronianus6620	-----AGCC	ATCCACAATG	CATAGGAC--	-----TACC
Tragusracemosus7108	-----AGCC	ATCCACAATG	CATAGGAC--	-----TACC
Traguskoelerioides6729	-----AGCC	ATCCACAATG	CATAGGAC--	-----TACC
Polevansiarigida7621	-----AGCC	ATCCACAATG	CATAGGAC--	-----TACC
Perotispatens9323	-----AGCC	GTCCACAATG	CATAGGAC--	-----TATC
Eragrostiscurvula6529	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Eragrostisobtusa6623	-----AGCC	ATCCACAATG	CATCGGAC--	-----TATC
Eragrostisobtusa6732	-----AGCC	ATCCACAATG	CATCGGAC--	-----TATC
Eragrostisechinochloidea7485	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Eragrostispseudobtusa7508	-----AGCC	ATCCACAATG	CATCGGAC--	-----TATC
Eragrostislehmanniana20	-----AGCC	ATCCACAATG	CATAGGAC--	-----TACC
Eragrostischloromelas6947	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Eragrostischloromelas7474	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Eragrostisracemosa6533	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Eragrostisbiflora6705	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Eragrostissuperba7470	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Eragrostiscapensis6551	-----AGCC	GTCCACAATG	CATAGGAC--	-----TATC
Eragrostiscapensis6532	-----AGCC	GTCCACAATG	CATAGGAC--	-----TATC
Pogonarthriasquarrosa7483	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Pogonarthriasquarrosa7504	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Catalepisgracilis6962	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Catalepisgracilis6653	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Cladoraphiscyperoides5704	-----AGCC	ATCCACAATG	CATAGTAC--	-----TATC
Cladoraphisspinosa6335	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Cladoraphiscyperoides4894	T-----	TAAGTAAGCC	ATCCACAATG	CATAGGAC--
Fingerhuthiaafricana7507	-----ATCC	ATCCACAATG	CATAGGAC--	-----TATC
Entoplocamiaaristulata9353	CCACAATGCA	TAAGTAATCC	ATCCACAATG	CATAGGAC--
Enneapogonscoparius18	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Enneapogoncenchroides7506	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Enneapogonscoparius7487	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Schmidtiaappophoroides6334	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Schmidtiaalihariensis7490	-----AGCC	ATCCACAATG	CATAGGACCA	TAGGACTATC
Schmidtiaalihariensis7486	-----AGCC	ATCCACAATG	CATAGGACCA	TAGGACTATC
Sporobolusafricanus7476	-----AGCC	ATCCACAATG	CATAGGAC--	-----TATC
Sporobolusconsimilis9354	-----AGCC	ATCCACAATG	CATAGGAT--	-----TATC
Sporobolusfimbriatus9328	-----AGCC	ATCCACAATG	CATAGGAC--	-----TACC



IMPERATACYLINDRICA	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
SORGHUMHALEPENSE	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
PANICUMLANIPES	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
PANICUMSTAFFIANUM	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
KARROOCHLOAPURPUREA6241	--AACTGATT	TTTTAATCCC	TTTAATTGAC	AATTGACATA	GATGCAAATA
CHAETOBROMUSINVOLUCRATES6284	--AATTGATT	TTTTAATCCC	TTTAATT---	----GACATA	GATGCAAATA
Cynodondactylon7480	---ATTGATT	TTTTAGTCTC	TTTAATT---	----GACATA	GATGCAAATA
Cynodondactylon	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusinecoracana	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Eleusineindica	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN	NNNNNNNNNN
Chlorisvirgata6616	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Chlorisvirgata7498	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Eustachyaspaloides5597	---ATTGATT	TTTTAGTCTC	TTTAATT---	----GACATA	GATGCAAATA
Harpochloafalx6530	TTTATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	AATGCAAATA
Harpochloafalx5118	TTTATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	AATGCAAATA
Harpochloafalx6670	TTTATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	AATGCAAATA
Rendliaaltera5133	---ATTAATT	TTTTAGTCTC	TTTAATT---	----GACATA	GATGCAAATA
Leptochloafusca7622	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Enteropogonmacrostachyus9339	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Dactylocteniumaegyptium9326	---TTTGATT	TTTTAGTCCC	ATTAATT---	----GACATA	AATGGAATA
Tragusberteronianus6620	---ATTGATT	TTT-AGTCTC	TTTAATT---	----GACATA	GATGCAAATA
Tragusracemosus7108	---ATTGATT	TTT-AGTCTC	TTTAATT---	----GACATA	GATGCAAATA
Traguskoelerioides729	---ATTGATT	TTT-AGTCTC	TTTAATT---	----GACATA	GATGCAAATA
Polevansiarigida7621	---ATTGATT	TTT-AGTCTC	TTTAATT---	----GACATA	GATGCAAATA
Perotispatens9323	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Eragrostiscurvula6529	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Eragrostisobtusa6623	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Eragrostisobtusa6732	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Eragrostisechinochloidea7485	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Eragrostispseudobtusa7508	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Eragrostislehmanniana20	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Eragrostischloromelas6947	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Eragrostischloromelas7474	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Eragrostisracemosa6533	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Eragrostisbiflora6705	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Eragrostisuperba7470	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Eragrostiscapensis6551	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Eragrostiscapensis6532	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Pogonarthriasquarrosa7483	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Pogonarthriasquarrosa7504	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Catalepisgracilis6962	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATCCAAATA
Catalepisgracilis6653	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATCCAAATA
Cladoraphiscyperoides5704	----TGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Cladoraphisspinosa6335	----TGATT	TTTT- ----C	TTTAATT---	----GACATA	GATGCAAATA
Cladoraphiscyperoides4894	----TGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Fingerhuthiaafricana7507	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Entoplocamiaaristulata9353	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGAAATA
Enneapogonscoparius18	---ATTGATC	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Enneapogoncenchroides7506	---ATTGAT-	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Enneapogonscoparius7487	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Schmidtiaappophoroides6334	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Schmidtialihariensis7490	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Schmidtialihariensis7486	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Sporobolusafricanus7476	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Sporobolusconsimilis9354	---ATTGATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA
Sporobolusfimbriatus9328	---ATTTATT	TTTTAGTCCC	TTTAATT---	----GACATA	GATGCAAATA







IMPERATACYLINDRICA	001-1000001100000000101000100????????????????????????????????
SORGHUMHALEPENSE	001-1000001100000000100000100????????????????????????????????
PANICUMLANIPES	001-1000001100000000001000101--001--000000000000000010-000
PANICUMSTAFFIANUM	001-1000001100000000001000101--001--000000000000000010-000
KARROOCHLOAPURPUREA6241	001--1000011000000001000011000100010100000001000100010001000
CHAETOBROMUSINVOLUCRATES6284	001-00000011000000001001-010001000001000000100010000010-000
Cynodondactylon7480	101-00010011000001-01000101000100000100000000000100000000001
Cynodondactylon	101-00010011000001-01000101000100000100000000000100000000001
Eleusinecoracana	111-00000011010001-01000101000100000-100000000001010-0000000
Eleusineindica	111-00000011010001-01000101000100000-100000000001010-0000000
Chlorisvirgata6616	101-00000011000000101000101000100000100000000000100000000000
Chlorisvirgata7498	101-00000011000000101000101000100000100000000000100000000000
Eustachyspaspaloides5597	101-000100110001-0-01000100001-0000010000000000010000000-10-
Harpochloafalx6530	101-000010110001-0-01000100100-10000--10-00-0000100000-1--0-
Harpochloafalx5118	101-000010110001-0-01000100100-10000--10-00-0000100000-1--0-
Harpochloafalx6670	101-000010110001-0-01000100100-10000--10-00-0000100000-1--0-
Rendliaaltera5133	101-000100110001-0-01000101000100000100000000000100000000001
Leptochloafusca7622	101-000100110001-0-0100010100010000010000000000010000000-10-
Enteropogonmacrostachyus9339	101-000100110001-0-0100010100010000010000000000000000000-10-
Dactylocteniumaegyptium9326	101-000100110001-0-01000101000100001-0000000000010-1-----0-
Tragusberteronianus6620	101-000100110001-0-0100010100010000010000000000010000000-10-
Tragusracemosus7108	101-000100110001-0-01??
Traguskoelerioides6729	101-000100110001-0-0100010100010000010000000000010000000-10-
Polevansiarigida7621	101-000100110001-0-0110010100010000010000000000010000000-10-
Perotispatens9323	001-001100110001-0-01000101000000000000000000000-1000000-10-
Eragrostiscurvula6529	101-0001001100000001000101000100000000000000000010000000000-
Eragrostisobtusa6623	101-00010011000000001000101000100000---1--0-000010000000000-
Eragrostisobtusa6732	101-00010011000000001000101000100000---1--0-000010000000000-
Eragrostisechinochloidea7485	101-00010011000000001000101000100000---1--0-000010000000000-
Eragrostispseudobtusa7508	101-00010011000000001000101000100000---1--0-000010000000000-
Eragrostislehmanniana20	101-0001001100000000100010100010000000000000000010000000000-
Eragrostischloromelas6947	101-000100110000000010001010001000000000000000000001000000000-
Eragrostischloromelas7474	101-000100110000000010001010001010000000100-000010000000000-
Eragrostisracemosa6533	101-000101110000000010001010001000000000100-000010????????????
Eragrostisbiflora6705	101-000100110000000010001010001010000000100-000010000000000-
Eragrostissuperba7470	101-00010011000000001000101000100000-1000000000010000000000-
Eragrostiscapensis6551	101-0001001100000000100010100010000000000-10-000010000000000-
Eragrostiscapensis6532	101-0001001100000000110001010001000000000100-001010000000--1-
Pogonarthriasquarrosa7483	101-000100110000000010001010001000000000--1-000010000000000-
Pogonarthriasquarrosa7504	101-000100110000000010001010001000000000--1-000010000000000-
Catalepisgracilis6962	101-000100110000000010001010001010000000100-000010000000000-
Catalepisgracilis6653	101-000100110000000010001010001010000000100-000010000000000-
Cladoraphiscyperoides5704	101-000100110000000010001010001010000000-10-000010000000000-
Cladoraphis spinosa6335	101-000100110000000010001010001010000000-10-000010000000000-
Cladoraphiscyperoides4894	101-000100110000000010001010001010000000-10-000010000000000-
Fingerhuthiaafricana7507	101-000100110000000010001010001000100000-10-000010000000000-
Entoplocamia aristulata9353	101-000100110000000010001010001000100010000-10-000010000000000-
Enneapogon scoparius18	101-000100110000000010001010001000000000001000110000000000-
Enneapogon cenchroides7506	101-000100110000000010001010001000000000001000110000000000-
Enneapogon scoparius7487	101-000100110000000010001010001000000000001000110000000000-
Schmidtia pappophoroides6334	101-000100110000000010001010001000000000001000010000000000-
Schmidtia kalihariensis7490	101-000100110000000010001010001000000000001000010000000000-
Schmidtia kalihariensis7486	101-000100110000000010001010001000000000001000010000000000-
Sporobolus africanus7476	000010001001000101000100010??????????????????????????????????
Sporobolus consimilis9354	00000001001000001000100010100010000001000000001-0100001000001
Sporobolus fimbriatus9328	000000010010100010001000101000100000000000000010100001000001

IMPERATACYLINDRICA	????????????????????????????????????0001000010010-10001-01-----0-
SORGHUMHALEPENSE	????????????????????????????????????00000000100-1-00001-01-----0-
PANICUMLANIPES	000000001001000000000000000000000001-0010-00001-1000101----00
PANICUMSTAFFIANUM	000000000001000000000000000000000001-0010-00001-1000101----00
KARROOCHLOAPURPUREA6241	010000000101100000010100000000000001-01-0-00001-0010101----00
CHAETOBROMUSINVOLUCRATES6284	0100000001001000000000100000000001-0010-00001-0010101----00
Cynodondactylon7480	0-1-0-000101????????001000000000100001-01-0-00001-0000100100100
Cynodondactylon	0-1-0-000101100000000000000000000001-01-0-00001-0000100100100
Eleusinecoracana	0-1-0-00010110000000000000000000000010-00001-0000100100100
Eleusineindica	0-1-0-00010110000000000000000000000010-00001-0000100100100
Chlorisvirgata6616	0-1-0-00010110100001000100000000001-01-0-00001-0000100-1--00
Chlorisvirgata7498	0-1-0-000101????????00100000000001-01-0-00001-0000100100100
Eustachyspaspaloides5597	0-1-0-000101??0100100000
Harpochloafalx6530	000001--1--0110110001-010100000100000010010--1111-1--011--
Harpochloafalx5118	0-1-0-000101????????????????????0001-1---00111-0000100000100
Harpochloafalx6670	0-1-0-00010110000000000100000000001-01-0-00001-0000100000100
Rendliaaltera5133	0-1-0-000101????????????00000000001-01-0-00001-0000100100100
Leptochloafusca7622	0-1-0-000101????????0001000000001--01-0-00001-0000100100100
Enteropogonmacrostachyus9339	0-1-0-000101????????00100000000001-01-0-00001-0000100100100
Dactylocteniumaegyptium9326	000001000101????????????????????????????????????0000100001100
Tragusberteronianus6620	00010-000101????????????????????00000000101-0-00001-0000100100100
Tragusracemosus7108	????????????????????????????????????0000101-0-00001-0000100100100
Traguskoelerioides6729	00010-000101????????????????????000000101-0-00001-0000100100100
Polevansiarigida7621	000-1-000101????????????0000000000000101-0-00001-0000100100100
Perotispatens9323	0-1-0-000101????????????????????????????????????001-0000100100100
Eragrostiscurvula6529	1-1-0-00010110000000000-1-000100001-0010-00001-0000101----00
Eragrostisobtusa6623	1-1-0-00011-10001000000000000000001-0010-00001-0000101----00
Eragrostisobtusa6732	1-1-0-00011-??
Eragrostisechinochloidea7485	1-1-0-00011-10001000000001000010001-0010-00001-000011----00
Eragrostispseudobtusa7508	1-1-0-00011-1000010000-1-00000001-0010-00001-0000101----00
Eragrostislehmanniana20	1-1-0-000101????0000001000000000000001-0010-00001-0000101----00
Eragrostischloromelas6947	1-1-0-000101100000000000000000000001-0010-00001-000011----00
Eragrostischloromelas7474	1-1-0-000101100100000000000000000001-0010-00001-0000101----00
Eragrostisracemosa6533	????????????100000000000000000000001-0010-00001-0000101----1-
Eragrostisbiflora6705	1-1-0-000101????????????0000000000001-0010-00001-0000101----00
Eragrostissuperba7470	1-1-0-00010110000000000-1-00000001-0010-00001-0000101----00
Eragrostiscapensis6551	1-1-0-0001011001000100010000100011--0010-0100010001101----00
Eragrostiscapensis6532	-0010-010101????????????????????????101--0010-0000000000101----00
Pogonarthriasquarrosa7483	1-1-0-10011-????????????0001000001--0010-00001-0000101----00
Pogonarthriasquarrosa7504	1-1-0-10011-10000000000000000000001--0010-00001-0000101----00
Catalepisgracilis6962	1-1-0-00010110000000000000100100001-0010-00001-0000101----00
Catalepisgracilis6653	1-1-0-000101????????000000100100001-0010-00001-0000101----00
Cladoraphiscyperoides5704	1-1-0-000101????????????????????????????????????00001-0000101----00
Cladoraphisspinosa6335	1-1-0-000101????????00000000000001-000000001-0000101----00
Cladoraphiscyperoides4894	1-1-0-000101????????????????????????????????????0001-0000101----00
Fingerhuthiaafricana7507	1-1-0-00010110010001000001000000101-01-0-00001-0000001----00
Entoplocamiaaristulata9353	1-1-0-000101????????????????????00001-01-0-00001-0000101----01
Enneapogonscoparius18	1-1-0-000101????????????00000000001-0010-00001-0000101----00
Enneapogoncenchroides7506	1-1-0-000101????????????????00000001-0010-00001-0000101----00
Enneapogonscoparius7487	1-1-0-000101100000000000001000000001-0010-00001-0000101----00
Schmidtiaappophoroides6334	1-1-0-000101????????????????00000001-0010-00001-0000101----00
Schmidtialahariensis7490	1-1-0-000101?000-1-0000-1-000000001-0000100001-0000101----00
Schmidtialahariensis7486	1-1-0-000101100000000000000000000001-0000100001-0000101----00
Sporobolusafricanus7476	????????????-1100010000000000000001-0010-00001-0000100100100
Sporobolusconsimilis9354	0-1-0-01010110010001000000000000001-0010-00001-0000100100100
Sporobolusfimbriatus9328	0-1-0-000101????????????????????????????????????00001-0000100100100



IMPERATACYLINDRICA	-1--01--1000010---1-000010?
SORGHUMHALEPENSE	-1--01--1000010---1-000010?
PANICUMLANIPES	????????????????????????????
PANICUMSTAFFIANUM	????????????????????????????
KARROOCHLOAPURPUREA6241	-00001--101---0---010000001
CHAETOBROMUSINVOLUCRATES6284	-0001-----010000101
Cynodondactylon7480	-1--01--10000001--1-0000101
Cynodondactylon	????????????????????????????
Eleusinecoracana	????????????????????????????
Eleusineindica	????????????????????????????
Chlorisvirgata6616	-1--01--10000000011-0000101
Chlorisvirgata7498	-1--01--1000000001-0000101
Eustachyaspaloides5597	-1--01--1000010---1-0000101
Harpochloafalx6530	-1--01--1000010---000000100
Harpochloafalx5118	-1--01--1000010---000000100
Harpochloafalx6670	-1--01--10000001--000000100
Rendliaaltera5133	11--01--10000-1---1-0000101
Leptochloafusca7622	-1--01--10000001--1-0000101
Enteropogonmacrostachyus9339	-1--01--1000010---1-0000101
Dactylocteniumaegyptium9326	-1--01--1001--0---1-0000111
Tragusberteronianus6620	-1--01--1001--0---1-0010101
Tragusracemosus7108	-1--01--1001--0---1-0010101
Traguskoelerioides6729	-1--01--1001--0---1-0010101
Polevansiarigida7621	01--01--10001-0---1-0010101
Perotispatens9323	-1--01--10000000011-0000101
Eragrostiscurvula6529	-1--01--1001--0---1-0000101
Eragrostisobtusa6623	-1--01--1001--0---1-0000101
Eragrostisobtusa6732	-1--01--1001--0---1-0000101
Eragrostisechinochloidea7485	-1--01--1001--0---1-0000101
Eragrostispsuedobtusa7508	-1--01--1001--0---1-0000101
Eragrostislehmanniana20	-1--01--1001--0---1-0000101
Eragrostischloromelas6947	-1--01--1001--0---1-0000101
Eragrostischloromelas7474	-1--01--101---0---1-0000101
Eragrostisracemosa6533	-1--01--1001--0---1-0000101
Eragrostisbiflora6705	-1--01--1001--0---1-0000101
Eragrostissuperba7470	-1--01--1001--0---1-0000101
Eragrostiscapensis6551	-1--01--1001--0---1-0000101
Eragrostiscapensis6532	-1--01--1001--0---1-0000101
Pogonarthriasquarrosa7483	-1--01--1001--0---1-0000101
Pogonarthriasquarrosa7504	-1--01--1001--0---1-0000101
Catalepisgracilis6962	-1--01--1001--0---1-0000101
Catalepisgracilis6653	-1--01--1001--0---1-0000101
Cladoraphiscyperoides5704	-00101--1001--0---001000101
Cladoraphisspinosa6335	-00101--1001--0---001001101
Cladoraphiscyperoides4894	-00100111001--0---001000101
Fingerhuthiaafricana7507	-1--01--1001--0---1-0000101
Entoplocamiaaristulata9353	-1--00001001--0---1-0000101
Enneapogonscoparius18	-01001--1001--0---1-0000111
Enneapogoncenchroides7506	-01001--1001--0---1-0100101
Enneapogonscoparius7487	-01001--1001--0---1-0000101
Schmidtiaapappophoroides6334	-01001--1001--0---1-0000101
Schmidtiaalihariensis7490	-01001--0001--0---1-0000101
Schmidtiaalihariensis7486	-01001--0001--0---1-0000101
Sporobolusafricanus7476	-1--01--10000000101-0000101
Sporobolusconsimilis9354	-1--01--11-----1-0000101
Sporobolusfimbriatus9328	-1--01--1000010---1-0000101

**APPENDIX L.** Table of correspondence between the indels and the regions they represent (Appendix K) generated by GapCoder for the combined data set.

<b>Indel Character</b>	<b>Sequence Region</b>	<b>Indel Character</b>	<b>Sequence Region</b>	<b>Indel Character</b>	<b>Sequence Region</b>
1832	9-9	1877	223-223	1922	591-591
1833	15-15	1878	223-224	1923	591-592
1834	34-34	1879	234-234	1924	598-598
1835	43-43	1880	239-240	1925	599-601
1836	53-53	1881	239-242	1926	599-599
1837	54-70	1882	239-239	1927	600-601
1838	60-60	1883	375-375	1928	601-601
1839	61-61	1884	390-390	1929	601-602
1840	64-65	1885	422-422	1930	601-603
1841	73-73	1886	425-425	1931	601-605
1842	86-86	1887	438-439	1932	602-603
1843	90-90	1888	438-438	1933	602-605
1844	92-92	1889	439-439	1934	602-606
1845	94-94	1890	443-455	1935	603-603
1846	102-106	1891	443-454	1936	605-607
1847	102-105	1892	443-456	1937	607-608
1848	103-106	1893	468-468	1938	608-608
1849	103-105	1894	483-486	1939	612-612
1850	104-106	1895	485-486	1940	614-614
1851	107-107	1896	490-490	1941	614-615
1852	112-112	1897	490-491	1942	618-619
1853	115-118	1898	492-492	1943	618-624
1854	116-118	1899	496-496	1944	619-619
1855	116-120	1900	497-497	1945	621-621
1856	116-117	1901	505-505	1946	622-623
1857	124-125	1902	510-510	1947	622-624
1858	125-125	1903	514-514	1948	623-623
1859	131-131	1904	515-515	1949	623-624
1860	138-138	1905	518-518	1950	623-625
1861	142-146	1906	524-524	1951	624-624
1862	147-147	1907	527-530	1952	624-625
1863	149-149	1908	527-527	1953	633-634
1864	152-152	1909	528-531	1954	633-635
1865	161-161	1910	528-530	1955	634-635
1866	169-172	1911	538-538	1956	634-636
1867	180-182	1912	543-543	1957	635-635
1868	181-182	1913	555-555	1958	641-641
1869	187-187	1914	562-562	1959	642-642
1870	192-192	1915	564-565	1960	646-646
1871	193-193	1916	565-565	1961	650-650
1872	195-195	1917	573-573	1962	652-660
1873	196-196	1918	581-582	1963	655-660
1874	197-197	1919	583-586	1964	671-671
1875	213-214	1920	589-591	1965	671-672
1876	213-215	1921	590-591	1966	677-677



<b>Indel Character</b>	<b>Sequence Region</b>	<b>Indel Character</b>	<b>Sequence Region</b>	<b>Indel Character</b>	<b>Sequence Region</b>
1967	681-681	2012	951-951	2057	1399-1402
1968	682-682	2013	954-957	2058	1401-1401
1969	682-683	2014	958-958	2059	1418-1421
1970	683-683	2015	987-991	2060	1435-1439
1971	684-684	2016	1009-1009	2061	1498-1500
1972	686-686	2017	1021-1026	2062	1509-1513
1973	689-689	2018	1021-1021	2063	1510-1513
1974	693-694	2019	1023-1026	2064	1518-1541
1975	694-694	2020	1025-1026	2065	1529-1530
1976	694-695	2021	1041-1045	2066	1538-1538
1977	695-695	2022	1041-1044	2067	1553-1554
1978	709-709	2023	1041-1043	2068	1558-1569
1979	719-719	2024	1041-1042	2069	1593-1617
1980	726-726	2025	1053-1053	2070	1594-1617
1981	727-727	2026	1093-1093	2071	1595-1617
1982	730-730	2027	1099-1099	2072	1603-1617
1983	749-749	2028	1101-1102	2073	1627-1637
1984	752-752	2029	1102-1102	2074	1627-1631
1985	771-776	2030	1111-1116	2075	1628-1637
1986	772-776	2031	1139-1190	2076	1636-1719
1987	773-776	2032	1167-1167	2077	1645-1666
1988	782-790	2033	1172-1172	2078	1647-1648
1989	783-786	2034	1178-1182	2079	1652-1660
1990	784-786	2035	1179-1182	2080	1689-1696
1991	784-789	2036	1187-1188	2081	1702-1716
1992	785-786	2037	1188-1188	2082	1705-1710
1993	793-797	2038	1193-1193	2083	1706-1710
1994	801-801	2039	1199-1203	2084	1707-1710
1995	802-802	2040	1212-1212	2085	1708-1710
1996	811-813	2041	1268-1268	2086	1708-1711
1997	822-827	2042	1352-1352	2087	1709-1710
1998	822-822	2043	1354-1391	2088	1709-1709
1999	870-871	2044	1355-1356	2089	1710-1710
2000	872-985	2045	1362-1366	2090	1738-1753
2001	872-873	2046	1362-1375	2091	1738-1752
2002	878-878	2047	1370-1370	2092	1745-1755
2003	899-903	2048	1377-1379	2093	1760-1760
2004	911-932	2049	1377-1377	2094	1764-1764
2005	912-930	2050	1381-1388	2095	1765-1769
2006	912-916	2051	1382-1388	2096	1778-1784
2007	912-928	2052	1384-1388	2097	1810-1810
2008	919-920	2053	1392-1476	2098	1821-1824
2009	923-928	2054	1395-1395		
2010	939-999	2055	1398-1402		
2011	947-947	2056	1399-1403		

**U.V.S. BIBLIOTEK**